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Complete Genome Sequence of *Chlamydia abortus* MRI-10/19, Isolated from a Sheep Vaccinated with the Commercial Live *C. abortus* 1B Vaccine Strain

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ABSTRACT We report the complete genome sequence of *Chlamydia abortus* MRI-10/19, recovered from the infected placenta of a sheep that had been vaccinated with the commercial live attenuated *C. abortus* 1B vaccine strain. Comparative analysis revealed 1 single nucleotide polymorphism (SNP) difference and 4 indels compared to the vaccine strain.

Chlamydia abortus, an obligate intracellular Gram-negative bacterium and a cause of enzootic abortion of ewes (EAE), is responsible for late-term abortion, stillbirths, and the birth of weak offspring (1). In Europe, the disease is controlled using the commercial live *C. abortus* 1B vaccine (2), which has been associated with infections and cases of abortion in vaccinated ewes (3–7).

Here, we report the complete genome sequence of *C. abortus* strain MRI-10/19, isolated from the placenta of a sheep that had been vaccinated with the commercial 1B strain (Ceva *Chlamydia*, Ceva Animal Health Ltd.) and showed evidence of gross lesions typical of EAE (8). The strain was isolated from pooled placental tissue following the inoculation of ground-up and filtered material onto HEp-2 cells (8). Elementary bodies were purified from infected cultures (9) and genomic DNA extracted using a DNeasy blood and tissue kit (Qiagen). The DNA concentration and purity were determined using a Qubit double-stranded DNA (dsDNA) broad-range (BR) assay kit (Invitrogen) and a NanoDrop One spectrophotometer (Thermo Scientific), respectively.

A genomic DNA library was prepared using the Nextera XT library preparation kit for sequencing on an Illumina HiSeq platform using a 250-bp paired-end protocol. The reads were adapter trimmed using Trimmomatic v0.30 (10) with a sliding window quality cutoff of Q15. Taxonomic classification to the species level as *C. abortus* was confirmed using Kraken v2.1.1 (11). A long-read genomic DNA library was prepared using a rapid barcoding kit (SQK-RBK004) and sequenced in a MinION FLO-MIN106 flow cell (MinKNOW v20.10.3), with integrated live base calling provided by Guppy v4.4.1 (Oxford Nanopore Technologies). The reads were filtered using FilTlong v0.2.0 (12) with a minimum cutoff of 5,000 bp. All trimmed raw data analysis was performed on the Galaxy platform (<http://usegalaxy.org.au/>) (13). The read quality was checked using FastQC (Galaxy v0.72+galaxy1) (14) and NanoPlot (Galaxy v1.28.2+galaxy1) (15). Sequencing resulted in 232,402 Illumina paired-end reads (average read length, 242 bp) and 363,047 filtered Nanopore reads (average read length, 6,842 bp; read N_{50} , 6,663 bp). Hybrid *de novo* assembly of the Illumina and Nanopore raw reads was carried out using the Unicycler pipeline (Galaxy v0.4.8.0) (16), producing a single contig comprising a circular chromosome of 1,144,464 bp with 39.9% GC content and oriented at the *hemB* gene. The average genome coverages for the Illumina and Nanopore read sequences were calculated as 97.4 \times and 2,107.7 \times , respectively, using BWA-MEM (Galaxy v0.7.17.1) (17). The assembly metrics were calculated using QUAST

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(Galaxy v5.0.2+galaxy1) (18). Annotation using the NCBI Prokaryotic Genome Annotation Pipeline v5.0 (19) identified 1,004 predicted genes and 1 rRNA operon. Default settings were used throughout for all utilized software packages.

Pairwise comparative analysis of the assembled sequence with the *C. abortus* 1B Cevac vaccine strain (GenBank accession number [LN589721.1](https://doi.org/10.1093/bioinformatics/btu170)) using Mauve v2015.02.26 (20) identified one SNP at position 131000 and indels in homopolymeric (poly-G) tracts at positions 320135, 684576, 687229, and 991350, identifying the strain as originating from infection with the commercial live vaccine and being responsible for the reported pathological placental lesions (8).

Data availability. The *C. abortus* MRI-10/19 genome sequence is available in GenBank/EMBL/DDBJ under accession number [CP070224](https://doi.org/10.1093/bioinformatics/btu170). The raw sequence reads are available under BioProject accession number [PRJNA700999](https://doi.org/10.1093/bioinformatics/btu170).

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REFERENCES

- Aitken ID, Longbottom D. 2007. Chlamydial abortion, p 105–112. In Aitken ID (ed), *Diseases of sheep*. Blackwell Publishing, Oxford, United Kingdom.
- Rodolakis A, Souriau A. 1983. Response of ewes to temperature-sensitive mutants of *Chlamydia psittaci* (var *ovis*) obtained by NTG mutagenesis. *Ann Rech Vet* 14:155–161.
- Laroucau K, Vorimore F, Sachse K, Vretou E, Siarkou VI, Willems H, Magnino S, Rodolakis A, Bavoil PM. 2010. Differential identification of *Chlamydia abortus* live vaccine strain 1B and *C. abortus* field isolates by PCR-RFLP. *Vaccine* 28:5653–5656. <https://doi.org/10.1016/j.vaccine.2010.06.064>.
- Wheelhouse N, Aitchison K, Laroucau K, Thomson J, Longbottom D. 2010. Evidence of *Chlamydia abortus* vaccine strain 1B as a possible cause of ovine enzootic abortion. *Vaccine* 28:5657–5663. <https://doi.org/10.1016/j.vaccine.2010.04.114>.
- Sargison ND, Truysers IGR, Howie FE, Thomson JR, Cox AL, Livingstone M, Longbottom D. 2015. Identification of the 1B vaccine strain of *Chlamydia abortus* in aborted placentas during the investigation of toxæmic and systemic disease in sheep. *N Z Vet J* 63:284–287. <https://doi.org/10.1080/00480169.2015.1018365>.
- Laroucau K, Aaziz R, Vorimore F, Menard MF, Longbottom D, Denis G. 2018. Abortion storm induced by the live *C. abortus* vaccine 1B strain in a vaccinated sheep flock, mimicking a natural wild-type infection. *Vet Microbiol* 225:31–33. <https://doi.org/10.1016/j.vetmic.2018.09.012>.
- Longbottom D, Sait M, Livingstone M, Laroucau K, Sachse K, Harris SR, Thomson NR, Seth-Smith HMB. 2018. Genomic evidence that the live *Chlamydia abortus* vaccine strain 1B is not attenuated and has the potential to cause disease. *Vaccine* 36:3593–3598. <https://doi.org/10.1016/j.vaccine.2018.05.042>.
- Caspe SG, Livingstone M, Frew D, Aitchison K, Wattedegedera SR, Entrican G, Palarea-Albaladejo J, McNeilly TN, Milne E, Sargison ND, Chianini F, Longbottom D. 2020. The 1B vaccine strain of *Chlamydia abortus* produces placental pathology indistinguishable from a wild type infection. *PLoS One* 15:e0242526. <https://doi.org/10.1371/journal.pone.0242526>.
- Thomson NR, Yeats C, Bell K, Holden MTG, Bentley SD, Livingstone M, Cerdeño-Tárraga AM, Harris B, Doggett J, Ormond D, Mungall K, Clarke K, Feltwell T, Hance Z, Sanders M, Quail MA, Price C, Barrell BG, Parkhill J, Longbottom D. 2005. The *Chlamydia abortus* genome sequence reveals an array of variable proteins that contribute to interspecies variation. *Genome Res* 15:629–640. <https://doi.org/10.1101/gr.3684805>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Wood DE, Lu J, Langmead B. 2019. Improved metagenomic analysis with Kraken 2. *Genome Biol* 20:257. <https://doi.org/10.1186/s13059-019-1891-0>.
- Wick RR. 2017. Filong: quality filtering tool for long reads. <https://github.com/rwick/filong>.
- Afgan E, Baker D, van den Beek M, Blankenberg D, Bouvier D, Čech M, Chilton J, Clements D, Coraor N, Eberhard C, Grüning B, Guerler A, Hillman-Jackson J, Von Kuster G, Rasche E, Soranzo N, Turaga N, Taylor J, Nekrutenko A, Goecks J. 2016. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2016 update. *Nucleic Acids Res* 44:W3–W10. <https://doi.org/10.1093/nar/gkw343>.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>.
- De Coster W, D'Hert S, Schultz T, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. *Bioinformatics* 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv* 1303.3997v2 [q-bio.GN]. <https://arxiv.org/abs/1303.3997v2>.
- Gurevich A, Saveliev V, Vyahh N, Tesler G. 2013. QUASt: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Darling ACE, Mau B, Blattner FR, Perna NT. 2004. Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res* 14:1394–1403. <https://doi.org/10.1101/gr.2289704>.