



SHORT COMMUNICATION

OPEN ACCESS

Embryo yield in llamas synchronized with two different intravaginal progesterone-releasing devices and superovulated with eCG

Juan F. Aller¹, Marcos C. Abalos², Francisco A. Acuña², Rosana Virgili³, Francisco Requena⁴ and Andrea K. Cancino⁵

¹ Instituto Nacional de Tecnología Agropecuaria (INTA), Estación Experimental Agropecuaria Balcarce. C.C. 276, 7620 Balcarce, Argentina. ² INTA, Estación Experimental Agropecuaria Abra Pampa. 4640 Abra Pampa, Jujuy, Argentina. ³ Universidad Nacional de Jujuy, Facultad de Ciencias Agrarias. 4600 S.S. Jujuy, Argentina. ⁴ Universidad de Córdoba, Facultad de Veterinaria, Campus Universitario Rabanales. 1471 Córdoba, Spain. ⁵ INTA, Estación Experimental Agropecuaria Bariloche. 8400 Bariloche, Argentina

Abstract

The objectives of this study were to compare the effects of two intravaginal devices (ID) containing the same dose (0.5 g) of progesterone (P₄) on subsequent ovarian response, embryo production and circulating P₄ concentration profile in llamas (*Lama glama*) treated with equine chorionic gonadotropin (eCG) for ovarian superstimulation. Female llamas were randomly assigned (n = 10 llamas per group) to one of the following groups and treated (Day 0) with an ID containing 0.5 g of vegetal P₄ to synchronize the emergence of a new follicular wave: i) DIB 0.5° and ii) Cronipres M15°. On Day 3 llamas were intramuscularly treated with 1000 IU of eCG. The IDs were removed on Day 7. Llamas were naturally mated (Day 9) and treated with GnRH analogue to induce ovulation. A second mating was allowed 24 h later. Embryos were collected between 7 and 8 days after the first mating. Blood samples were taken every day from Day 0 to Day 7 to measure circulating P₄ concentrations. The results indicated that DIB device maintained greater plasma P₄ levels as compared to Cronipres until Day 2. However, the mean (± SD) number of corpora lutea and recovered embryos was not affected (p < 0.05) by the type of ID (5.3 ± 2.6 vs 4.2 ± 2.2 and 3.5 ± 2.7 vs 2.6 ± 3.0 for DIB and Cronipres, respectively). In conclusion, both DIB and Cronipres devices can be successfully used to synchronize the emergence of follicular wave prior to a single dose of eCG in superovulation protocol in llamas.

Additional key words: South American camelids; superovulation; hormonal treatment; embryo.

Abbreviations used: CL (corpora lutea); eCG (equine chorionic gonadotropin), GnRH (gonadotropin-releasing hormone); ID (intravaginal device); LH (luteinising hormone); MOET (multiple ovulation and embryo transfer); P₄ (progesterone); SAC (South American camelids)

Citation: Aller, J. F.; Abalos, M. C.; Acuña, F. A.; Virgili, R.; Requena, F.; Cancino, A. K. (2015). Short communication. Embryo yield in llamas synchronized with two different intravaginal progesterone-releasing devices and superovulated with eCG. Spanish Journal of Agricultural Research, Volume 13, Issue 3, e04SC01, 6 pages. http://dx.doi.org/10.5424/sjar/2015133-7308.

Received: 29 Dec 2014. **Accepted:** 20 Jul 2015

Copyright © **2015 INIA.** This is an open access article distributed under the Creative Commons Attribution License (CC by 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Funding: This research was supported by National Institute for Agricultural Technology (National Project N° 1115053) and ArgenINTA Foundation.

Competing interests: The authors have declared that no competing interests exist. Correspondence should be addressed to Juan F. Aller: alleratucha.juan@inta.gob.ar

The South American camelids (SAC), also known as the New World Camelidae, are mainly reared in the High Andes having an important economic and social role in countries such as Argentina, Bolivia, Chile, Peru and in recent years, Ecuador. Eighty-six per cent of the world's 3.5 million alpacas are found in Peru; likewise, the greatest percentage (>70%) of the world's 3.1 million llamas are found in Bolivia (Novoa, 1981). The greater population of guanacos is found in the Argentinean Patagonia, with an estimated number of 600,000 animals and the vicuña population is distributed among all the aforementioned countries. In those countries,

several programs for genetic improvement (meat and fibre) have been undertaken based on assisted reproductive techniques.

Multiple ovulation and embryo transfer (MOET) allow the decrease of generational interval, optimization of mating programs and increasing the reproductive potential from high genetic value females. In llama and alpaca, the MOET is more used than artificial insemination due to poor results using frozen semen (Bravo *et al.*, 2013). Ovarian superstimulation in SAC is performed under different general conditions: i) simulating a luteal phase by using exogenous proges-

terone/progestogen, ii) during a luteal phase following ovulation induced by administration of hCG (human chorionic gonadotropin), LH (luteinising hormone), GnRH (gonadotropin-releasing hormone) or iii) after ablation of all follicles greater than 5 mm using transvaginal ultrasound-guided follicle aspiration. The simplest method to control follicular dynamics and synchronization of the emergence of a new follicular development in llama is to use an intravaginal device (ID) (Chaves *et al.*, 2002) or an ear implant containing progestogen (Bourke *et al.*, 1995).

Progesterone may also have an important effect on oocyte quality decreasing the apoptosis rate of cumulus cells during maturation (Salhab *et al.*, 2011). On the other hand, a positive relationship between greater circulating levels of P₄ during follicular growth and embryo quality have been recently described in dairy cows (Rivera *et al.*, 2011). The precise effect of elevated P₄ during the growth of the dominant follicle on the oocyte and embryo quality is unknown (Fair & Lonergan, 2012) but it may be related to reduced exposure of the developing oocyte to LH, avoiding its premature maturation (Cerri *et al.*, 2011).

In Argentina, different kinds of IDs containing different amounts of P_4 for use in cattle have been developed by pharmaceutical industry in the last 10 years and are exported to all South America, especially to the Andean countries where the SAC are raised. Moreover, IDs containing low dose (≤ 0.5 g) of P_4 should preferably be used to reduce environmental pollution after use in animals.

These IDs, that contain the same P₄ dose, vary in shape and surface contact area, variations which could have an effect on the synchronization obtained. Thus, differences in physical characteristics among IDs produce different plasma P₄ concentrations in cattle (Rathbone *et al.*, 1998; 2002). In a recent study, van Werven *et al.* (2013) found that PRID® produced significantly greater circulating P₄ concentration compared to CIDR® and suggested that both levels of P₄ content and differences in shape and contact surface area might contribute to circulating P₄ concentrations.

Therefore, the objectives of the present work were to compare the effects of two commercially available and commonly used IDs containing low dose of P_4 and varying in their structure and shape on subsequent bioavalability in blood plasma, ovarian response and embryo yield in llamas treated with eCG for ovarian superstimulation.

This work was carried out at Abra Pampa Experimental Station of the National Institute of Agricultural Technology (INTA), located in the province of Jujuy (22° 49′S, 65° 47′W; 3,483 m above sea level) during non-breeding season. Animals weighed 102.3 ± 7.8 kg,

with a body condition score of 4.6 ± 1.1 (1 = emaciated to 9 = obese; Richards et al., 1986) determined by palpation. Llamas were selected (n = 20 from a group of 60) and randomly allocated in equal numbers (n = 10) to one of two groups to be treated (Day 0) with an intravaginal device (i) DIB 0.5® (Syntex, Argentina) (DIB group) or (ii) Cronipres M15® (Biogenesis-Bago, Argentina) (Cronipres group) and both IDs containing 0.5 g of progesterone. Each ID was removed seven days later. Ovarian follicular development was stimulated by intramuscular administration of 1000 IU of eCG (Novormon 5000®, Syntex, Argentina) (Day 3). Llamas were naturally mated (Day 9) with males of proven fertility and immediately treated with 100 µg of GnRH analogue of gonadorelin (Gonasyn GDR®, Syntex, Argentina) as an additional stimulus to induce ovulation. A second mating was allowed 24 h later.

Embryo recovery from the donor females was performed non-surgically 7 days after the first mating as described previously (Aller et al., 2002). Briefly, each female was sedated with 10 mg acepromazine (Acedan®, Holliday, Argentina) and caudal epidural anaesthesia with 3 mL of 2% lidocaine hydrochloride was induced before uterine flushing. Each uterine horn was flushed using 14-Fr Rusch two-way catheter and 250 mL of Ringer lactate supplemented with 1% heatinactivated cow serum. The flushing medium was filtered (EmCon, Minitüb, Germany) and searching embryos was performed using a stereomicroscope at magnification x40. The recovered embryos were transferred to holding medium (Syngro holding medium®, Bioniche Animal Health, Canada) and kept at room temperature and classified according to IETS standards for cattle embryos (IETS Manual, 1998). After embryo collection, ovarian responses in the donor females were evaluated by transrectal ultrasonography using a real time scanner with a 5-MHz linear array transducer (Honda HS 101V, Japan).

Blood samples were collected on Days 0, 1, 2, 3, 4, 7 and 9 by jugular venipuncture into tubes containing sodium heparin (Fada Pharma, Argentina) and immediately centrifuged at 1500 xg for 20 min. The blood plasma was stored at -20°C until hormone analyses. Concentrations of P_4 were measured by a radioimmunoassay commercial kit (Siemens Medical Solutions Diagnostics, Los Angeles, CA, USA) for use in bovine and validated for llama (Aba *et al.*, 1999). Samples were evaluated in duplicate and all samples were analyzed in a single assay. The intra- assay coefficient of variation was < 9%, for concentrations between 0.1 and 40.0 ng/mL. The estimated sensitivity of this method was 0.01 ng/mL.

Data analyses were performed using SAS (1989). Number of unovulated follicles ≥ 7 mm, number of

corpora lutea (CL) and number of recovered embryos were compared between groups using one-way ANOVA. Plasma P_4 concentration was analyzed by ANOVA for repeated measures (PROC MIXED); the model included the effects of treatment, day and treatment by day interaction, with day as a repeated effect. Proportional data were compared by Chi square test. Correlation coefficients (Pearson correlation) between number of CL and number of recovered embryos were calculated (Steel & Torrie, 1980). Probability of p < 0.05 was considered to be statistically significant.

On Day 9, all llamas were sexually receptive and successfully mated. After device removal, a slight vaginitis was observed; however, the fertility was not compromised because ejaculation is deep intracornual in this species. The mean number of unovulated follicles ≥ 7 mm, CL and recovered embryos were not affected (p > 0.05) by treatment (Table 1). The recovery rates (total number of recovered embryos relative to the total number of CL) were 66.0 (35/53) and 61.9% (26/42) for DIB and Cronipres groups respectively (p > 0.05). All recovered embryos, regardless of group, were graded as excellent quality hatched blastocysts. Embryos could not be collected from 4 of the 20 animals (two animals of each group). Approximately 20% of the animals did not respond to the superovulatory treatment. Significant positive correlations between number of CL and number of recovered embryos for DIB and Cronipres groups were detected (r = 0.68, p = 0.02 and r = 0.80, p = 0.005, respectively).

The circulating P_4 concentrations of females from the DIB and Cronipres groups are shown in Fig. 1. An effect of Day \times Treatment interaction was detected on plasma P_4 concentrations. Significant differences

Table 1. Ovarian response and embryo yield (mean \pm SD; range in parenthesis) in llamas treated with 1000 IU of eCG (equine chorionic gonadotropin) after synchronization of follicular wave emergence with two different intravaginal devices containing 0.5 g of progesterone

	DIB Group (n = 10) 1	Cronipres Group (n = 10) ¹	p
No. of follicles ≥ 7 mm on day of embryo recovery	$2.5 \pm 2.7 (0-9)$	3.3 ± 3.2 (0-8)	0.55
No. of CL ² on day of embryo recovery	$5.3 \pm 2.6 (3-11)$	$4.2 \pm 2.2 (3-10)$	0.31
No. of recovered	$3.5 \pm 2.7 (0-8)$	$2.6 \pm 3.0 \ (0-10)$	0.48

¹ Two females of each group did not respond to the superovulatory treatment (0 embryo). ² CL: corpora lutea.

(p < 0.05) in P₄ concentration between groups were determined at 24 and 48 h after ID insertion; however these differences were not observed from Day 3 onwards. Plasma P₄ concentration (mean \pm SD) in llamas that yielded ≥ 2 embryos in both Groups (DIB = 35.6 ± 19.0 ng/mL; Cronipres = 32.0 ± 10.6 ng/mL) were not different (p < 0.05) from those in llamas yielding single or no embryo (DIB = 28.8 ± 16.8 ng/mL; Cronipres = 22.6 ± 14.2 ng/mL).

The main finding from this experiment was that both intravaginal devices (DIB and Cronipres) containing low dose of progesterone used during superovulation treatment produced a similar ovarian response and number of recovered embryos in llamas.

Several research groups have looked for possible associations between endogenous (natural luteal phase) or exogenous progesterone priming (treatment used to control the ovarian follicular dynamics) and the response to superovulation as a potential explanation for some of the persistent variability in superovulation seen between animals (Kanitz *et al.*, 2002; Mapletoft *et al.*, 2002). In this study, the aim was to investigate any differences in quantity and quality of embryos produced in response to either DIB or Cronipres progesterone ID combined with eCG for ovarian superstimulation in llamas.

The use of progesterone to synchronize the follicular wave emergence obviates the need to know the specific follicular stage when starting the superovula-

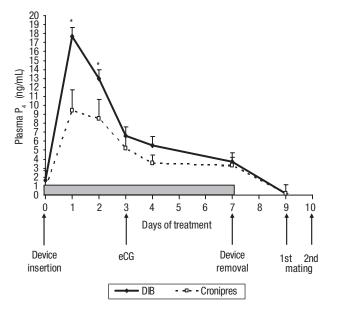


Figure 1. Mean (\pm SD) concentrations of P₄ in plasma of llamas treated with an intravaginal device containing 0.5 g of progesterone (DIB or Cronipres) during 7 days and 1000 IU eCG for superovulation. * DIB differed from Cronipres on Day 1 and Day 2 (p < 0.05). P₄: progesterone; eCG: equine chorionic gonadotropin.

tory treatment and this treatment should be initiated near the time of follicular wave emergence to produce the maximal superovulatory response (Adams $et\ al.$, 1994). In llamas, Aller $et\ al.$ (2010) observed that the follicular wave emergence occurred approximately on Day 4 (\pm one day) after medroxiprogesterone acetate intravaginal sponge insertion. Therefore, in the present study the eCG treatment for ovarian stimulation was administered on Day 3 after ID insertion.

Superovulatory treatment can be initiated in natural luteal phase, but require the use of ultrasonography to detect the growing dominant follicle and then to induce ovulation with human chorionic gonadotrophin (San Martin *et al.*, 1968) or luteinizing hormone (Fernandez-Baca *et al.*, 1970). Therefore, we chose exogenous progesterone because this hormone allows regulate the ovarian function indirectly through LH secretion, rather than by direct actions on the ovaries. The P₄ device is very simple to apply in field conditions to control the ovarian follicular dynamics and predicts the emergence of a new follicular wave and thus to start the ovarian superstimulatory treatment without the use of ultrasound.

In the present study the number of CL and embryos recovered per llama did not differ between DIB and Cronipres group. Huanca *et al.* (2009) using progestin-realising vaginal sponges obtained a greater number of CL (8.6) but the number of recovered embryos (3.5) was similar to our study. Additionally, Carretero *et al.* (2010), who used daily intramuscular administration of progesterone during five days to inhibit follicular growth obtained 2.9 embryos per female. Progestogen implants inserted over a period of 7 days combined with 1000 IU of eCG on Day 5 yielded a low embryo recovery (1.3 embryos per donor female); however, the same protocol using CIDR® improved the embryo recovery (2.0 embryos) (Bourke *et al.*, 1992).

The recovery rates for DIB and Cronipres groups were significantly higher than the recovery rates obtained from other small-scale studies in llama (Correa *et al.*, 1997, 34.5%; Ratto *et al.*, 1997, 16.9%). On the other hand, in a very large data set collected recently by Vaughan *et al.* (2013) from commercial alpaca embryo transfer records the recovery rate was 38.8% (4188 embryos/10796 ovulations).

The effect of elevated progesterone during follicular growth has been linked to improved embryo quality (Lonergan, 2011). Additionally, Rivera *et al.* (2011) showed that high P₄ during ovarian superstimulation treatment of lactating dairy cows increased the quality of embryos collected on Day 7 after estrus. In the present study, all recovered embryos were hatched blastocysts graded as excellent quality. The associations

between number of CL and recovered embryos for DIB and Cronipres groups were similar to that observed by Vaughan *et al.* (2013) in alpacas (r = 0.54). However, no significant correlation (r = 0.12) was detected by Aller *et al.* (2010), possibly as consequence of the high number of ovarian follicles (12.4) induced by the superovulatory treatment; therefore, the ovarian bursa can be displaced leading to a loss of oocytes into the abdominal cavity and a low number of recovered embryos.

Plasma P₄ concentrations rose quickly after ID insertion (Day 0) with peak concentration attained on Day 1. Significant differences of circulating P₄ concentrations between DIB and Cronipres devices were observed at this day. Following 48 h after device removal, P₄ concentrations were not significantly different to pre-treatment concentrations. Treatments with an intravaginal CIDR® device containing 0.33 g of progesterone (Chaves et al., 2002) and medroxiprogesterone acetate-vaginal sponge (Aba et al., 1999; Huanca et al., 2009) were successfully used to control ovarian follicular dynamics. Progesterone concentrations similar to the one observed in our study was described by Chaves et al. (2002) where a rapid increase was observed at Day 1 (~10 ng/mL) and sharply decreased until Day 4 (2 ng/mL).

The two types of ID used in the present study have the same P_4 content (0.5 g), therefore differences in contact surface area (DIB ~95 cm² vs Cronipres ~60 cm²) might contribute to differences in plasma P_4 concentrations. Additionally, because the shape of the two IDs differ greatly (DIB = V-shape vs. Cronipres = double L-like with three hood containing progesterone) it may influencing the surface area of the devices in direct contact with the vagina wall since the rate of diffusion of P_4 from ID to bloodstream could be considered similar, because the two IDs have the same type of outer layer material (inert silicone).

The results supported the hypothesis that the shape and contact surface area with the vagina wall of two ID (DIB and Cronipres) containing the same P_4 levels have effect on the plasma P_4 concentrations. In spite of the differences in physical characteristic and the plasma P_4 concentrations observed between devices, the ovarian superstimulation protocols using eCG combined with DIB or Cronipres were equally effective to induce a good superovulatory response and embryo production.

Acknowledgments

Authors acknowledge the expert technical assistance of the Abra Pampa Experimental Station technical staff.

References

- Aba MA, Quiroga MA, Auza N, Forsberg M, Kindahl, H, 1999. Control of ovarian activity in llamas (*Lama glama*) with medroxiprogesterone acetate. Reprod Dom Anim 34: 471-476. http://dx.doi.org/10.1111/j.1439-0531.1999.tb01406.x
- Adams GP, Nasser LF, Bo GA, Garcia A, del Campo, MR, Mapletoft RJ, 1994. Superovulatory response of ovarian follicles of wave 1 versus wave 2 in heifers. Theriogenology 42: 1103-1113. http://dx.doi.org/10.1016/0093-691X(94)90858-3
- Aller JF, Rebuffi GE, Cancino AK, Alberio RH, 2002. Successful transfer of vitrified llama (*Lama glama*) embryos. Anim Reprod Sci 73: 121-127. http://dx.doi.org/10.1016/S0378-4320(02)00120-3
- Aller JF, Cancino AK, Rebuffi GE, Alberio RH, 2010. Effect of estradiol benzoate used at the start of a progestagen treatment on superovulatory response and embryo yield in lactating and non-lactating llamas. Anim Reprod Sci 119: 322-328. http://dx.doi.org/10.1016/j.anireprosci.2010.02.001
- Bourke DA, Adam C.L, Kyle CE, Young P, McEvoy TG, 1992. Superovulation and embryo transfer in the llama. Proc First Int Camel Conf; Allen WR, Higgins AJ, Maythew IG, Snow D, Wade JF (Eds). R&W Publ., Newmarket, UK. pp: 129-137.
- Bourke DA, Kyle CE, McEvoy TG, Young P, Adam CL, 1995. Superovulatory responses to eCG in llamas (*Lama glama*). Theriogenology 44: 255-268. http://dx.doi.org/10.1016/0093-691X(95)00175-8
- Bravo PW, Alarcón V, Baca L, Cuba Y, Ordoñez C, Salinas J, Tito F, 2013. Semen preservation and artificial insemination in domesticated South American camelids. Anim Reprod Sci 136: 157-163. http://dx.doi.org/10.1016/j.anireprosci.2012.10.005
- Carretero MI, Miragaya M, Chaves MG, Gambarotta M, Agüero A, 2010. Embryo production in superstimulated llamas pre-treated to inhibit follicular growth. Small Rumin Res 88: 32-37. http://dx.doi.org/10.1016/j.small-rumres.2009.11.006
- Cerri RI, Chebel RC, Rivera F, Narciso CD, Oliveira RA, Thatcher WW, 2011. Concentration of progesterone during the development of the ovulatory follicle: I. Ovarian and embryonic responses. J Dairy Sci 94: 3342-3351. http://dx.doi.org/10.3168/jds.2010-3734
- Chaves MG, Aba M, Agüero A, Egey J, Berestin V, Rutter B, 2002. Ovarian follicular wave pattern and the effect of exogenous progesterone on follicular activity in nonmated llamas. Anim Reprod Sci 69: 37-46. http://dx.doi.org/10.1016/S0378-4320(01)00173-7
- Correa JE, Ratto MH, Gatica R, 1997. Superovulation in llamas (*Lama glama*) with pFSH and equine chorionic gonadotrophin used individually or in combination. Anim Reprod Sci 46: 289-296. http://dx.doi.org/10.1016/S0378-4320(96)01615-6
- Fair T, Lonergan P, 2012. The role of progesterone in oocyte acquisition of developmental competence. Reprod Dom Anim 47 (Suppl. 4): 142-147. http://dx.doi.org/10.1111/j.1439-0531.2012.02068.x

- Fernandez-Baca S, Hansel, W, Novoa C, 1970. Corpus luteum function in the alpaca. Biol Reprod 3: 252-261.
- Huanca W, Cordero A, Huanca T, Cardenas O, Adams GP, Ratto MH, 2009. Ovarian response and embryo production in llamas treated with equine chorionic gonadotropin alone or with a progestin-releasing vaginal sponge at the time of follicular wave emergence. Theriogenology 72: 803-808. http://dx.doi.org/10.1016/j.theriogenology.2009.05.019
- IETS, 1998. Manual IETS; Stringfellow DA & Seidel SM (Eds), 3rd Edition. Savoy, IL, USA.
- Kanitz W, Becker F, Schneider F, Kanitz E, Leiding C, Nohner H, Pöhland R, 2002. Superovulation in cattle: practical aspects of gonadotropin treatment and insemination. Reprod Nutr Dev 42: 587-599. http://dx.doi.org/ 10.1051/rnd:2002045
- Lonergan P, 2011. Influence of progesterone on oocyte quality and embryo development in cows. Theriogenology 76: 1594-1601. http://dx.doi.org/10.1016/j.theriogenology.2011.06.012
- Mapletoft RJ, Bennett Steward K, Adams GP, 2002. Recent advances in the superovulation in cattle. Reprod Nutr Dev 42: 601-611. http://dx.doi.org/10.1051/rnd:2002046
- Novoa C, 1981. La conservación de especies nativas en América Latina. Animal Genetic Resources Conservation and Management, FAO Anim Prod Health, Paper 24, FAO, Rome, pp. 349-363.
- Rathbone MJ, Macmillan KL, Inskeep K, Burggraaf S, Bunt CR, 1998. Fertility regulation in cattle. J Control Release 54: 117-148. http://dx.doi.org/10.1016/S0168-3659(98) 00003-0
- Rathbone MJ, Bunt CR, Ogle CR, Burggraaf S, Macmillan KL, Burke CR, Pickering KL. 2002. Reengineering of the commercially available bovine intravaginal insert (CIDR insert) containing progesterone. J Control Release 85: 105-115. http://dx.doi.org/10.1016/S0168-3659(02) 00288-2
- Ratto MH, Gatica R, Correa JE, 1997. Timing of mating and ovarian response in llamas *(Lama glama)* treated with pFSH. Anim Reprod Sci 48: 325-330. http://dx.doi.org/10.1016/S0378-4320(97)00047-X
- Richards MW, Spitzer JC, Warner MB, 1986. Effect of varying levels of postpartum nutrition at body condition at calving on subsequent reproductive performance in beef cattle. J Anim Sci 62: 300-306.
- Rivera FA, Mendonca LG, Lopes GJr, Santos JEP, Perez RV, Amstalden M, Correa-Calderón A, Chebel RC, 2011. Reduced progesterone concentration during growth of the first follicular wave affects embryo quality but has no effect on embryo survival post transfer in lactating dairy cows. Reproduction 141: 333-342. http://dx.doi.org/10.1530/REP-10-0375
- Salhab M, Tosca L, Cabau C, Papillier P, Perreau C, Dupont J, Mermillod P, Uzbekova S, 2011. Kinetics of gene expression and signaling in bovine cumulus cells throughout IVM in different mediums in relation to oocyte developmental competence, cumulus apoptosis and P4 secretion. Theriogenology 75: 90-104. http://dx.doi.org/10.1016/j. theriogenology.2010.07.014
- San-Martin M, Copaira M, Zuniga J, Rodriguez R, Bustinza G, Acosta L, 1968. Aspects of reproduction in the alpaca.

- J Reprod Fertil 16: 395-399. http://dx.doi.org/10.1530/jrf.0.0160395
- SAS, 1989. SAS/STAT® User's Guide, Version 6, vol. 2, 4th ed. SAS Institute Inc., Cary, NC, USA. 848 pp.
- Steel RGD, Torrie JH, 1980. Principles and procedures of statistics, 2nd ed. McGraw-Hill, NY, USA.
- Van Werven T, Waldeck F, Souza AH, Floch S, Englebienne M, 2013. Comparison of two intravaginal progesterone
- releasing devices (PRID-Delta vs CIDR) in dairy cows: Blood progesterone profile and field fertility. Anim Reprod Sci 138: 143-149. http://dx.doi.org/10.1016/j.anireprosci.2013.02.010
- Vaughan J, Mihm M, Wittek T, 2013. Factors influencing embryo transfer success in alpacas-A retrospective study. Anim Reprod Sci 136: 194-204. http://dx.doi.org/10.1016/j.anireprosci.2012.10.010