

IMPACT OF GENETIC ANCESTRY ON THE DISTRIBUTION OF INTERFERON- λ 4 RS12979860 POLYMORPHISM IN A GLOBAL POPULATION OF BUENOS AIRES, ARGENTINA



IMPACTO DE LA ANCESTRÍA GENÉTICA EN LA DISTRIBUCIÓN DEL POLIMORFISMO DE INTERFERÓN- λ 4 RS12979860 EN UNA POBLACIÓN GLOBAL DE BUENOS AIRES, ARGENTINA

Mansilla F.C.¹, Avena S.A.^{2,3,4}, Dejean C.B.^{2,3}, Turco C.S.¹, Capozzo A.V.^{1,4}

¹Instituto de Virología e Innovaciones Tecnológicas (IVIT), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) – Instituto Nacional de Tecnología Agropecuaria (INTA), Hurlingham, Buenos Aires, Argentina.

²Centro de Ciencias Naturales, Ambientales y Antropológicas (CCNAA), Universidad Maimónides, Buenos Aires, Argentina.

³Sección Antropología Biológica, Instituto de Ciencias Antropológicas (ICA), Facultad de Filosofía y Letras, Universidad de Buenos Aires, Buenos Aires, Argentina.

⁴Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Argentina.

Corresponding author:

Mansilla F.C.
mansilla.florencia@inta.gob.ar

 ORCID 0000-0002-8325-0579

ABSTRACT

Human interferon- λ 4 is a cytokine involved in early stages of antiviral responses. Strikingly, some allelic variants with diminished antiviral activity reduce the susceptibility to viral infections, thus they would have suffered a positive selection pressure throughout the evolutionary history of the genus *Homo*. An intronic variant within the IFN λ 4 locus (rs12979860, T>C) emerged as one of the main gene determinants of the response to HCV and other viruses. The rs12979860-C allele has a differential frequency in African, European and Native American populations, though South American data are scarce. Here we characterize for the first time the distribution of rs12979860 genotypes in a sample of the global population of Buenos Aires, Argentina, assessing its association with European, Native American and African parental components. The rs12979860 genotypes were determined by PCR-RFLP in DNA samples from donors of a blood banks of Buenos Aires (n=96), whose genetic individual ancestry (European, African or Native American) had been previously determined using molecular markers. The distribution of rs12979860-CC, CT and TT was 29.17%, 50.0% and 20.83%, respectively. A significant increase in the frequency of CC among donors with a strong European contribution and a greater impact of the Native American component among donors carrying the T allele were observed. Native American and European components were associated to the rs12979860 distribution in a sample of the global population of Buenos Aires, while no differences were directly attributable to the African ancestry. Considering interferon's key role in antiviral responses, our results may contribute to both bioanthropological and immunogenetic studies associated with infectious diseases.

Key words: ancestry, Buenos Aires, IFN λ 4 polymorphism, rs12979860 distribution.

RESUMEN

El interferón- λ 4 humano es una citoquina involucrada en la respuesta antiviral. Algunas variantes alélicas con menor actividad antiviral, paradójicamente, reducen la susceptibilidad a infecciones virales, por lo que habrían sufrido una presión de selección positiva en la historia evolutiva del género *Homo*. Una variante dentro del locus de IFN λ 4 (rs12979860, T>C), con distribución diferencial en poblaciones africanas, europeas y nativas americanas, surgió como uno de los principales determinantes genéticos de la respuesta al HCV y otros virus. Aquí caracterizamos por primera vez la distribución de los genotipos de rs12979860 en una muestra de la población cosmopolita de Buenos Aires, Argentina, evaluando el impacto de su ancestría. Se determinaron diferentes genotipos de rs12979860 por PCR-RFLP en muestras de ADN de donantes de bancos de sangre de Buenos Aires (n=96), cuya ancestría individual había sido previamente determinada mediante diferentes marcadores moleculares. La distribución global de rs12979860-CC, CT y TT fue 29,17%; 50,0% y 20,83%, respectivamente. Se observó un aumento significativo de la frecuencia del genotipo CC entre individuos con fuerte aporte europeo y un mayor impacto del componente nativo-americano entre portadores del alelo T. Los componentes nativo-americano y europeo se asociaron a la distribución rs12979860 en una muestra poblacional global de Buenos Aires, mientras que no se vieron diferencias directamente asociadas a la ancestría africana. Considerando el papel clave del interferón en la respuesta antiviral, nuestros resultados pueden contribuir a estudios con un enfoque bioantropológico así como a estudios inmunogenéticos asociados a enfermedades infecciosas.

Palabras clave: ancestría, Buenos Aires, polimorfismo en IFN λ 4, distribución de rs12979860.

Cite this article as:

Mansilla F.C., Avena S.A., Dejean C.B., Turco C.S., Capozzo A.V. 2022. IMPACT OF GENETIC ANCESTRY ON THE DISTRIBUTION OF INTERFERON- λ 4 RS12979860 POLYMORPHISM IN A GLOBAL POPULATION OF BUENOS AIRES, ARGENTINA. BAG: Journal of Basic and Applied Genetics XXXIII (2): 19-25.

Received: 03/04/2022

Revised version received: 06/30/2022

Accepted: 08/08/2022

General Editor: Elsa Camadro

DOI: 10.35407/bag.2022.33.02.02

ISSN online version: 1852-6233

INTRODUCTION

Lambda interferons (IFN λ) are cytokines rapidly produced by most vertebrates during the innate immune response, constituting the first line of defense against viral infections (Lazear *et al.*, 2015). IFN λ 1, 2 and 3 were identified in 2003 (Kotenko *et al.*, 2003; Sheppard *et al.*, 2003) and in 2013 a functional form of IFN λ 4 was firstly characterized (Prokunina-Olsson *et al.*, 2013). The IFN λ 4 locus (19q13.2) is highly polymorphic (Fang *et al.*, 2020) and it was reported that some allelic variants can modulate the susceptibility, progression and response to treatments against different viral infections (Chatterjee, 2010; Bravo *et al.*, 2014; Angulo *et al.*, 2015; Ispiroglu *et al.*, 2017; da Silva Cezar *et al.*, 2020). Interestingly, the most favorable alleles in this regard correspond to mutations that are in strong linkage disequilibrium and restrict the expression, stability or antiviral activity of IFN λ 4 (Booth and George, 2013; O'Brien *et al.*, 2014; Prokunina-Olsson, 2019).

Throughout the evolutionary history of the genus *Homo* these mutations have suffered a positive selection pressure resulting in a differential global distribution which is correlated to the ancestry of different human populations and may affect the immune response to different pathogens (Key *et al.*, 2014; Bamford *et al.*, 2018). An intronic variant that reduces the antiviral activity of IFN λ 4 (rs12979860, T>C) was characterized as the main gene determinant of the response against Hepatitis C Virus (HCV). The rs12979860-T allele is associated with lower sustained virologic response (SVR) rates and a lower percentage of treatment success (Ge *et al.*, 2009). On the other hand, the CC genotype was strongly associated with spontaneous resolution and lower susceptibility to HCV infection (Thomas *et al.*, 2009; Pedernana *et al.*, 2012; Indolfi *et al.*, 2014a; Fan *et al.*, 2016). Moreover, genotyping of rs12979860 is recommended to predict the patient's response to different antiviral treatments (Sharafi *et al.*, 2012; Ramamurthy *et al.*, 2018). Different correlations between rs12979860 and clinical phenotypes associated with other viral infections have also been reported, conditioning the susceptibility, evolution and/or response to treatment against Hepatitis B and D (Ispiroglu *et al.*, 2017), Dengue (da Silva Cezar *et al.*, 2020), HIV (Chatterjee, 2010; Zaidane *et al.*, 2018), CMV (Bravo *et al.*, 2014; Chmelova *et al.*, 2019) and coronaviruses (Hamming *et al.*, 2013).

The rs12979860-C allele has a global frequency of 0.23–0.55 in African populations; 0.53–0.80 for Europeans and 0.72–1.00 for Asians, with higher frequencies in eastern Asia. Data about the distribution of these variants in South American populations are scarce and tend to be biased due to the small sample size and the genetic admixture of the populations assessed. The Argentinean population's ancestry is the result of

a deep miscegenation, product of different migratory waves during the last centuries, which means that the European, Native American and African components (frequently underestimated) are present at different degrees in the gene pool of different cosmopolitan populations of the country (Avena *et al.*, 2012). In this regard, the immunogenetic profiling of IFN λ 4-rs12979860, and the association with its ancestry, may be a potential tool in both anthropological and biomedical studies associated with infectious diseases. The objective of this study was to determine the distribution of the allelic variants of rs12979860 in a cosmopolitan population of Buenos Aires, Argentina, whose ancestry had been previously determined by assessing a set of 106 biallelic SNPs (Ancestry Informative Markers) widely spaced and balanced throughout the genome, that can discriminate Native American, African and European ancestry (Avena *et al.*, 2012).

MATERIALS AND METHODS

Study Design

This study comprised DNA samples from unrelated donors from both public and private hospitals blood banks in Buenos Aires, Argentina (n=96). Informed consent was obtained from all individual participants included in the study. Most of them (89/96) also agreed to provide information about the region/country of birth of all their grandparents, which was included in the data analysis. The study was approved by the Ethics Committee of the Hospital Italiano of Buenos Aires and was performed in accordance with the ethical standards adopted in the Declaration of Helsinki.

rs12979860 genotyping

Different genotypes of rs12979860 were determined by PCR-RFLP, as it was previously described (Sharafi *et al.*, 2012). A 241 bp fragment was amplified by endpoint PCR (Taq Pegasus®, Productos Bio-Lógicos, Bs. As., Argentina) following a standard cycle (5 min at 94° C; 35 cycles of 20 s at 94° C, 20 s at 59° C and 20 s at 72° C; and 5 min at 72° C) and then digested with Bsh12361 restriction enzyme (Thermo Fisher, DE, USA; 1U/reaction) for 1 h at 37° C. The primers used were 5'GCGGAAGGAGCAGTTGCGCT3' (Fw) and 5'TCTCTCCCCAAGTCAGGCAACC3' (Rv) and the resulting fragments (rs12979860-CC = 196 + 45 bp; rs12979860-CT = 241 + 196 + 45 bp; rs12979860-TT = 241 bp) were revealed by agarose gel electrophoresis (3%) stained with GelRed (Biotium, CA, USA).

Statistical analysis

The allelic frequencies were determined, and Hardy-Weinberg equilibrium was assessed using the chi-square test (Microsoft Excel GenAIEx 6.5, Peakall and Smouse, 2012) to compare the genotype distribution. Differences associated to European, Native American or African component were determined using T test (GraphPad Prism 9). In all statistical analysis a $p < 0.05$ was considered as statistically significant and $\alpha = 0.05$ was set as the risk level.

RESULTS AND DISCUSSION

The average individual ancestry was estimated as 69.4% European, 26.3% Native American and 4.3% African. Frequencies lower than 0.02 were not included in the data analysis since they may be associated to technical artifacts. The European component was present in every tested sample, with individual frequencies ranging from 0.02 to 1. The Native American component was also detected but to a lesser extent, in 79% of the samples (frequencies 0.02-0.8). Finally, the African ancestry was detected in 41% of the samples, with a frequency range from 0.02 to 0.23 (Figure 1, modified from Avena *et al.*, 2012). This evidences the multiplicity of origins of Buenos Aires' population, resulting of the miscegenation between Native Americans, enslaved Africans who came mainly from West Africa and Mozambique until the first half of the 19th century (Fejerman *et al.*, 2005) and European immigrants, mainly from Italy and Spain, who arrived in the country between 1870 and 1960 (Avena *et al.*, 2006; Muzzio *et al.*, 2018). These results are in line with previously published data (Avena *et al.*, 2006), further challenging the European self-perception as Argentina's identity.

Several studies have reported the distribution of the rs12979860 genotypes in different populations, mainly assessing its correlation with the susceptibility to different viral infections and response to antiviral

treatments (Wu *et al.*, 2012; Porto *et al.*, 2015; Taheri *et al.*, 2015; Echeverría *et al.*, 2018). The correlation of this distribution and the local ancestry of these populations as well as its implications have also been assessed (Indolfi *et al.*, 2014b; Rizzo *et al.*, 2016), though this is the first report in an Argentinean global population. The overall distribution of rs12979860-CC, CT and TT was 29.17%, 50.0% and 20.83%, respectively. Hardy-Weinberg equation was used to calculate the genetic variation of this population at equilibrium. Significant differences were not detected (chi-square test: 0.00469; $p = 0.99766$), thus suggesting that the impact of possible microevolutionary mechanisms and population structure is not significant. The allelic frequencies for C and T were 54.17% and 45.83%, respectively. These results differ from data reported in HCV chronically infected patients of a public center in Buenos Aires, with an allelic frequency of C=0.6 and 45.0% of heterozygosity (Machicote *et al.*, 2018). This higher frequency of rs12979860-C is expected as it is known that this allele is favorable in both acute and chronic HCV infection. In this regard, the differences observed between healthy and infected individuals highlight the impact of assessing global populations when studying the distribution of this kind of markers.

A significant increase in the frequency of CC genotype was observed among donors with a strong European contribution (Figure 2a, $p < 0.05$). Our results also suggest a greater impact of the Native American component among donors carrying the T allele (both CT and TT genotypes), although differences were marginally significant (Figure 2b). No differences in the rs12979860 distribution were directly attributable to the African component (Figure 2c, $p > 0.05$), represented at low levels in our sample.

Based on previously reported data on the composition and immigration patterns of the admixed population of Buenos Aires (Avena *et al.*, 2012), we defined our parental population including sub-Saharan Africans (involved in slavery trafficking) and Europeans from Italy and Spain (Avena *et al.*, 2006). To minimize bias, we only considered

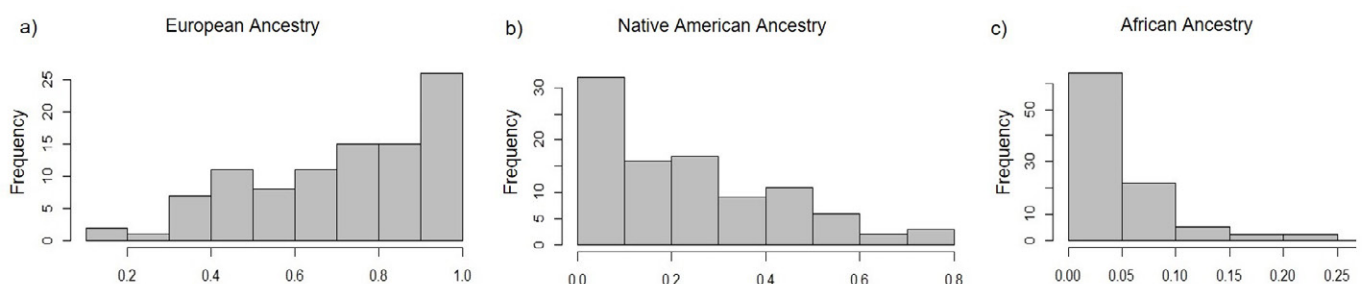


Figure 1. Frequency distribution of the individual European (a), Native American (b) and African ancestry (c) among healthy donors from Buenos Aires, Argentina, enrolled in this study (n=96). Modified from Avena *et al.* (2012).

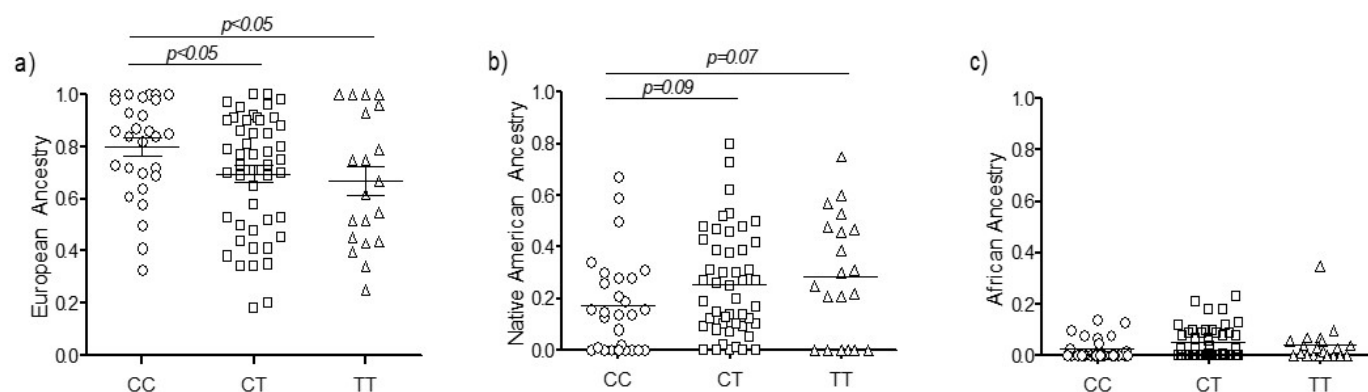


Figure 2. Distribution of European (a), Native American (b) and African ancestry (c) among individuals carrying rs12979860-CC, CT and TT genotypes. $p < 0.05$ were considered as statistically significant.

reported data on the rs12979860 distribution (Table S1) from non-cosmopolitan populations with a sample size greater than 50. Ethiopian Jews and Sephardic Jews from Rome, Italy, were also excluded, as these groups tend to be endogamous and have a different origin, which may introduce certain bias to our analysis. The mean frequency of the rs12979860-C allele for this parental population is 0.654 for Europeans, 0.298 for Africans and 0.518 for Native Americans (table S1). However, data available regarding the Native American component are scarce and are often based either on cosmopolitan admixed populations or studies with very small sample sizes and variable results. Despite the lack of a robust sample to perform comparisons, our results suggest that populations with greater autochthonous ancestry tend to exhibit higher frequencies of the rs12979860-T allele.

Further studies are needed to fully characterize the distribution of this polymorphism in Latin America, as available data seem to be contradictory. To explain this it is important, regarding cosmopolitan populations, to disclose their composition and their genetic ancestry in order to determine their parental populations' contribution. Latin American cosmopolitan populations are known to be admixed, but the European, Native American and sub-Saharan contributions have marked regional differences. Hence the relevance of studying cases such as the one here described considering the genetic ancestry of the population under study.

The frequency of rs12979860-C in Buenos Aires' individuals was similar to previously reported data for populations from Tuscany ($C=0.603$) in Italy, which are among the lowest compared to other West European populations (Table S1). The reported frequency of this allele in an Iberian population, however, was higher than the one described in our study ($C=0.705$, Table S1).

Although immigrants from both Italy and Spain are the main determinants of the European ancestry of Buenos Aires' population (Avena *et al.*, 2006), it is to note that most of the immigrants in Buenos Aires (and Argentina) were of Italian origin (Avena *et al.* 2006). This may explain, at least partially, the frequencies here described.

In order to further characterize the European contribution to the rs12979860 distribution we considered, when available, the self-reported data about grandparents' origins. Interestingly, a total of 53 individuals declared the nonexistence of grandparents of European origin (8/53:CC, 33/53:CT and 12/53:TT), while only 36 individuals reported at least one grandparent from Italy, Spain/Portugal or other European countries (14/36:CC, 16/36:CT and 6/36:TT). This may be attributed to the fact that the vast majority of immigrants arrived in Buenos Aires before 1950. In our sample, the presence of Iberian ancestry seems to be underrepresented, as genealogical data suggest that the self-reported Italian ancestry was 33.0% higher than Iberian ancestry. Altogether, our results may be explained by the higher presence of Italian ancestry among European descendants in our sample, as well as by the admixture of these individuals with Native Americans and Africans or afro-descendants with a higher rs12979860-T frequency, thus increasing the heterozygosity and the rs12979860-T frequency. However, it is important to consider that, despite being very useful especially in regions with recent immigration patterns (Avena *et al.* 2012), this kind of surveys must be carefully analyzed, since different social and economic aspects may influence the individual self-perceived ancestry, as it was recently reported (Paschetta *et al.* 2021).

Notably, most of the populations that have been included in large-scale immunogenomic studies were

of European origin, and might include certain bias by demographic, social and economic conditions of non-randomly selected individuals (Peng *et al.*, 2021). This may have affected the representativeness of the sample, thus compromising the conclusions of those studies. Therefore, increasing the genetic diversity while considering these structural inequalities is mandatory in order to obtain more reliable results. The PCR-RFLP protocol here applied was previously described and fully validated against PCR-sequencing, with a concordance of 100% in the results obtained for C/T alleles (Sharafi *et al.*, 2012). In this regard, the use of a simple low-cost and high-yielding technique is paramount, since it allows small regional laboratories with limited resources to conduct population genetic studies, thus reducing the sampling bias that may occur in large cosmopolitan cities. This is particularly relevant in regions such as South America, in which the availability of qPCR or sequencing platforms is still limited.

During the last years, there has been a growing interest on the impact of genetic ancestry on the immune response against viral infections (Mersha and Abebe, 2015). The molecular determinants responsible for those associations are being increasingly understood, and interferon pathways and their expression patterns seem to be influenced by genetic ancestry (Miretti and Beck, 2006; Randolph *et al.*, 2021), as suggested by our results.

In the context of the COVID-19 pandemic and considering that IFN λ 4 can elicit an antiviral response against RNA viruses, including some coronaviruses, several studies have assessed whether rs12979860 is involved in SARS-CoV-2 susceptibility and COVID-19 outcome. In this regard, it was reported that the T allele was overexpressed in COVID-19 patients compared to the general healthy population (36.2% vs. 26.4%), thus, this allele was proposed as a possible risk factor for COVID-19 (Saponi-Cortes *et al.*, 2021). This was also supported by Rahimi *et al.* (2021), who demonstrated a positive correlation between the survival rate in COVID-19 patients and the rs12979860-CC genotype, which is also favorable to control other infectious diseases caused by RNA viruses. On the other hand, a higher frequency of the CC genotype among COVID-19 patients was reported in a different study, suggesting that people with the C allele (both CT or CC genotypes) are more susceptible to SARS-CoV-2 infection (Agwa *et al.*, 2021). However, only slight differences between infected and control groups are shown (44.7% vs. 44.0%, respectively) and allelic frequencies are the same for both groups (C=34.0%, T=66.0%). In that study, it was also reported that 52.6% of the TT genotypes were classified as severe disease compared to 45.8% and 34.9% in the TC and CC genotypes, respectively (Agwa *et al.*, 2021), which seem to be in line with the results published by Saponi-Cortes *et al.* (2021) and Rahimi *et*

al. (2021). It is to be noted, also, that the differences shown by Agwa *et al.* (2021) may not be exclusively explained by rs12979860 variants, considering that comorbidities were found in 57.4% of the infected group (and in 18.0% of controls). This highlights the relevance of carrying out a properly designed and unbiased sampling as well as a cautious analysis of the results in order to discern this type of controversies when assessing the differential distribution of these variants in different populations.

CONCLUSIONS

Given its importance and its apparent association with different infectious diseases, there is a growing interest in assessing IFN λ 4 polymorphisms. As a whole, this study describes for the first time the distribution of rs12979860 polymorphism in a healthy sample of the population of Buenos Aires, Argentina, further demonstrating that these frequencies are associated to the composition of the population. This, in addition to being useful in anthropological studies, may contribute to the study of different infectious diseases for which interferon antiviral responses are key.

ACKNOWLEDGMENTS

We thank Dr. Karina Trono for critical reading of the manuscript. This research was funded by the services provided by AC's group through STAN-CONICET. Other support came from PIP CONICET 2111.

BIBLIOGRAPHY

- Agwa S.H.A., Kamel M.M., Elghazaly H., Abd Elsamee A.M., Hafez H., Girgis S.A., Elarab H.E., Ebeid F.S.E., Sayed S.M., Sherif L., Matboli M. (2021) Association between Interferon-Lambda-3 rs12979860, TLL1 rs17047200 and DDR1 rs4618569 Variant Polymorphisms with the Course and Outcome of SARS-CoV-2 Patients. *Genes*, 12(6). <https://doi.org/10.3390/GENES12060830>
- Angulo J., Pino K., Echeverría-Chagas N., Marco C., Martínez-Valdebenito C., Galeno H., Villagra E., Vera L., Lagos N., Becerra N., Mora J., Bermúdez A., Cárcamo M., Díaz J., Miquel J.F., Ferrer M., López-Lastra M. (2015) Association of Single-Nucleotide Polymorphisms in IL28B, but Not TNF- α , With Severity of Disease Caused by Andes Virus. *Clinical Infectious Diseases*, 61(12): e62–e69. <https://doi.org/10.1093/CID/CIV830>
- Avena S.A., Goicoechea S.A., Rey J., Dugoujon J.M., Dejean C.B., Carnese F.R. (2006) Gene mixture in a population sample from Buenos Aires City. *Medicina*, 66(2): 113–118.
- Avena S., Via M., Ziv E., Pérez-Stable E.J., Gignoux C.R., Dejean C., Huntsman S., Torres-Mejía G., Dutil J., Matta J.L., Beckman K., Burchard E.G., Parolin M.L., Goicoechea A., Acreche N., Boquet M., Ríos Part M.D.C., Fernández V., Rey J., Fejerman L. (2012) Heterogeneity in genetic admixture across different regions of Argentina. *PLoS One*, 7(4). <https://doi.org/10.1371/JOURNAL.PONE.0034695>

- Bamford C.G.G., Aranday-Cortes E., Filipe I.C., Sukumar S., Mair D., Filipe A. da S., Mendoza J.L., Garcia K.C., Fan S., Tishkoff S.A., McLauchlan J. (2018) A polymorphic residue that attenuates the antiviral potential of interferon lambda 4 in hominid lineages. *PLoS Pathogens*, 14(10). <https://doi.org/10.1371/journal.ppat.1007307>
- Booth D., George J. (2013) Loss of function of the new interferon IFN- λ 4 may confer protection from hepatitis C. *Nat. Genet.* 45(2): 119–120. <https://doi.org/10.1038/NG.2537>
- Bravo D., Solano C., Giménez E., Remigia M.J., Corrales I., Amat P., Navarro D. (2014) Effect of the IL28B Rs12979860 C/T polymorphism on the incidence and features of active cytomegalovirus infection in allogeneic stem cell transplant patients. *J. Med. Virol.* 86(5): 838–844. <https://doi.org/10.1002/jmv.23865>
- Chatterjee K. (2010) Host genetic factors in susceptibility to HIV-1 infection and progression to AIDS. *J. Genet.* 89(1): 109–116. <https://doi.org/10.1007/s12041-010-0003-4>
- Chmelova K., Frankova S., Jirsa M., Neroldova M., Sticova E., Merta D., Senkerikova R., Trunecka P., Spicak J., Sperl J. (2019) IL28B rs12979860 T allele protects against CMV disease in liver transplant recipients in the post-prophylaxis and late period. *Transpl. Infect. Dis.* 21(4): e13124. <https://doi.org/10.1111/tid.13124>
- da Silva Cezar R.D., da Silva Castanha P.M., Matos Freire N., Mola C., Feliciano do Carmo R., Tenório Cordeiro M., Baptista P., Silva Vasconcelos L.R., Moura P., da Silva Teixeira V.G. (2020) Association between interferon lambda 3 rs12979860 polymorphism and clinical outcome in dengue virus-infected children. *Int. J. Immunogenet.* 47(4): 351–358. <https://doi.org/10.1111/iji.12477>
- Echeverría N., Chiodi D., López P., Sanchez Ciceron A., Angulo J., López-Lastra M., Silvera P., Canavesi A., Bianchi C., Colistro V., Cristina J., Hernandez N., Moreno P. (2018) IL28B gene polymorphism rs12979860, but not rs8099917, contributes to the occurrence of chronic HCV infection in Uruguayan patients. *Virol. J.* 15(1): 1–10. <https://doi.org/10.1186/s12985-018-0946-2>
- Fan W., Xie S., Zhao X., Li N., Chang C., Li L., Yu G., Chi X., Pan Y., Niu J., Zhong J., Sun B. (2016) IFN- λ 4 desensitizes the response to IFN- α treatment in chronic hepatitis C through long-term induction of USP18. *J. Gen. Virol.* 97(9): 2210–2220. <https://doi.org/10.1099/JGV.0.000522>
- Fang M.Z., Jackson S.S., O'Brien T.R. (2020) IFNL4: Notable variants and associated phenotypes. *Gene*, 730: 144289. <https://doi.org/10.1016/j.gene.2019.144289>
- Fejerman L., Carnese F.R., Goicoechea A.S., Avena S.A., Dejean C.B., Ward R.H. (2005) African ancestry of the population of Buenos Aires. *Am. J. Phys. Anthropol.* 128(1): 164–170. <https://doi.org/10.1002/AJPA.20083>
- Ge D., Fellay J., Thompson A.J., Simon J.S., Shianna K.V., Urban T.J., Heinzen E.L., Qiu P., Bertelsen A.H., Muir A.J., Sulkowski M., McHutchison J.G., Goldstein D.B. (2009) Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature*, 461(7262), 399–401. <https://doi.org/10.1038/nature08309>
- Hamming O.J., Terczyńska-Dyla E., Vieyres G., Dijkman R., Jørgensen S.E., Akhtar H., Siupka P., Pietschmann T., Thiel V., Hartmann R. (2013) Interferon lambda 4 signals via the IFN λ receptor to regulate antiviral activity against HCV and coronaviruses. *EMBO J.* 32(23): 3055–3065. <https://doi.org/10.1038/emboj.2013.232>
- Indolfi G., Mangone G., Calvo P.L., Bartolini E., Regoli M., Serranti D., Calitri C., Tovo P.A., De Martino M., Azzari C., Resti M. (2014a) Interleukin 28B rs12979860 single-nucleotide polymorphism predicts spontaneous clearance of hepatitis C virus in children. *J. Pediatr. Gastroenterol. Nutr.* 58(5): 666–668. <https://doi.org/10.1097/MPG.0000000000000275>
- Indolfi G., Mangone G., Bartolini E., Nebbia G., Calvo P.L., Moriondo M., Tovo P.A., De Martino M., Azzari C., Resti M. (2014b) Comparative analysis of rs12979860 SNP of the IFNL3 gene in children with hepatitis C and ethnic matched controls using 1000 Genomes Project data. *PLoS One*, 9(1): e0085899. <https://doi.org/10.1371/JOURNAL.PONE.0085899>
- Ispiroglu M., Bahcecioglu I.H., Demirel U., Yalniz M. (2017) Impact of interleukin 28B rs12979860 C/T polymorphism on severity of disease and response to treatment in hepatitis delta. *J. Infect. Dev. Ctries.* 11(1): 58–64. <https://doi.org/10.3855/jidc.6872>
- Key F.M., Peter B., Dennis M.Y., Huerta-Sánchez E., Tang W., Prokunina-Olsson L., Nielsen R., Andrés A.M. (2014) Selection on a Variant Associated with Improved Viral Clearance Drives Local Adaptive Pseudogenization of Interferon Lambda 4 (IFNL4). *PLoS Genetics*, 10(10): e1004681. <https://doi.org/10.1371/journal.pgen.1004681>
- Kotenko S.V., Gallagher G., Baurin V.V., Lewis-Antes A., Shen M., Shah N.K., Langer J.A., Sheikh F., Dickensheets H., Donnelly R.P. (2003) IFN- λ s mediate antiviral protection through a distinct class II cytokine receptor complex. *Nat Immunol.* 4(1): 69–77. <https://doi.org/10.1038/ni875>
- Lazear H.M., Nice T.J., Diamond M.S. (2015) Interferon- λ : Immune Functions at Barrier Surfaces and Beyond. *Immunity*, 43(1): 15–28. <https://doi.org/10.1016/j.immuni.2015.07.001>
- Machicote A., Flichmann D., Arana E., Paz S., Fainboim H., Fainboim L., Fernández P. M., Muñoz F.J., Aires B. (2018) IL28B SNPs rs12979860 and rs8099917 Are Associated with Inflammatory Response in Argentine Chronic HCV Patients. *Int. J. Clin. Med.* 9: 79–91. <https://doi.org/10.4236/ijcm.2018.92009>
- Mersha T.B., Abebe T. (2015) Self-reported race/ethnicity in the age of genomic research: its potential impact on understanding health disparities. *Hum. Genomics*, 9(1): 1–15. <https://doi.org/10.1186/S40246-014-0023-X>
- Miretti M.M., Beck S. (2006) Immunogenomics: molecular hide and seek. *Hum. Genomics*, 2(4): 244–251. <https://doi.org/10.1186/1479-7364-2-4-244>
- Muzzio M., Motti J.M.B., Paz Sepulveda P.B., Yee M., Cooke T., Santos M.R., Ramallo V., Alfaro E.L., Dipierri J.E., Bailliet G., Bravi C.M., Bustamante C.D., Kenny E.E. (2018) Population structure in Argentina. *PLoS One*, 13(5): e0196325. <https://doi.org/10.1371/JOURNAL.PONE.0196325>
- O'Brien T.R., Prokunina-Olsson L., Donnelly R.P. (2014) IFN- λ 4: the paradoxical new member of the interferon lambda family. *J. Interferon Cytokine Res.* 34(11): 829–838. <https://doi.org/10.1089/JIR.2013.0136>
- Paschetta C., de Azevedo S., Ramallo V., Cintas C., Perez O., Navarro P., Bandieri L., Quinto-Sanchez M., Adhikari K., Bortolini M.C., Poletti Ferrara G., Gallo C., Bedoya G., Rothhammer F., Acuña Alonzo V., Ruiz-Linares A., Gonzalez-Jose, R. (2021) The impact of socioeconomic and phenotypic traits on self-perception of ethnicity in Latin America. *Sci Rep* 11(1):12617. <https://doi.org/10.1038/s41598-021-92061-x>
- Peakall R., Smouse P.E. (2012) GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research--an update. *Bioinformatics*, 28(19): 2537–2539. <https://doi.org/10.1093/BIOINFORMATICS/BTS460>
- Pedergnana V., Abdel-Hamid M., Guernon J., Mohsen A., Le Foulter L., Theodorou I., Mohamed M.K., Fontanet A., Plancoulaine S., Abel L. (2012) Analysis of IL28B variants in an Egyptian population defines the 20 Kilobases minimal region involved in spontaneous clearance of hepatitis C virus. *PLoS ONE*, 7(6): e38578. <https://doi.org/10.1371/journal.pone.0038578>
- Peng K., Safonova Y., Shugay M., Popejoy A.B., Rodriguez O.L., Breden F., Brodin P., Burkhardt A.M., Bustamante C., Cao-Lormeau V.M., Corcoran M.M., Duffy D., Fuentes-Guajardo M., Fujita R., Greiff V., Jönsson V.D., Liu X., Quintana-Murci L., Rossetti M., ... Mangul S. (2021) Diversity in immunogenomics: the value and the challenge. *Nature Methods*, 18(6): 588–591. <https://doi.org/10.1038/S41592-021-01169-5>
- Porto L.C., Fabrício-Silva G.M., Poschetzky B., Perez R., Carneiro dos Santos R., Cavalini L. (2015) Association of cytokine gene

- polymorphisms with hepatitis C virus infection in a population from Rio de Janeiro, Brazil. *Hepat. Med.* 7: 71-79. <https://doi.org/10.2147/HMER.S89447>
- Prokunina-Olsson L. (2019) Genetics of the Human Interferon Lambda Region. *J. Interferon Cytokine Res.* 39(10): 599-608. <https://doi.org/10.1089/jir.2019.0043>
- Prokunina-Olsson L., Muchmore B., Tang W., Pfeiffer R.M., Park H., Dickensheets H., Hergott D., Porter-Gill P., Mumy A., Kohaar I., Chen S., Brand N., Tarway M., Liu L., Sheikh F., Astemborski J., Bonkovsky H.L., Edlin B.R., Howell C.D., ... O'Brien T.R. (2013) A variant upstream of IFNL3 (IL28B) creating a new interferon gene IFNL4 is associated with impaired clearance of hepatitis C virus. *Nature Genetics*, 45(2): 164-171. <https://doi.org/10.1038/ng.2521>
- Rahimi P., Tarharoudi R., Rahimpour A., Mosayebi Amroabadi J., Ahmadi I., Anvari E., Siadat S.D., Aghasadeghi M., Fateh A. (2021) The association between interferon lambda 3 and 4 gene single-nucleotide polymorphisms and the recovery of COVID-19 patients. *Virol. J.* 18(1): 1-7. <https://doi.org/10.1186/S12985-021-01692-Z>
- Ramamurthy N., Marchi E., Ansari M.A., Pedergnana V., Mclean A., Hudson E., Bowden R., Spencer C.C.A., Barnes E., Klenerman P. (2018) Impact of Interferon Lambda 4 Genotype on Interferon-Stimulated Gene Expression During Direct-Acting Antiviral Therapy for Hepatitis C. *Hepatology*, 68(3): 859-871. <https://doi.org/10.1002/hep.29877>
- Randolph H.E., Fiege J.K., Thielen B.K., Mickelson C.K., Shiratori M., Barroso-Batista J., Langlois R.A., Barreiro L.B. (2021) Genetic ancestry effects on the response to viral infection are pervasive but cell type specific. *Science*, 374(6571): 1127-1133. <https://doi.org/10.1126/SCIENCE.ABG0928>
- Rizzo S.R.C.P., Gazito D., Pott-Junior H., Latini F.R.M., Castelo A. (2016) Prevalence of IFNL3 gene polymorphism among blood donors and its relation to genomic profile of ancestry in Brazil. *Braz. J. Infect. Dis.* 20(6): 619-622. <https://doi.org/10.1016/J.BJID.2016.10.002>
- Saponi-Cortes J.M.R., Rivas M.D., Calle-Alonso F., Sanchez J.F., Costo A., Martin C., Zamorano J. (2021) IFNL4 genetic variant can predispose to COVID-19. *Sci. Rep.* 11(1): 1-4. <https://doi.org/10.1038/S41598-021-00747-Z>
- Sharafi H., Pouryasin A., Alavian S.M., Behnava B., Keshvari M., Mehrnoush L., Salimi S., Kheradvar O. (2012) Development and validation of a simple, Rapid and inexpensive PCR-RFLP method for genotyping of common IL28b polymorphisms: A useful pharmacogenetic tool for prediction of Hepatitis C treatment response. *Hepat. Mon.* 12(3): 190-195. <https://doi.org/10.5812/hepatmon.849>
- Sheppard P., Kindsvogel W., Xu W., Henderson K., Schlutsmeyer S., Whitmore T.E., Kuestner R., Garrigues U., Birks C., Roraback J., Ostrander C., Dong D., Shin J., Presnell S., Fox B., Haldeman B., Cooper E., Taft D., Gilbert T., ... Klucher K.M. (2003) IL-28, IL-29 and their class II cytokine receptor IL-28R. *Nat. Immunol.* 4(1): 63-68. <https://doi.org/10.1038/ni873>
- Taheri S., Aygen B., Korkmaz K., Yildiz O., Zararsiz G., Canatan H. (2015) Characterization of the Interleukin-28B Gene rs12979860 C/T Polymorphism in Turkish Chronic Hepatitis C Patients and Healthy Individuals. *Balk. Med. J.* 32(2): 147-155. <https://doi.org/10.5152/BALKANMEDJ.2015.15156>
- Thomas D.L., Thio C.L., Martin M.P., Qi Y., Ge D., Ohuigin C., Kidd J., Kidd K., Khakoo S.I., Alexander G., Goedert J.J., Kirk G.D., Donfield S.M., Rosen H.R., Tobler L.H., Busch M.P., McHutchison J.G., Goldstein D.B., Carrington M. (2009) Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature*, 461(7265): 798-801. <https://doi.org/10.1038/nature08463>
- Wu L.S., Wang H., Geng X.P. (2012) Two IL28B polymorphisms are associated with the treatment response of different genotypes of hepatitis C in different racial populations: A meta-analysis. *Exp. Ther. Med.* 3(2): 200-206. <https://doi.org/10.3892/ETM.2011.385>
- Zaidane I., Wakrim L., Oulad Lahsen A., Bensghir R., Chihab H., Jadid F.Z., El fihry R., Lamdini H., Fayssel N., Marhoum El Filali K., Oudghiri M., Benjelloun S., Ezzikouri S. (2018) Interleukin 28B rs12979860 genotype and Human Immunodeficiency Virus type 1: Susceptibility, AIDS development and therapeutic outcome. *Hum. Immunol.* 79(1): 70-75. <https://doi.org/10.1016/J.HUMIMM.2017.10.011>