



BRIEF REPORT

Identification and molecular characterization of Orf virus infection in occupationally exposed women in South America



Andrea Peralta^{a,*}, Carlos Flores-Olivares^{b,1}, Andrea Verna^{c,d},
Erika González-Altamiranda^d, Ernesto Odriozola^d, Carolina Madariaga^e,
Anselmo Odeón^d, Guido A. König^a, Germán Cantón^d

^a Instituto de Agrobiotecnología y Biología Molecular (IABIMO), Instituto Nacional de Tecnología Agropecuaria (INTA), Consejo Nacional de Investigaciones Científicas y Tecnológicas (CONICET), Argentina

^b Facultad de Ciencias, Universidad Mayor, Chile

^c Consejo Nacional de Investigaciones Científicas y Tecnológicas (CONICET), Argentina

^d Grupo de Sanidad Animal, INTA EEA Balcarce, Argentina

^e Universidad Santo Tomás, Chile

Received 30 December 2021; accepted 18 July 2022

Available online 30 September 2022

KEYWORDS

Contagious Ecthyma;
Dermatitis;
Human Orf;
Zoonosis;
Molecular
characterization

Abstract Contagious Ecthyma (CE) is a severe exanthematous dermatitis caused by the Orf virus (ORFV) that mainly affects domestic small ruminants such as sheep and goats. It is a worldwide-distributed occupational zoonosis, particularly infecting those in close contact with animals or animal products such as shepherds, farmers and veterinarians, among others. In the present work, we report the first human CE case confirmed in Argentina. A phylogenetic analysis based on four gene sequences of the isolated strain responsible for the disease showed that this isolate grouped with other ORFV sequences that caused reported CE cases in sheep from the same Argentine province. We also sequenced a sample from a Chilean human case reported in 2017, whose phylogenetic analysis showed that it groups together with other Argentine isolates from locations close to the border with Chile.

© 2022 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

* Corresponding author.

E-mail address: peralta.andrea@inta.gob.ar (A. Peralta).

¹ Current affiliation: Facultad de Ciencias Agropecuarias, Universidad del Alba, Chile.

PALABRAS CLAVE

Ectima contagioso;
Dermatitis;
Orf en humanos;
Zoonosis;
Caracterización molecular

Identificación y caracterización molecular de virus Orf en mujeres de Sudamérica expuestas laboralmente

Resumen El ectima contagioso (EC) es una dermatitis exantemática grave causada por el virus Orf (ORFV), que afecta mayormente a pequeños rumiantes domésticos, como ovinos y caprinos. Es una zoonosis ocupacional con distribución mundial, infecta a humanos en estrecho contacto con animales o sus productos, como granjeros, esquiladores y veterinarios, entre otros. En este trabajo se informa el primer caso humano de EC confirmado en Argentina. Un análisis filogenético basado en cuatro genes de la cepa responsable de este caso mostró que el aislamiento agrupa con otras secuencias de ORFV que causaron casos en ovinos en la misma provincia argentina. También se secuenció una muestra del caso de ectima humano reportado en Chile en 2017 y el análisis filogenético mostró que dicho aislamiento forma un grupo con otros aislamientos argentinos de localidades cercanas a la frontera con Chile.

© 2022 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Contagious Ecthyma (CE) is a severe exanthematous dermatitis affecting small domestic ruminants such as sheep and goats, although it can also be detected in wild animals⁶. CE is caused by the Orf virus (ORFV), member of the *Poxviridae* Family, genus *Parapoxvirus* (PPV)³. ORFV has a linear double-stranded DNA genome, of approximately 135 kbp in size and exhibits an unusual high GC content (~64%). The central region of the genome contains 88 genes which are present in the subfamily *Chordopoxvirinae* and mostly occur in a common order and orientation. The terminal regions of the genome are variable and there are genes associated with virulence, pathogenesis, tropism and immune response modulators (*orf001* to *orf008* and *orf112* to *orf134*).

ORFV is an epitheliotropic virus that causes lesions on the lips, nostrils, oral mucosa and udders of sheep and goats. ORFV can remain infective for many weeks, especially in scabs, thus enabling widespread infection in the flock¹⁰.

CE is also a zoonotic disease that can spread to humans after contact with infected animals or contaminated fomites². Human skin lesions due to ORFV are usually self-limiting and resolve spontaneously within 4–6 weeks, except in immunocompromised individuals⁵.

Human ORFV infections are not usually reported in South American clinical practice. Considering the widespread distribution of CE in flocks all over the region^{1,11,12}, ORFV

infection in humans may be underdiagnosed, which may lead to overtreatment or unnecessary medical procedures¹⁵.

The first human ORFV case diagnosed in Argentina was detected in a 52-year-old female sheep keeper from a farm in Villarino Department (south of Buenos Aires province). The flock of the farm had suffered a CE outbreak (lamb skin samples were identified as sMbu15) that affected more than 90% of young lambs (<3 months old) (Fig. 1A) weeks before the disease was confirmed in the keeper. The patient showed erythematous papules on the back of both hands (Fig. 1B) that evolved into inflammatory nodules with a diameter of approximately 2 cm (Fig. 1C). Initially, she attended a primary health care center, where the signs were attributed to a complication of her diabetic condition. Two weeks later, the lesions progressed to the left forearm, with erythematous, edematous linear appearance and evident pain, compatible with lymphangitis. Once ORFV infection was confirmed in lambs from the flock she cared for, she was advised to return to the health center where a skin sample was biopsied (hMbu15) and sent for analysis. Informed consent was obtained from the individual that participated in the study.

In Chile, the first human ORFV case was reported by Flores et al.⁴ Briefly, a young veterinary student received a goring wound in her right hand when she was performing a clinical examination of apparently healthy goats from a

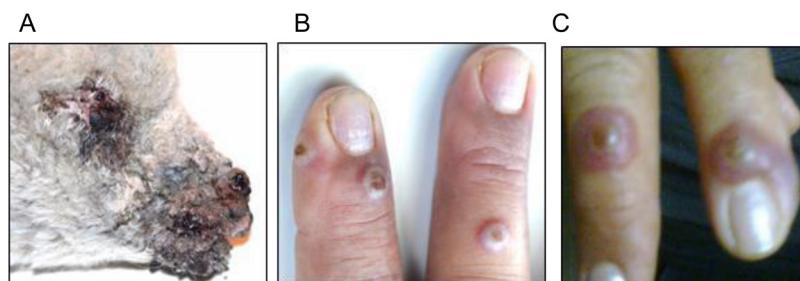


Figure 1 Lesions caused by ORFV infection in an Argentine rural worker. (A) Photograph of a dramatic ORFV infection in a 3-month-old affected lamb. (B) Three erythematous papules are observed on the back of the right hand. (C) Few days later, papules evolved into inflammatory nodules measuring 2 cm in diameter.

flock. Two weeks later, she noted the appearance of typical EC lesions and consulted a dermatologist, who biopsied the skin samples (hChi17) for PCR diagnosis. No further analysis was performed.

In this work, we analyzed hMbu15 and hChi17 and compared them with other ruminant ORFV strains by using different genes: *orf011*, *orf020*, *orf109*, and *orf127*. We present the first human CE case confirmed in Argentina along with a phylogenetic analysis of its etiological agent.

First, we confirmed an ORFV infection in a human scab sample (hMbu15) by using an *orf045* internal region gene amplification by PCR. This diagnostic PCR proved to have 100% efficiency in detected viral genomes in different samples⁸. Subsequently, DNA samples hChi17 and hMbu15 were used as a template for PCR reactions to amplify an *orf011* gene fragment and the complete open reading frame of *orf020*, *orf109* and *orf127* genes, as previously described¹². The *orf011* gene encodes a highly immunogenic major envelope protein B2L. The internal region of the *orf011* gene (nt 388 to 981) has been mostly used for the phylogenetic analysis because it is the virus sequence most deposited in databases, thus allowing strains from all over the world to be compared. Furthermore, we analyzed the *orf020* and *orf127* genes, which codify virulence factors VIR and VIL10 respectively, and *orf109* because it encodes an envelope mature protein (EEV), one of the most variable genes within the PPV genus. The sequences were submitted to the GenBank databases under the following accession numbers: *orf011* internal region: MH161454, MH161455; *orf020*: MH161456, MH161457; *orf109*: MH161458, MH161459 and *orf127*: MH161460, MH161461.

The sequence of the *orf011* internal region gene⁷ is commonly used for the OFRV phylogenetic analysis⁹. The identity matrix showed that hMbu15 was 100% identical to sMbu15, while hChi17 showed 99.8% and 100% identity with other ORFV strains isolated from goats of Rio Negro (gCom14) and San Luis (gCnc15) provinces in Argentina, respectively.

As expected, a maximum likelihood (ML) phylogenetic tree analysis resulted in a group that contained hChi17 together with the above mentioned ORFV isolates from Argentina (gCom14 and gCnc15) and Brazil (NE2, NE1, Bahia and SJ1) (Fig. 2A). This group contains exclusively goat ORFV strains, with a significant bootstrap support (Fig. 2A). On the other hand, hMbu15 constitutes a heterogeneous group together with other ORFV isolates obtained from sheep (sMbu15, sBal15, sChu14, sPil14, sPi13) and goats (gNeu14) from Argentina as well as isolates from goats in Brazil (MT05, 578/08, 561/11 and 27/12), Uruguay (UY19/10) and North America (orf-mu, orf-ta and orf-sh) (Fig. 2A). This particular tree group exhibits many branches with low or intermediate bootstrap values, probably due to a high nucleotide sequence identity among all included samples in the phylogenetic analysis^{13,14}.

To improve this analysis, we extended the study to other ORFV genes: *orf020* and *orf127*, which were described as virulence factors, and *orf109*, which codifies an extracellular mature virion protein that is orthologous to vaccinia virus A33R protein³. In accordance with the finding using the *orf011* gene, hMbu15 and sMbu15 were 100% identical in

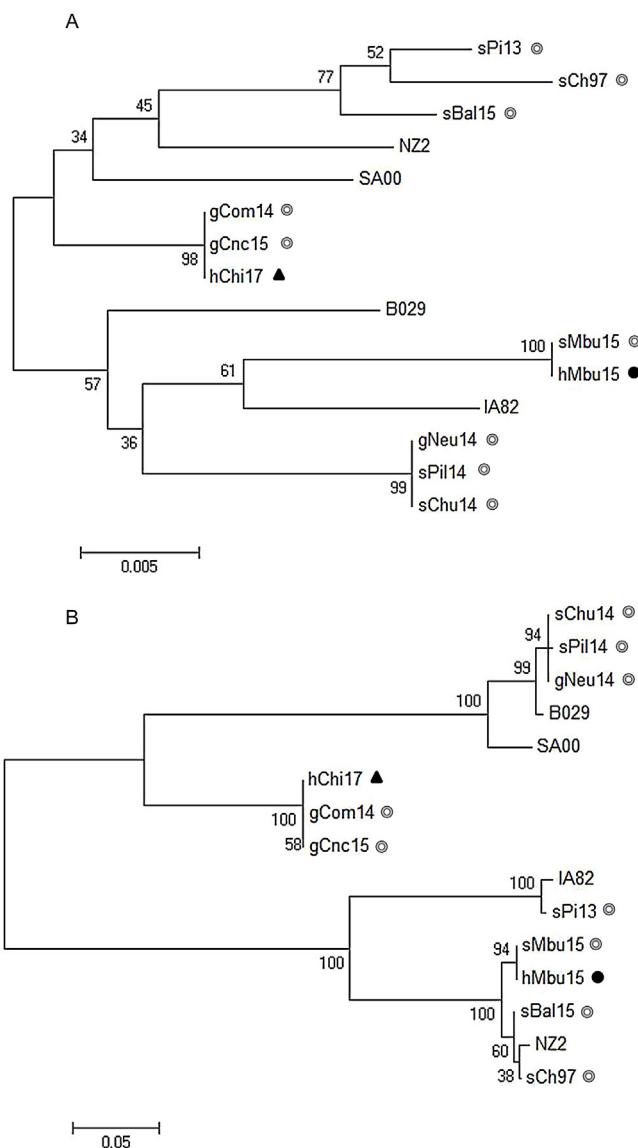


Figure 2 Phylogenetic analysis of South American human ORFV strains. The phylogenetic relationships were constructed by the maximum likelihood algorithm using the MEGA 6.0 software. All positions with less than 50% site coverage were eliminated. The tree with the highest log likelihood is shown. Initial trees for the heuristic search were obtained automatically by applying Neighbor-Join and BioNJ algorithms to a matrix of pairwise distances. Numbers at nodes represent % of 500 bootstrap replicates. White circles indicate Argentinian sheep and goat samples, the dark circle indicates the Argentinian human sample, while dark triangle, the Chilean human sample. (A) Phylogenetic analysis based on 517 nucleotide sequence positions of the *orf011* internal region; the Tamura-3 parameter was selected as the best-fit evolution model. (B) Phylogenetic analysis based on 2046 nucleotide sequence positions of the *orf011*, *orf020*, *orf109* and *orf127* concatenated sequences; the Kimura-2 parameter was selected as the best-fit evolution model. Only strains with the four molecular marker sequences available at the Gene Bank were included.

all genes analyzed, which confirms the origin of the human infection. Moreover, the analysis also revealed 100% identity between hChi17 and the Argentine gCom14 and gCnc15 in *orf109* and *orf127*. However, the *orf020* gene exhibited some variability between hChi17 with gCom14 and gCnc15 (0.2 and 0.5% respectively). Subsequently, a ML phylogenetic tree was constructed with concatenated *orf011*, *orf020*, *orf109* and *orf127* sequences to simplify the visualization of the global analysis (Fig. 2B).

As expected, hMbu15 and sMbu15 were identical and formed a branch with other ORFV strains from Buenos Aires province (sBal15 and sCh97), with 100% bootstrap support, and separated from other strains detected in Argentina (Fig. 2B). This finding could indicate the heterogeneity of ORFV strains from different regions in Argentina.

The phylogenetic analysis with four molecular markers along the genome also confirmed no close phylogenetic relationship between hMbu15 and hChi17 samples. Surprisingly, hChi17 was closely related to other goat ORFV strains from Río Negro (gCom14) and San Luis (gCnc15) provinces in Argentina. A phylogenetic analysis obtained from the four concatenated molecular markers showed that gCom14, gCnc15 and hChi17 form a group with high confidence support (Fig. 2B), which indicates a close relationship between the three isolates. Moreover, they were 100% identical in two of the four markers studied (*orf109* and *orf127* genes), which may indicate that these strains share a common origin. There is a natural border between Argentina and Chile (The Andes range of mountains), but apparently this barrier does not prevent the exchange of animals or contaminated fomites that move the virus from one region to another. Nevertheless, studies including more ORFV strains from Chile and Argentina are necessary to confirm this hypothesis and further studies could help to establish new policies on the ruminant movements between both countries.

The first human CE case from Argentina was initially misdiagnosed by medical practitioners. This is a common finding in the region since healthcare staff are usually unaware of the occurrence of this zoonosis. It is necessary to warn physicians about the probability of ORFV infection in humans, especially when they are in contact with susceptible animals. Zoonotic diseases, such as CE, could represent a threat to human health as was widely demonstrated by the COVID-19 pandemic.

This study presents the first molecular comparison of human ORFV strains from Argentina and Chile. The obtained data increase the knowledge of the ORFV strains circulating in the region at a molecular level and show the importance of performing an update about human CE in the region, both for workers with exposure risk and healthcare providers.

Conflict of interest

The authors have no conflict of interest to declare.

Authors' contributions

AP, CFO, and GC contributed to the conception and design of the study. AP performed the PCRs for the four

molecular markers and the sequence analysis. GAK performed the phylogenetic analysis. AV, EGA and AO performed diagnostic PCR. CFO, EO and CM characterized the outbreaks in the field, identified human cases and took epidemiological data.

All authors contributed to manuscript revision, read and approved the submitted version.

Acknowledgment

Financial support was provided by PNBI01131032 and PNSA1115055, Instituto Nacional de Tecnología Agropecuaria (INTA), Argentina.

References

1. Abrahão JS, Borges IA, Mazur C, Lobato ZIP, Ferreira PCP, Bonjardim JA, Trindade GS, Kroon EG. Looking back: a genetic retrospective study of Brazilian Orf viruses isolates. *Vet Rec.* 2012;171:476.
2. Bayindir Y, Bayraktar M, Karadag N, Ozcan H, Kayabas U, Otlu B, Durmaz R, Doganay M. Investigation and analysis of a human Orf outbreak among people living on the same farm. *New Microbiol.* 2011;34:37–43.
3. Damon I. Poxviridae and their replication. In: *Fields virology*. New York: Raven Press Ltd; 2007. p. 2079–81.
4. Flores C, Gonzalez E, Verna A, Peralta A, Madariaga C, Odeón A, Cantón G. Virus Orf en humanos, confirmación molecular de un caso clínico en Chile. *Rev Chilena Infectol.* 2017;34:607–9.
5. Geerinck K, Lukito G, Snoeck R, De Vos R, De Clercq E, Vanrenterghem Y, Degrue H, Maes B. A case of human orf in an immunocompromised patient treated successfully with cidofovir cream. *J Med Virol.* 2001;64:543–9.
6. Guo J, Rasmussen J, Wünschmann A, de La Concha-Bermejillo A. Genetic characterization of Orf viruses isolated from various ruminant species of a zoo. *Vet Microbiol.* 2004;99:81–92.
7. Inoshima Y, Morroka A, Sentsui H. Detection and diagnosis of parapoxvirus by the polymerase chain reaction. *J Virol Methods.* 2000;84:201–8.
8. Kottaridi C, Nomikou K, Lelli R, Markoulatos P, Mangana O. Laboratory diagnosis of contagious ecthyma: comparison of different PCR protocols with virus isolation in cell culture. *J Virol Met.* 2006;134:119–24.
9. Lojkic I, Cac Z, Beck A, Bedekovic T, Cveticic Z, Sostaric B. Phylogenetic analysis of Croatian orf viruses isolated from sheep and goats. *Virol J.* 2010;7:314.
10. Nandi S, De UK, Chowdhury S. Current status of contagious ecthyma or orf disease in goat and sheep. A global perspective. *Small Rumin Res.* 2011;96:73–82.
11. Olivero N, Reolon E, Arbiza J, Berois M. Genetic diversity of Orf virus from sheep in Uruguay. *Arch Virol.* 2018;163:1285–91.
12. Peralta A, Robles C, Martinez A, Alvarez L, Valera A, Calamante G, König GA. Identification and molecular characterization of Orf virus in Argentina. *Virus Genes.* 2015;50:381–8.
13. Peralta A, Robles C, Micheloud J, Rossanigo C, Martinez A, Carosio A, Konig G. Phylogenetic analysis of ORF viruses from five Contagious Ecthyma outbreaks in Argentinian goats. *Front Vet Sci.* 2018;5:134.
14. Schmidt C, Cargnelutti JF, Brum MCS, Traesel CK, Weiblen R, Flores EF. Partial sequence analysis of B2L gene of Brazilian Orf viruses from sheep and goats. *Vet Microbiol.* 2013;162:245–53.
15. Steinhart B. Orf in humans: dramatic but benign. *CJEM.* 2005;7:417–9.