

# New Insights into Soybean Biological Nitrogen Fixation

Ignacio A. Ciampitti\* and Fernando Salvagiotti\*

## ABSTRACT

Soybean biological N<sub>2</sub> fixation (BNF) relationships with fertilizer N and yield response have been comprehensively reviewed in the scientific literature. However, the study of the N-gap between N uptake and N supplied by N<sub>2</sub> fixation, and the partial N balance (fixed N in aboveground biomass – N seeds) needs further investigation. Therefore, the goals of this synthesis-analysis were to (i) quantify seed production per unit of fixed N under different amounts of N derived from the atmosphere (NDFA, %), (ii) study the N-gap and explore limitations of N<sub>2</sub> fixation (kg ha<sup>-1</sup>) for satisfying plant N demand, and (iii) calculate a partial N balance for soybean and determine its relationship with the N<sub>2</sub> fixation process. Data was gathered from 1955 through 2016 using studies reporting BNF, seed yield, and plant N uptake (*n* = 733 data points). The main outcomes of this review were (i) as NDFA increased, seed production per N<sub>2</sub> fixation decreased (from 0.033 to 0.017 Mg yield kg<sup>-1</sup> N from low, 28%, to high, 80%, NDFA); (ii) N-gap increased faster when NDFA values were above 80% and after plant N content was above 370 kg N ha<sup>-1</sup> suggesting that the crop needs additional N for coping yield potential; and (iii) when excluding roots, the partial N balance calculation revealed negative values across all NDFA levels. Future studies should consider a holistic approach to quantify the contribution of BNF in overall N cycling, including N contribution from roots, and to better understand the soil × plant × rhizobia interactions.

## Core Ideas

- As N<sub>2</sub> fixation (%) increased, seed production per N<sub>2</sub> fixation decreased.
- The N-gap between crop N uptake and N supplied by N<sub>2</sub> fixation rose when contribution from biological N<sub>2</sub> fixation increased.
- The partial N balance revealed negative values across all N derived from the atmosphere levels.
- Yield was negatively related to partial N balance when N derived from the atmosphere was below 42%.

**S**OYBEAN (*GLYCINE max* L.) is one of the most important crops grown worldwide as source of both protein and oil. Three countries are the main soybean producers at the global scale: Argentina, Brazil, and the United States; they comprise 16, 32, and 33%, respectively, of the estimated global soybean production (USDA NASS, 2017). From a historical perspective, soybean yield, harvested area, and total production have been increasing in all three countries but at different rates (SoyStat, 2016). Historical gains in soybean seed yields are primarily due to increases in seed biomass, which demonstrates an improvement in seed partitioning efficiency (Koester et al., 2014). The increase in partitioning efficiency and seed biomass requires larger N demand (Balboa et al., 2018), primarily met by biological N<sub>2</sub> fixation (BNF) and soil N mineralization.

In soybean, N derived from the atmosphere (NDFA) via BNF can range from 0 to 98% of the total N uptake, representing 0 to 337 kg N ha<sup>-1</sup> (Salvagiotti et al., 2008), depending on rhizobia activity. However, N removal from the system (i.e., by seed N) is determined by different factors that affect seed yield and N harvest index (NHI; seed N uptake to total N uptake). In a recent study, Tamagno et al. (2017) showed that NHI in soybean grown in three different regions ranged from 44 to 91% (at R7–R8 growth stages), with yields ranging from ~1 to 8 Mg ha<sup>-1</sup>. The previous information related to NDFA contribution and NHI calculation for soybeans portrays the complexity of estimating N contribution of soybeans to the rotation. A review study summarizing 108 scientific papers published from 1966 to 2006 documented an average NDFA contribution of 50 to 60% (Salvagiotti et al., 2008). Comparable NDFA estimations were documented in Argentina: 60% (ranging 46–71%) (Collino et al., 2015) and up to 80% in less fertile soils in Brazil (Alves et al., 2003). The review by Salvagiotti et al. (2008) suggested that the NDFA contribution was not sufficient for high-yielding soybeans (>7 Mg ha<sup>-1</sup>). Salvagiotti et al. (2009) showed a slight increase in seed yield in crops that yielded more than 5 Mg ha<sup>-1</sup> when N was supplied without affecting the N<sub>2</sub> fixation process. However, the question, whether N<sub>2</sub> fixation alone can supply N for a high-yielding soybean while maintaining a neutral partial N balance (fixed N in aboveground biomass minus N removed in seeds), remains unanswered.

Published in *Agron. J.* 110:1185–1196 (2018)

doi:10.2134/agronj2017.06.0348

Supplemental material available online

Available freely online through the author-supported open access option

Copyright © 2018 by the American Society of Agronomy

5585 Guilford Road, Madison, WI 53711 USA

This is an open access article distributed under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

I.A. Ciampitti, Dep. of Agronomy, Kansas State Univ., Manhattan, KS 66506; F. Salvagiotti, Dep. Agronomía, EEA INTA Oliveros, Ruta 11 km 353 (C 2206), Santa Fe, Argentina. Received 23 June 2017. Accepted 1 Apr. 2018. \*Corresponding authors (ciampitti@ksu.edu; salvagiotti.fernando@inta.gob.ar).

**Abbreviations:** BNF, biological N<sub>2</sub> fixation; NHI, N harvest index; NIE, N internal efficiency; N-seed, seed N content, NDFA, N derived from the atmosphere.

The “N-gap” between crop N uptake and N supplied by N<sub>2</sub> fixation has not yet been estimated at varying yield and NDFA levels. A better understanding of the so-called N-gap could allow development of potential N management strategies to further boost soybean yields and profitability. These strategies should take into account the trade off between inorganic N fertilizer and fixed N (Salvagiotti et al., 2008). High-yielding soybean environments should be accompanied by high N<sub>2</sub> fixation activity, as suggested by van Kessel and Hartley (2000). Collino et al. (2015) showed greater N<sub>2</sub> fixation as yield potential increased. However, if N<sub>2</sub> fixation contribution remains constant (or increasing less than proportional to yield) at increasing productivity levels, then mineral soil N contribution should meet the crop demand. Consequently, N could be mined from the soil producing negative partial N balance and affecting soil health.

Soybean breeding efforts have been focused on improving partitioning efficiency (seed yield per unit of biomass) (Kumudini et al., 2001, 2002; Koester et al., 2014) and increasing the duration of early reproductive stages. However, the reduction in N<sub>2</sub> fixation rates between R5 and R7 stages (Zapata et al., 1987), even when not always consistent, could lead to N limitation during seed filling, as recently documented by Tamagno and Ciampitti (2017). The same authors found that treatments with high fertilizer N rate extended the duration of the seed filling, without modifying seed growth rate, by 5 d as compared with treatments depending only on BNF, with overall yields for the high N fertilization of 4 Mg ha<sup>-1</sup>. Thus, it can be hypothesized that reaching maximum yield cannot solely depend on N<sub>2</sub> fixation to satisfy soybean N demand. Following this rationale, a better assessment of the N-gap between crop N uptake and N supplied by N<sub>2</sub> fixation is needed.

In summary, three main critical points will be addressed in this review: (i) quantify seed production per kg of fixed N under different NDFA proportions, (ii) quantify N-gap and explore limitations of N<sub>2</sub> fixation for satisfying plant N demand, and (iii) calculate a partial N balance for soybean and determine its relationship with N<sub>2</sub> fixation and provide an overall value for the fixed N contribution to the “soybean N credit” or “soybean rotation effect”, excluding the potential contribution of N from BNF coming from the roots.

## DATABASE AND ANALYSIS

### Database

The database consisted of information previously collected from scientific literature over the past decades (Salvagiotti et al., 2008: 38 studies), but it also included 22 additional scientific studies, for a total of 60 datasets from the entire globe from 1955 through 2016 (Table 1). In all cases, the database comprised studies primarily focused on quantifying N<sub>2</sub> fixation in soybean as affected by multiple genotypes × environment × management practices combinations. Countries represented in the database included the United States (15 datasets, *n* = 201 observations), China (7, *n* = 91), Thailand (6, *n* = 92), Argentina (4, *n* = 113), Australia (4, *n* = 58), Brazil (4, *n* = 17), Japan (4, *n* = 28), Canada (2, *n* = 41), Austria (3, *n* = 22), India (2, *n* = 13), Nigeria (2, *n* = 15), France (1, *n* = 8), Indonesia (1, *n* = 2), Syria (1, *n* = 9), Kenya (1, *n* = 8), South Korea (1, *n* = 3), Zambia (1, *n* = 4), and Zimbabwe (1, *n* = 8). Data inclusion

followed criteria defined in previous review papers (Ciampitti and Vyn, 2012, 2013, 2014; Ciampitti and Prasad, 2016). Briefly, information must report data on seed yield at harvest and data at the end of the crop cycle (mainly R6.5–R7) of total plant N content, N<sub>2</sub> fixation (g m<sup>-2</sup>), NDFA (%), and seed N content. Multiple N<sub>2</sub> fixation determination techniques have been reported in different studies: (i) N difference method (8 studies), (ii) ureides determination in the xylem sap or stems (17 studies), (iii) <sup>15</sup>N dilution technique and <sup>15</sup>N natural abundance technique (34 studies), and (iv) acetylene reduction method (1 study); for complete citations of all techniques refer to Salvagiotti et al. (2008) and Unkovich and Pate (2000). The majority of data was retrieved from tables, some from equations, and a small proportion from digitized figures; if only NDFA was reported, total magnitude of N fixed was determined by multiplying the NDFA (%) contribution by its respective plant N content per unit area value. When BNF was estimated by the ureide or the acetylene reduction method, N<sub>2</sub> fixation was calculated by integrating measurements that were made multiple times over the season (i.e., there were not point-in-time measurements) (Peoples et al., 1989).

### Variables Evaluated

For the purpose of this review, total plant N content refers to the aboveground plant N content (stem, leaves, podwalls, and seed) determined close to maturity (as describe above, R6.5–R7), excluding belowground fraction. A similar concept was followed for the BNF variables, excluding any contribution from roots, because most of the studies did not provide this information. If not reported in the original publication, plant N content per unit area was determined by multiplying plant biomass and its respective N concentration (dry mass basis). Total N<sub>2</sub> fixation was reported in the research studies or calculated as the N<sub>2</sub> fixation (%) multiplied by its respective plant N content. For this review, the term N<sub>2</sub> fixation was utilized to refer the quantity of soybean N fixed at the end of the season (kg N ha<sup>-1</sup>), while the term NDFA (%) refer to the contribution of N<sub>2</sub> fixation as a proportion of plant N content, expressed in relative terms. Seed yield was reported on an area basis and adjusted to 13% moisture content.

The partial N balance was calculated as follows:

$$\text{Partial N balance} = \text{Fixed N in aboveground biomass} - \text{N seeds.}$$

Negative partial N balance indicates that the amount of N exported from harvesting soybean seeds is larger than N fixed by the crop, and thus a net “soil N depletion” may occur. Alternatively, a positive partial N balance portrays a situation where N<sub>2</sub> fixation contribution to the soybean exceeds seed N export, resulting in a net positive N budget. To consider the potential contribution of roots, an adjusted partial N balance was calculated by assuming that 24% of the total plant N content is located in the roots (Rochester et al., 1998).

### Database Analysis

For the entire database, a descriptive analysis was performed using the R function “hist” to prepare all histograms (R Development Core Team, 2009) and to estimate total number

Table 1. Number of study, country, experimental design, year of experimentation, BNF method, fertilizer N rates, and treatments evaluated for each different soybean study included in the meta-database. Other studies from Salvagioti et al. (2008)†.

No	Country	Author; Yr	Design	Years	BNF Method	N rate	Treatments evaluated
1	China	Yong et al., 2015	Split-plot	2012/13	<sup>15</sup> N natural abundance	3 (0, 180, 240 kg N ha <sup>-1</sup> )	Planting patterns × N rates × crop rotation
2	Syria	Al-Chammaa et al., 2014	Randomized Complete Block (RCB)	Not reported	<sup>15</sup> N isotopic dilution	20 kg N ha <sup>-1</sup> as labeled	Phosphorus levels × rates of sheep manure
3	Japan	Tewari et al., 2005	Not reported	2002	<sup>15</sup> N isotopic dilution	16 basal + 100 kg N ha <sup>-1</sup>	Deep placement of <sup>15</sup> N-labeled × inoculation
4	Argentina	Di Ciocco et al., 2008	RCB	2004/05	<sup>15</sup> N using three methods	10 kg N ha <sup>-1</sup> as labeled	BNF methods × tillage system
5	Nigeria	Sanginga et al., 1997	Split-/Split-split-plot	1994/95	<sup>15</sup> N isotopic dilution	20 kg N ha <sup>-1</sup> as labeled	Varieties × growth periods × inoculation
6	Nigeria	Sanginga et al., 2002	RCB	1996/97	<sup>15</sup> N isotopic dilution	20 kg N ha <sup>-1</sup> as labeled	<sup>15</sup> N labeling approach × varieties
7	Kenya	Kihara et al., 2011	Factorial	2007	<sup>15</sup> N isotopic dilution	4,6 kg N ha <sup>-1</sup> as labeled	Tillage × crop residue × cropping systems
8	India	Singh et al., 2014	RCB	1972/10	N balance method	10 different N rates	Nutrient management in soybean and wheat
9	South Korea	Park et al., 2005	RCB	2001/02	Ureides	0 or 3 kg N ha <sup>-1</sup>	Cropping systems × N levels × varieties
10	Japan	Shimada et al., 2012	Split-plot	2006/07	Ureides	36 kg N ha <sup>-1</sup> as dolomitic limestone	Water table × soybean varieties
11	USA	Salvagioti et al., 2009	RCB	2006/07	Ureides	0 or 180 kg N ha <sup>-1</sup> of different fertilizer sources	N fertilizer management
12	Zimbabwe	Zingore et al., 2008	RCB	2002/05	<sup>15</sup> N natural abundance	70 kg N ha <sup>-1</sup> as ammonium nitrate (maize)	Crop rotation × manure
13	Brazil	Alves et al., 2006	RCB	2000/02	<sup>15</sup> N natural abundance	No-N reported	Crop rotation
14	Brazil	Macedo and Miranda, 2001	RCB	1997/98	<sup>15</sup> N natural abundance	No-N reported	Tillage systems × crop rotation
15	Brazil	Neves et al., 1985	RCB	1983	Ureides	2 (0 or 60 kg N ha <sup>-1</sup> )	N rates × inoculation
16	Argentina	Collino et al., 2015	On-farm research	2004/11	<sup>15</sup> N natural abundance	No-N reported	On-farm research
17	Canada	Lynch and Smith, 1993	Completely Randomized	Not reported	N difference	No-N reported	Exposure to a low root-zone temperature
18	China	Xinmin et al., 1993	RCB	Not reported	<sup>15</sup> N isotopic dilution	No-N reported	Varieties and ability to fix N
19	China	Mengpei et al., 1986	Not reported	1982-1985	<sup>15</sup> N isotopic dilution	N check, and N-P-K combinations	Fertilizer rates × residue on yields and BNF
20	China	Jinfan et al., 1987	Not reported	1985	<sup>15</sup> N isotopic dilution	No-N reported	N fixation on three soils × soybean varieties
21	Argentina	Santachiara et al., 2017	RCB	2012/13	<sup>15</sup> N natural abundance	No-N reported	Soybean varieties (70 genotypes) × N fixation
22	USA	Tamagno and Ciampitti, 2017	RCB	2016	Ureides	0 or 112 kg N ha <sup>-1</sup>	N rates × timing × N fixation across several sites in the US Midwest

† From Salvagioti et al. 2008 (38 studies): Alvarez et al., 1995; Gan et al., 2002; Hughes and Herridge, 1989; Zotarelli, 2000; Bezdicek et al., 1978; Weber, 1966; Kucey et al., 1988a,b; Rennie et al., 1988; Amarger et al., 1979; Zapata et al., 1987; George et al., 1988; Kundu et al., 1996; Sisworo et al., 1990; Afza et al., 1987; Hardarson et al., 1984; Jefing et al., 1992; Guafa et al., 1993; Guffy et al., 1989; Peoples et al., 1995; Bergersen et al., 1992; Cassman et al., 1993; Leffel et al., 1992; Vasilas and Ham, 1985; Munyinda et al., 1988; Ham and Cadwell, 1978; Bhango and Albritton, 1976; Johnson et al., 1975; Ravuri and Hume, 1993; Vasilas and Fuhrman, 1993; Herridge, 1982; Zhang et al., 1986; Thies et al., 1995; Toomsan et al., 1995; Takahashi et al., 1991; Gan et al., 2002; Tewari et al., 2004; Israel and Burton, 1997.

Table 2. Descriptive statistics of the meta-database relative to soybean yield (adjusted to 13% moisture), Plant N content at the end of the season (dry basis), N contribution from biological N fixation ( $N_2$  fixation) at the end of the season in absolute values and expressed in relative terms, N derived from the atmosphere, NDFA %, in aboveground biomass.

Parameter	Unit	n	Mean	SD	Minimum	25% Q	Median	75% Q	Maximum
Seed yield	Mg ha <sup>-1</sup>	733	3.1	1.4	0.10	2.0	2.9	4.1	8.3
Plant N content	kg ha <sup>-1</sup>	733	245	108	7.0	162	228	331	538
$N_2$ fixation (all N)	kg ha <sup>-1</sup>	733	137	82	0.0	72	127	194	372
$N_2$ fixation (no N)	kg ha <sup>-1</sup>	473	142	78	0.0	83	130	194	372
NDFA % (all N†)	Unitless	733	56	21	0.0	44	59	72	98
NDFA % (no N‡)	Unitless	473	58	19	0.0	46	60	73	98

† All N refers to summary statistics for all data sets, including treatments with or without fertilizer-N.

‡ No N refers to summary statistics for the data sets without including any fertilizer-N.

Table 3. Descriptive statistics of the meta-database relative to the partial N balance, calculated as the N contribution  $N_2$  fixation minus the total N removed in the seed, seed yield (adjusted to 13% moisture), seed N content (dry basis),  $N_2$  fixation (absolute terms) at the end of the season (mainly R6.5 to R7 growth stage), N derived from the atmosphere, and NDFA (expressed in relative terms to total plant N content), for each NDFA group (as defined in Fig. 1A).

Parameter	Unit	n	Mean	SD	Minimum	25% Q	Median	75% Q	Maximum
Partial N balance (all N†)	kg ha <sup>-1</sup>	460	-47	55	-279	-76	-38	-11	111
Partial N balance <sub>roots</sub> ‡	kg ha <sup>-1</sup>	460	-13	66	-279	-46	-4.8	29	181
Seed yield	Mg ha <sup>-1</sup>	460	3.2	1.5	0.1	2.0	3.0	4.3	8.3
Seed N content	kg N ha <sup>-1</sup>	460	183	84	3.9	125	167	251	409
$N_2$ fixation	kg ha <sup>-1</sup>	460	136	86	0.0	66	127	198	372
NDFA%	Unitless	460	55	21	0.0	43	58	71	94
Partial N balance (no N§)	kg ha <sup>-1</sup>	190	-33	49	-159	-60	-35	-2	110
NDFA% Groups									
					0–44% NDFA				
Seed yield	Mg ha <sup>-1</sup>	122	2.9	1.2	0.5	1.8	2.8	3.4	6.1
Partial N balance	kg ha <sup>-1</sup>	122	-100	55.1	-279	-141	-106	-51.0	5.0
$N_2$ fixation	kg ha <sup>-1</sup>	122	62.5	49.2	0.0	25.4	50.5	89.0	197
					44–72% NDFA				
Seed yield	Mg ha <sup>-1</sup>	236	3.2	1.5	0.1	2.1	3.0	4.4	7.3
Partial N balance	kg ha <sup>-1</sup>	236	-38.5	38.7	-153	-59.0	-37.0	-14.0	74.0
$N_2$ fixation	kg ha <sup>-1</sup>	236	145	71.2	0.0	89.1	134	205	321
					72–98% NDFA				
Seed yield	Mg ha <sup>-1</sup>	102	3.6	1.7	0.4	2.4	3.3	4.7	8.3
Partial N balance	kg ha <sup>-1</sup>	102	-3.4	33.0	-80.0	-26.2	-5.0	8.2	110.0
$N_2$ fixation	kg ha <sup>-1</sup>	102	202	86	26.0	135	198	268	372

† All N refers to summary statistics for the partial N balance, including treatments with or without fertilizer-N.

‡ Partial N balance<sub>roots</sub>, calculations assuming an average N contribution from belowground biomass of 24% (Rochester et al., 1998).

§ No N refers to summary statistics for the partial N balance without including any fertilizer-N.

of observations ( $n$ ), mean, standard deviation (SD), minimum, 25–75% quartile, median, and maximum all variables collected in this paper (Tables 2 and 3). Two databases were formed with equal numbers of observations for (i) quantify N-gap including seed yield (Mg ha<sup>-1</sup>), plant N content (kg N ha<sup>-1</sup>),  $N_2$  fixation (kg N ha<sup>-1</sup>), and NDFA (%) (Database 1, Table 2); and (ii) estimate partial N balance comprising seed yield and  $N_2$  fixation (Database 2, Table 3). Database 1 is presented in Fig. 1, 2, and 3 (Table 2), while Database 2 was utilized for Fig. 4 (Table 3). Histograms were calculated for NDFA, seed yield, and plant N content (Fig. 1A, B, C), with Gaussian models fitted for each NDFA group (GraphPad Prism 6; Motulsky and Christopoulos, 2003). For the frequency distributions of NDFA, three groups were formed from < 25th, 25th–75th, and >75th: (i) low NDFA 0–44% ( $n = 187$ ); (ii) medium NDFA 44–72% ( $n = 372$ ); and (iii) high NDFA 72–98% ( $n = 174$ ) (Fig. 1A). The seed yield-to-plant N content (Fig. 2A) relationship was characterized by determining envelopes portraying the maximum and minimum

boundaries, 0.99 and 0.01 quantiles (Koenker, 2005). The linear components of the relationship between seed yield and  $N_2$  fixation were tested for each NDFA group (Fig. 2B) (F test, Mead et al., 1993) and compared with a global fit (GraphPad Prism 6, Motulsky and Christopoulos, 2003). These relationships were also shown by method of estimating  $N_2$  fixation to check for potential literature bias in this review (Supplementary Fig. S1). There was no clear trend of BNF method as related to low or high yields, plant N content, and/or  $N_2$  fixation values. Quantile regression was also utilized to estimate (Koenker, 2005) 10, 25, 50, 75, and 99 percentiles, for the relationship between  $N_2$  fixation and plant N content (Fig. 3A). In addition, a boundary function (0.99 quantile) was fitted to identify the expected highest fixed N values for a given level of plant N content. Linear and quadratic models were tested for each quantile regression and tested for equality of slopes among percentile lines. A similar approach was previously implemented by several researchers for identifying the highest yields for a given resource supply

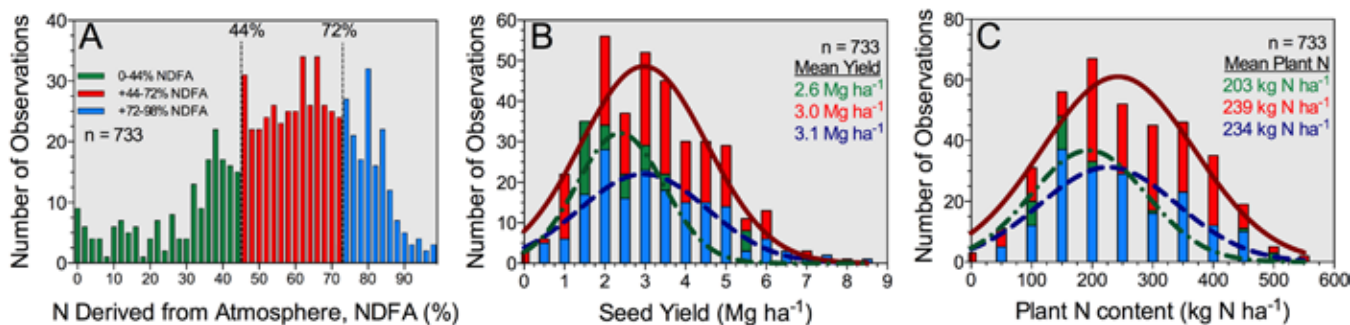


Fig. 1. Histogram of frequency for contribution of N from biological N fixation (BNF), expressed as the N derived from the atmosphere (NDA) (A), seed yield, expressed in  $\text{Mg ha}^{-1}$  and adjusted to 13% moisture content (B), and plant N content, expressed in  $\text{kg N ha}^{-1}$  in dry basis (C), all relative to the NDA groups: 0 to 44% NDA (green); +44 to 72% NDA (red), and +72 to 98% NDA (blue). Lines in (B) and (C) represent the Gaussian fit for each BNF group.

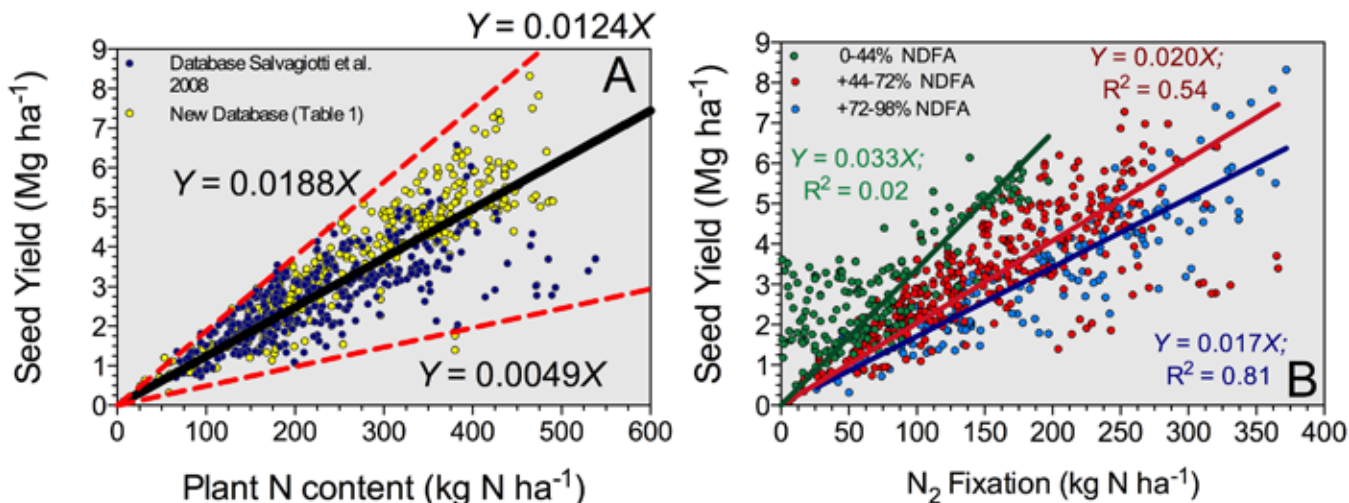


Fig. 2. Relationship between seed yield (adjusted to 13% moisture content) versus plant N content (dry basis) (A), and  $\text{N}_2$  fixation ( $\text{kg N ha}^{-1}$ ) for different NDA groups (B). In (A), the solid line is the average fit of the data, with a slope of  $0.0124 \text{ Mg grain kg}^{-1} \text{ N}$ . Dashed red lines show the boundaries of maximum N dilution (upper) and maximum N accumulation (lower). In (B), green, red, and blue lines depict the best-fitted line for the 0 to 44%, +44 to 72% and +72 to 98% NDA groups, respectively.

(Tittonell et al., 2008; Hochman et al., 2009). Changes in the  $\text{N}_2$  fixation to plant N content ratio for the boundary function were further studied via calculation of a segmental linear regression for values of plant N content above  $200 \text{ kg N ha}^{-1}$  and then  $370 \text{ kg N ha}^{-1}$ . The N-gap, calculated as the difference between crop N uptake and N supplied by  $\text{N}_2$  fixation for each quantile regression line was plotted against plant N content to quantify changes in response models for this relationship (Fig. 3B). The partial N balance was studied for understanding its statistical distribution (Fig. 4A), cumulative frequency (Fig. 4B), and final association with seed yield (Fig. 4C) as related to the three groups previously defined for NDA (Fig. 1A).

### SEED YIELD, PLANT N CONTENT, AND $\text{N}_2$ FIXATION

The analysis of the pooled data for yield and plant N content indicated a similar variation considering the 50% interquartile range (50% IQR, from 25–75% quartile), from 2.0 to  $4.1 \text{ Mg ha}^{-1}$  for seed yield and from 162 to  $331 \text{ kg N ha}^{-1}$  for plant N content (Table 2). Overall mean seed yield was  $3.1 \text{ Mg ha}^{-1}$  with yield distribution slightly skewed toward high values (Fig. 1B) and a maximum value of  $8.3 \text{ Mg ha}^{-1}$  (Table 2). Slightly lower mean seed yield and maximum value for yield were documented by Salvagiotti et al. (2008), which

were  $2.7 \text{ Mg ha}^{-1}$  and  $5.9 \text{ Mg ha}^{-1}$ , respectively. Therefore, this review increased the number of cases with high yields (Table 2). Plant N content at maturity averaged  $245 \text{ kg N ha}^{-1}$ , which also was slightly greater than the  $219 \text{ kg N ha}^{-1}$  mean value documented by Salvagiotti et al. (2008).

Mean fixed N was  $137 \text{ kg N ha}^{-1}$ , showing a maximum value of  $372 \text{ kg N ha}^{-1}$  (Table 2). In relative terms,  $\text{N}_2$  fixation (%) reached the maximum point at 98%. Overall, the NDA was 56% and presented a 50% IQR from 44 to 72% (Table 2). Under dryland soybean conditions, NDA was reported to be 50% (Unkovich and Pate, 2000). In a large study from 41 sites in Argentina, average NDA was 58%, with a 50% IQR from 46 to 71% (Collino et al., 2015). An overall NDA of 52% was synthesized by Salvagiotti et al. (2008), with a 50% IQR ranging from 36 to 69%. When the database did not comprise studies that applied fertilizer-N, fixed N increased to a mean value of  $142 \text{ kg N ha}^{-1}$  (from  $137 \text{ kg N ha}^{-1}$ ) and an overall NDA of 58% (from 56%) (Table 2). Thus, NDA for the BNF process could range from 50 to 60% for soybean systems around the globe.

The N internal efficiency (NIE, i.e., the slope of the yield-to-plant N content relationship) was  $0.0124 \text{ Mg kg}^{-1} \text{ N}$  (Fig. 2A). The new dataset provided in this review (yellow circles; Fig. 2A) added higher yields and associated plant N content levels as compared to Salvagiotti et al. (2008). Superior yield values required

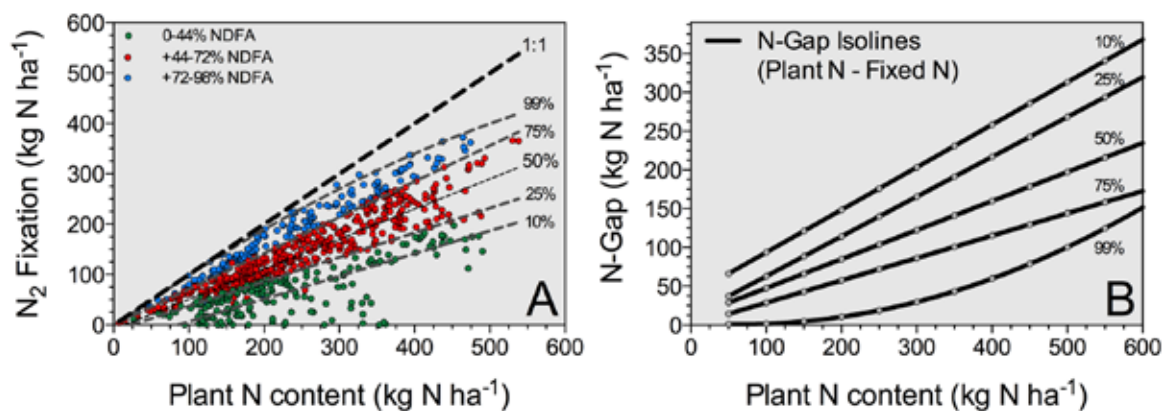


Fig. 3. Relationship between the contribution of  $N_2$  fixation and plant N content (both expressed in  $kg\ N\ ha^{-1}$ ) (A), and the N-gap (plant N– fixed N) relative to the plant N content (B). Quantile regression lines were fitted for the relationship in (A), representing NDFA isolines. For (A), the percentile 99 line (boundary function) adjusted was  $Y = -3.47 + 1.07X - 0.0005X^2$ . For (B), quantile regression lines, in (A), were used to calculate isolines of the difference between plant N content minus fixed N, herein term as N-gap isolines.

greater plant N content, but variation in this factor increased with yields; note the maximum dilution ( $0.0188\ Mg\ kg^{-1}\ N$ ) and accumulation ( $0.0049\ Mg\ kg^{-1}\ N$ ) boundary lines. Similar NIE and boundaries for the yield-to-N content relationship were previously documented by Salvagiotti et al. (2008). Then, for a constant  $300\ kg\ N\ ha^{-1}$ , seed yield is expected to be 1.5, 3.6, and  $5.6\ Mg\ ha^{-1}$  for the maximum N accumulation (minimum boundary), median (average), and N dilution (maximum boundary) curves, respectively (Fig. 2A). The latter portrays the differential internal N use efficiency in soybean and the influence of other factors namely environment, management, and cultivar. It seems from the distribution of data points in Fig. 2A that there is not a clear plateauing in seed yield at high levels of N uptake as was observed in other graminaceous crops like corn (*Zea mays* L.) (Setiyono et al., 2010), rice (*Oryza sativa* L.) (Witt et al., 1999), or wheat (*Triticum aestivum* L.) (Liu et al., 2006), where NIE decreases as N content increases, suggesting that, in soybean, NIE remains constant even at high levels of seed yield (above  $7\ Mg\ ha^{-1}$ ). This different behavior between cereals and soybean (and maybe other legumes) might be related to the fact that total plant N content in the former will be limited by soil plus fertilizer N sources, while in the latter the BNF process is mostly satisfying many times when plant N demand is not provided by soil. In addition, larger N concentration in vegetative and reproductive organs in legumes (e.g., soybeans) relative to cereals (e.g., corn) (Tamagno et al., 2017; Ciampitti and Vyn, 2011) could be another source for explaining the differences in NIE. When compared with other legumes (pigeon pea, *Cajanus cajan* L.; and peanut, *Arachis hypogaea* L.), soybeans presented larger losses of N in leaves (finishing with lower N concentrations), thus presenting differential N dynamics (Devries et al., 1989). In summary, more data beyond  $500\ kg\ N\ ha^{-1}$  are needed to confirm if NIE will remain in a constant trend for soybeans.

Seed yield and  $N_2$  fixation were linearly related in each of the NDFA groups (Fig. 2B). Overall, the yield-to- $N_2$  fixation relationship presented a larger variability relative to the yield-to-N content association. For the low NDFA group (green circles; Fig. 2B),  $N_2$  fixation presented a maximum value of  $197\ kg\ N\ ha^{-1}$  and seed yield of  $6.1\ Mg\ ha^{-1}$ . In this group, when  $N_2$  fixation was below  $50\ kg\ N\ ha^{-1}$  and seed yield ranging from  $>0$  to  $<4\ Mg\ ha^{-1}$ , lower yield was obtained in parallel with

low NDFA below 20% (Fig. 2B). This group was characterized by studies that included fertilizer N application that averaged  $106\ kg\ N\ ha^{-1}$  across all treatments, with high nitrate values around the nodulation zone or presence of low soil pH values, ineffective rhizobia, or biotic stresses affecting the nodulation process (Salvagiotti et al., 2008). This group showed low NDFA and N content more dependent on soil (or fertilizer) N. Changes in the slope of the yield-to- $N_2$  fixation relationship were primarily explained by NDFA, with higher slope ( $0.020X$  or  $50\ kg\ N\ Mg^{-1}$ ) for the medium group relative to the high NDFA group (slope  $0.017X$  or  $59\ kg\ N\ Mg^{-1}$ ) (Fig. 2B). The reduction of more than 50% on the yield-to- $N_2$  fixation slope from the low to the high NDFA group reflects a decrease in the degree of dependency to other external N sources (soil + fertilizer; Fig. 2B) for satisfying N demand in the latter group. When comparing the effects of N fertilization in this meta-analysis (i.e., grouping  $0\ kg\ N\ ha^{-1}$  vs.  $>10\ kg\ N\ ha^{-1}$ ) (data not shown), the slope for the yield-to- $N_2$  fixation relationship was not statistically different, between the N-fertilized and the zero-N fertilization group, averaging  $0.0192X$  (i.e.,  $52\ kg\ N\ Mg^{-1}$ ).

When fertilizer N was applied at rates above  $100\ kg\ N\ ha^{-1}$ , this slope was slightly modified, lowering  $N_2$  fixation per unit of N uptake when considering each sub-database (slope =  $42\ kg\ N\ Mg^{-1}$ ;  $n = 259$ , N fertilizer rate  $110\ kg\ N\ ha^{-1}$ , seed yield  $3.0\ Mg\ ha^{-1}$ , and  $N_2$  fixation  $129\ kg\ N\ ha^{-1}$ ; vs. slope =  $48\ kg\ N\ Mg^{-1}$ ;  $n = 292$ , N fertilization  $0\ kg\ N\ ha^{-1}$ , seed yield  $2.8\ Mg\ ha^{-1}$ , and  $N_2$  fixation  $132\ kg\ N\ ha^{-1}$ ). These results suggest that application of N fertilizer partially inhibited  $N_2$  fixation decreasing the efficiency of N fixed per unit of yield, but further testing at multiple sites should confirm this research outcome. Several estimations of “energy costs” comparing soil N uptake and N fixed biologically at the cellular level are elusive for answering the question whether BNF represents a significant cost for the plant (Schubert, 1982; Salsac et al., 1984; Andrews et al., 2009). Generally, several assumptions are made for this type of analysis, and different values may come up depending on processes taken into account in the calculations. Schubert (1982) estimated that the total cost for both, BNF and inorganic N uptake are 92 ATP. However, Andrews et al. (2009) estimated that energy costs for BNF is 5 to 7% greater than for nitrate plus ammonium uptake.

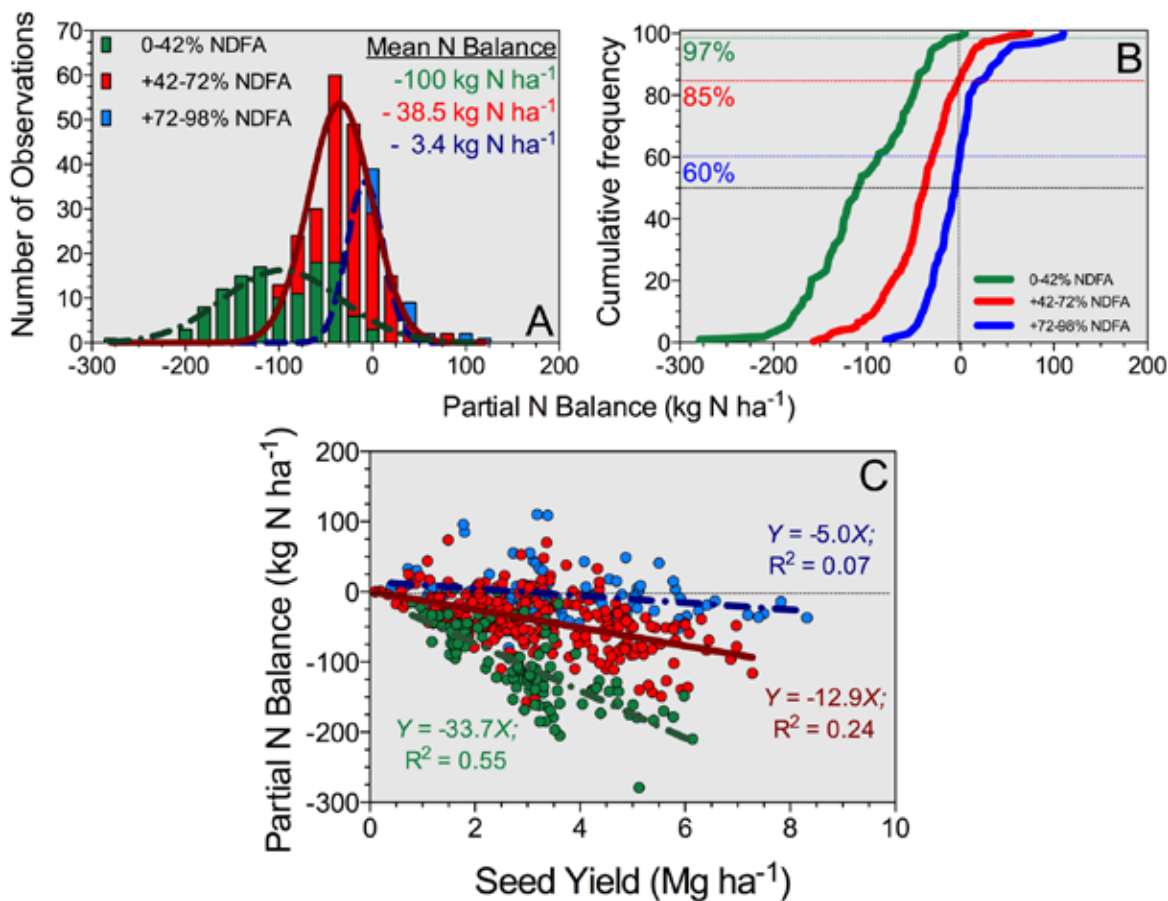


Fig. 4. Histogram of frequency for partial N balance (A), cumulative distribution frequency for partial N balance (B), and relationship between partial N balance and seed yield adjusted to 13% moisture content (C). Green, red, and blue lines depict the best-fitted line for the 0–44%, +44–72%, and +72–98% NDFa groups, respectively. Lines in (A) represent the Gaussian fit for each NDFa group.

### N-GAP AND MAXIMUM N<sub>2</sub> FIXATION

A relationship between N<sub>2</sub> fixation and plant N content was established to provide a better assessment of the N-gap that is plant N content minus N<sub>2</sub> fixation (Fig. 3A). Linear models better explained the relationship for percentiles 10 to 80 (percentile 80 not shown in the figure), but they presented different slopes among them. Regression lines from percentiles 10 to 25 shared an equal slope of 0.47X, but slopes increased less than proportionally until percentile 80 (i.e., percentile 50 and 75 showed a slope of 0.62X and 0.71X, respectively). Above percentile 80, a quadratic model better fit the relationship without changing the linear term of the equation between 90 and 99 percentiles (0.80X linear term) (Fig. 3A). Analyzing percentile 25 (low NDFa levels), when plant N content was 50 kg N ha<sup>-1</sup>, the expected N-gap was 37 kg N ha<sup>-1</sup> and it rose to 183 kg N ha<sup>-1</sup> when plant N content was 400 kg N ha<sup>-1</sup> (i.e., an eight-fold increase in plant N demand was accompanied by a five-fold increase in the N-gap). However, when the NDFa increased (i.e., 75 quantile regression line), the N-gap was reduced close to 15 kg N ha<sup>-1</sup> when plant N content was 50 kg N ha<sup>-1</sup> and rose to 115 kg N ha<sup>-1</sup> when plant N uptake was 400 kg N ha<sup>-1</sup> (i.e., an eight-fold increase for both N content and N-gap) (Fig. 3B).

Many factors affect the N<sub>2</sub> fixation, including genotype × environment × management practices interactions. The main issue to be addressed in this review is to quantify the N<sub>2</sub> fixation maximum capacity. The boundary function (i.e., percentile

99 in Fig. 3A) represents the maximum attainable N<sub>2</sub> fixation contribution at each level of plant N content. As a first step the slope for the percentile 99 function was compared to the 1:1 line, and presented equal slope until 200 kg N ha<sup>-1</sup> (F test; Mead et al., 1993). The slope of  $-0.001$  kg N fixed kg<sup>-1</sup> N uptake portrays that as plant N content increases N<sub>2</sub> fixation decreases. As a second step, a segmental lineal regression was fitted for all plant N content values above 200 kg N ha<sup>-1</sup> to calculate the inflection point in which the N fixed to N uptake ratio changes, this plant N content value was obtained at 370 kg N ha<sup>-1</sup>. Therefore, for the N-gap analysis, after 200 kg N ha<sup>-1</sup>, the size of the N-gap increased at an estimated linear rate of 0.22 kg N-gap kg<sup>-1</sup> N content, and then drastically changing at 370 kg N ha<sup>-1</sup> with a slope of 0.46 kg N-gap kg<sup>-1</sup> N content, more than a twofold change (Fig. 3B). Thus, larger plant N content implies a greater dependency on external N sources to achieve higher yields. Maximum N<sub>2</sub> fixation values ranging from 337 to 372 kg N ha<sup>-1</sup> (with a concomitant NDFa from 68% to 86%) were gathered from Herridge (1982), Tewari et al. (2004), Santachiara et al. (2017), and Tamagno and Ciampitti (2017). In our dataset, only 3% ( $n = 23$ ) showed seed yields above 6 Mg ha<sup>-1</sup>, with an average NDFa of 67% (ranging from 33% to 81%), representing a mean N<sub>2</sub> fixation contribution of 279 kg N ha<sup>-1</sup> and with an N-gap of 137 kg N ha<sup>-1</sup>. These results clearly show that soybean crops with a high contribution from BNF cannot attain high yielding (>7 Mg ha<sup>-1</sup>) and potential N

uptake. Very few observations ( $n = 6$ ) attained yield levels above  $7.5 \text{ Mg ha}^{-1}$  with an NDFA of 74%, total  $\text{N}_2$  fixation contribution of  $331 \text{ kg N ha}^{-1}$  and a N-gap of  $116 \text{ kg N ha}^{-1}$ . The latter outcomes are the first ones to concomitantly portray high-yielding soybean within the high NDFA group (+72%). In summary, N-gap of soybean that received a high proportion of N from BNF increased at a faster rate when overpassing  $370 \text{ kg N ha}^{-1}$  (Fig. 3B), with overall mean and maximum yield environments from  $4.6$  to  $+7 \text{ Mg ha}^{-1}$  (Fig. 2A), respectively.

## PARTIAL N BALANCE AND $\text{N}_2$ FIXATION

The partial N balance (excluding BNF contribution from roots) presented an overall mean of  $-47 \text{ kg N ha}^{-1}$ , with  $-75$  to  $-11 \text{ kg N ha}^{-1}$  values for the 50% IQR (Table 3). For the dataset collected, only 17% of all observations showed a positive partial N balance, averaging  $22 \text{ kg N ha}^{-1}$ , while more than 80% of the data showed N balance of  $-61 \text{ kg N ha}^{-1}$  (Fig. 4A). Based on the NDFA groups (Fig. 1A), the partial N balance for the low BNF group presented an average of  $-100 \text{ kg N ha}^{-1}$  with a 50% IQR from  $-141$  to  $-51 \text{ kg N ha}^{-1}$ , an overall yield of  $2.9 \text{ Mg ha}^{-1}$ , and  $\text{N}_2$  fixation of  $62.5 \text{ kg N ha}^{-1}$  (Table 3). The medium NDFA group presented an average partial N balance of  $-38.5 \text{ kg N ha}^{-1}$  with a 50% IQR from  $-59$  to  $-14 \text{ kg N ha}^{-1}$ , a yield of  $3.2 \text{ Mg ha}^{-1}$ , and  $\text{N}_2$  fixation of  $145 \text{ kg N ha}^{-1}$ . The high NDFA group presented an average of  $-3.4 \text{ kg N ha}^{-1}$  with a 50% IQR from  $-26$  to  $8 \text{ kg N ha}^{-1}$ , with an overall yield of  $3.6 \text{ Mg ha}^{-1}$ , and  $\text{N}_2$  fixation of  $202 \text{ kg N ha}^{-1}$  (Table 3). The partial N balance for the high NDFA group presented a distribution centered around zero but with heavy tails toward both negative and positive values (Fig. 4A).

Cumulative frequencies for the partial N balance for each NDFA group are presented in Fig. 4B. For the low NDFA only 3% of the data ( $n = 4$ ) presented positive partial N balance with 97% portraying a negative balance. The proportion of observations with positive partial N balance increased in the medium NDFA group, reaching 15% of all the datasets ( $n = 35$ ). Lastly, the high NDFA group presented 40% of all observations with positive partial N balance ( $n = 41$ ). Potential sources of error for the partial N balance are lack of accounting for potential N loss via leaf drop and the contribution of belowground parts. In the former case, N harvest index (used as an indicator of differences in N leaf drop) of the dataset evaluated was 0.70 units, ranging the 50% IQR from 0.60 to 0.80. Similar variation was recently documented by Tamagno et al. (2017) in a synthesis-analysis with an overall mean of 0.75 units. Then, this slight lower NHI difference obtained from two independent data sets allowed us to estimate N balances using the current data set. Nonetheless, N from dropped leaves is a potential source of error that should be properly estimated in future N balance studies for the soybean crop. Regarding the second source of error, i.e., N contribution from belowground parts, Table 3 shows a new balance including an additional 24% of N that is contributed by roots (Rochester et al., 1998), which still resulted in a  $-13 \text{ kg N ha}^{-1}$  balance (Table 3). Notwithstanding the adjustment of the partial N balance summing up this additional N contribution from BNF at R7 (Rochester et al., 1998), this method may still underestimate the total root N contribution because N losses from roots and nodules occur during the growing season (Brophy and Heichel, 1989; Ofosu-Budu et al., 1990; Ta et al., 1986). For instance,

Rochester et al. (1998) determined that N content in belowground components at R7 represents approximately 60% of N uptake in R5. However, lower NDFA contribution from roots, ranging from 1 to 9%, was recently documented by Gelfand and Robertson (2015). For example, root NDFA contribution could range, on average, from 13 (9% NDFA) to 34 (24% NDFA)  $\text{kg N ha}^{-1}$  when considering the estimations from Gelfand and Robertson (2015) and Rochester et al. (1998), respectively. As clarified by Anglade et al. (2015), N derived from rhizodeposits are not well contained in a defined physical structure and root N contribution from BNF for all roots, more precisely thinner roots, is very challenging. Variations in root estimates presented in the research literature (Rochester et al., 1998; Gelfand and Robertson, 2015) might as well come from different root sampling techniques, variations in sampling depth, and lack of complete retrieval of in-field N rhizodeposition from thinner roots. It is evident that, after reviewing the scientific literature summarized by Salvagiotti et al. (2008) and considering this current review, more efforts should be focused on collecting data concerning the contribution of roots to obtain a more precise quantification of BNF impact on the partial N balance. In addition, measuring N gains in above- and belowground plant fractions due to BNF are needed, but also, in parallel, monitoring N losses (e.g., including N-metabolites via root excretion) are required.

## SOYBEAN N CREDIT

For soybean N-credit, commonly utilized in US maize-soybean systems for making N-fertilizer recommendations in maize, it could be hypothesized that this N-credit is entirely dependent on soil N mineralization of soybean residues with low C to N ratio (Bundy et al., 1993; Wu et al., 1998; Gentry et al., 2001; 2013). Green and Blackmer (1995) suggested that the N-credit, when sowing corn after soybean as compared with corn as a previous crop, was due to a larger N immobilization in the latter case. In addition, Maloney et al. (1999) and Bergerou et al. (2004) stated that the BNF process plays a minor role in the positive effect of soybean in a maize-soybean rotation, the so-called "soybean rotation effect". An alternative situation could be a transfer of soil N to the following crops, since less soil N is removed when soybean is in the rotation because it uses N derived from BNF, the so-called "N sparing effect" (Chalk, 1998). Even when excluding fertilizer N observations ( $n = 190$ ), the partial N balance still presented an overall  $-33 \text{ kg N ha}^{-1}$  with a very similar data distribution (relative when all fertilizer N points were considered) (50% IQR from  $-59$  to  $-2 \text{ kg N ha}^{-1}$ ) (Table 3). In any case, it seems that there likely would be a net gain of the partial N balance in the rotation system from BNF, and it may occur at both medium and high BNF groups (Fig. 4B), contributing to the "soybean rotation effect", but there is almost no contribution of the BNF process for the low BNF group (i.e., NDFA below 42%). Seed yield was the main factor driving changes in the partial N balance for the low NDFA group (Fig. 4C), with a more negative N balance as yields increased, showing a decrease of  $33.7 \text{ kg}$  in the N balance per  $\text{Mg}$  of yield. However, this relationship became weaker (i.e., low  $R^2$ ) as the NDFA level improved (Fig. 4C). The latter highlights the complexity of N dynamics. A more comprehensive approach looking at losses via greenhouse gases or N leaching (e.g.,  $\text{N}_2\text{O}$ ; Yang and Cai, 2005; Ciampitti et al., 2008; Itakura et al., 2013; Uchida and Akiyama, 2013) and root



excretion (Brophy and Heichel, 1989; Ofosu-Budu et al., 1990; Ta et al., 1986), and also N gains or inputs from atmospheric deposition and irrigation water, will improve knowledge and assist in identifying critical components for more precisely estimating the N budget in cropping systems dominated by soybeans.

## OUTCOMES AND FUTURE RESEARCH PRIORITIES

The most noteworthy outcomes of this review are that (i) as the contribution of NDFA increased, seed production per N<sub>2</sub> fixation decreased (from 0.033 to 0.017 Mg yield kg<sup>-1</sup> N from low, 28%, to high, 80%, NDFA; Fig. 1B); (ii) the N-gap increased greatly when NDFA values were above 80% and after a plant N content was above 370 kg N ha<sup>-1</sup>; (iii) when excluding roots, the partial N balance calculation revealed negative values across all the NDFA levels; (iv) the partial N balance was related to N<sub>2</sub> fixation, with positive balance most likely to occur for 40% ( $n = 41$  points), 15% ( $n = 35$ ), and 3% ( $n = 4$ ) of the database for the high (above 72% N<sub>2</sub> NDFA), medium (44 to 72% NDFA), and low BNF (below 44% NDFA) groups, respectively; (v) seed yield was stronger (greater  $R^2$ ) related to negative partial N balance only for the low NDFA group, with no clear trend for the medium and high NDFA groups; and lastly, (vi) the quantity of N contributed from BNF seems negligible, by itself, to be considered as a “soybean N credit” in a maize–soybean cropping rotation, primarily for the low and medium BNF groups. Under this scenario, the apparent N contribution from soybean seems to be primarily related to a greater soil N supply from a more positive N mineralization/immobilization balance in soybean–corn cropping systems.

The observed rise in the N-gap in high-yielding conditions, even with high BNF, suggests the need of having an additional source for supplying N to the crops. This provision has to come primarily from highly efficient *Rhizobium* strains adapted to environments with high plant N demand. However, the development of strategies that supply N at low rates during the cycle (especially during the seed-filling period), reducing the negative impact of soil nitrate concentration on BNF, seems to be a likely solution. These alternatives may involve the inclusion of legume cover crops in the rotation or the use of slow release N fertilizers. Nonetheless, both approaches should evaluate further the impact on soybean productivity, N budget at the crop and system level, and on the environment.

Two research priorities were identified from this review. The first priority should focus on exploring a more holistic study of the N cycling within soybean, first by including a better understanding of the BNF contribution from different plant parts, specifically including roots. Sampling methods and timing within the crop cycle are crucial factors that will affect estimation of root contribution to BNF. The second priority should be to pursue a better understanding of the soil × plant × rhizobia interactions on plant N processes (N remobilization, BNF, and N uptake) in high-yielding soybean systems, especially during the seed filling period, and their contribution to yield and/or seed protein formation process, focusing on the relative importance of contemporaneous and remobilized N. This should be complemented with well calibrated simulation crop models, because it is difficult to have, at the present, field studies that determine N derived from BNF in soybeans that yield more than 7 Mg ha<sup>-1</sup>.

## SUPPLEMENTARY MATERIAL

Supplementary Fig. S1. Relationship between seed yield (adjusted to 13% moisture content) versus N<sub>2</sub> fixation (dry basis, kg N ha<sup>-1</sup>) (panel A), and N<sub>2</sub> fixation (kg N ha<sup>-1</sup>) versus plant N content (dry basis, kg N ha<sup>-1</sup>) (panel B) for all N<sub>2</sub> fixation methods gathered in this review paper.

## ACKNOWLEDGMENTS

Mr. S. Tamagno, G. Balboa, L. Moro Rosso, and Mrs. D. Hansel are acknowledged for their assistance in collecting soybean studies in the last decades. This study was supported by the International Plant Nutrition Institute, (IPNI, Project GBL 62), K-State Research and Extension (KSRE), and INTA Oliveros. Dr. M.B. Kirkham is thanked for her helpful comments provided on a earlier version of this manuscript. This is contribution no. 17-388-J from the Kansas Agricultural Experiment Station.

## REFERENCES

- Afza, R., G. Hardarson, F. Zapata, and S.K.A. Danso. 1987. Effects of delayed soil and foliar N fertilization on yield and N<sub>2</sub> fixation of soybean. *Plant Soil* 97:361–368. doi:10.1007/BF02383226
- Anglade, J., G. Billen, and J. Garnier. 2015. Relationships for estimation N<sub>2</sub> fixation in legumes: Incidence for N balance of legume-based cropping systems in Europe. *Ecosphere* 6:1–24. doi:10.1890/ES14-00353.1
- Amarger, N., A. Mariotti, F. Mariotti, J.C. Durr, C. Bourguignon, and B. Lagacherie. 1979. Estimate of symbiotically fixed nitrogen in field grown soybeans using variations in <sup>15</sup>N natural abundance. *Plant Soil* 52:269–280. doi:10.1007/BF02184565
- Andrews, M., P.J. Lea, J.A. Raven, and R.A. Azevedo. 2009. Nitrogen use efficiency. 3. Nitrogen fixation: Genes and costs. *Ann. Appl. Biol.* 155:1–13. doi:10.1111/j.1744-7348.2009.00338.x
- Al-Chammaa, M., F. Al-Ain, and K. Khalifa. 2014. Growth and nitrogen fixation in soybean as affected by phosphorus fertilizer and sheep manure using <sup>15</sup>N isotopic dilution. *Commun. Soil Sci. Plant Anal.* 45:487–497. doi:10.1080/00103624.2013.863908
- Alvarez, R., J.H. Lemcoff, and A.H. Merzari. 1995. Nitrogen balance in a soil cultivated with soybeans. *Cienc. Suelo* 13:38–40.
- Alves, B.J.R., R.M. Boddey, and S. Urquiaga. 2003. The success of BNF in soybean in Brazil. *Plant Soil* 252:1–9. doi:10.1023/A:1024191913296
- Alves, B.J.R., L. Zotarelli, F. Marques Fernandes, J.C. Heckler, R.A. Tavares De Macedo, R.M. Boddey, C.P. Jantalia, and S. Urquiaga. 2006. Fixação biológica de nitrogênio e fertilizantes nitrogenados no balanço de nitrogênio em soja, milho e algodão. *Pesqui. Agropecu. Bras.* 41:449–456. doi:10.1590/S0100-204X2006000300011
- Balboa, G.R., V.O. Sadras, and I.A. Ciampitti. 2018. Shifts in soybean yield, nutrient uptake, and nutrient stoichiometry: A historical synthesis-analysis. *Crop Sci.* 58:43–54. doi:10.2135/cropsci2017.06.0349
- Bhango, M.S., and D.J. Albritton. 1976. Nodulating and non-nodulating Lee soybean isolines response to applied nitrogen. *Agron. J.* 68:642–645. doi:10.2134/agronj1976.00021962006800040027x
- Bergerou, J.A., L.E. Gentry, M.B. David, and F.E. Below. 2004. Role of N<sub>2</sub> fixation in the soybean N credit in maize production. *Plant Soil* 262:383–394. doi:10.1023/B:PLSO.0000037057.35160.ec
- Bergersen, F.J., G.L. Turner, R.R. Gault, M.B. Peoples, L.J. Morthorpe, and J. Brockwell. 1992. Contributions of nitrogen in soybean crop residues to subsequent crops and to soils. *Aust. J. Agric. Res.* 43:155–169. doi:10.1071/AR9920155
- Bezdicke, D.F., D.W. Evans, B. Abede, and R.E. Witters. 1978. Evaluation of peat and granular inoculum for soybean yield and N fixation under irrigation. *Agron. J.* 70:865–868. doi:10.2134/agronj1978.0021962007000050037x

- Brophy, L.S., and G.H. Heichel. 1989. Nitrogen release from roots of alfalfa and soybean grown in sand culture. *Plant Soil* 116:77–84.
- Bundy, L.G., T.W. Andraski, and R.P. Wolkowski. 1993. Nitrogen credits in soybean–corn sequences on three soils. *Agron. J.* 85:1061–1067. doi:10.2134/agronj1993.00021962008500050020x
- Cassman, K.G., P.W. Singleton, and B.A. Linquist. 1993. Input/output analysis of the cumulative soybean response to phosphorus on an Ultisol. *Field Crops Res.* 34:23–36. doi:10.1016/0378-4290(93)90108-Y
- Chalk, P.M. 1998. Dynamics of biologically fixed N in legume-cereal rotations: A review. *Aust. J. Agric. Res.* 49:303–316. doi:10.1071/A97013
- Ciampitti, I.A., E.A. Ciarlo, and M.E. Conti. 2008. Nitrous oxide emissions from soil during soybean (*Glycine max* (L.) Merrill) crop phenological stages and stubble decomposition period. *Biol. Fertil. Soils* 44:581–588. doi:10.1007/s00374-007-0241-7
- Ciampitti, I.A., and P.V.V. Prasad. 2016. Historical synthesis-analysis of changes in grain nitrogen dynamics in sorghum. *Front. Plant Sci.* 7:275. doi:10.3389/fpls.2016.00275
- Ciampitti, I.A., and T.J. Vyn. 2011. A comprehensive study of plant density consequences on nitrogen uptake dynamics of maize plants from vegetative to reproductive stages. *Field Crop. Res.* 121:2–18. doi:10.1016/j.fcr.2010.10.009.fcr.2010.10.009
- Ciampitti, I.A., and T.J. Vyn. 2012. Physiological perspectives of changes over time in maize yield dependency on nitrogen uptake and associated nitrogen efficiencies: A review. *Field Crops Res.* 133:48–67. doi:10.1016/j.fcr.2012.03.008
- Ciampitti, I.A., and T.J. Vyn. 2013. Grain nitrogen source changes over time in maize: A review. *Crop Sci.* 53:366–377. doi:10.2135/cropsci2012.07.0439
- Ciampitti, I.A., and T.J. Vyn. 2014. Understanding global and historical nutrient use efficiencies for closing maize yield gaps. *Agron. J.* 106:2107-2117.
- Collino, D.J., F. Salvaggiotti, A. Peticari, C. Piccinetti, G. Ovando, S. Urquiaga, and R.W. Racca. 2015. Biological nitrogen fixation in soybean in Argentina: Relationships with crop, soil, and meteorological factors. *Plant Soil* 392:239–252. doi:10.1007/s11104-015-2459-8
- Devries, J.D., J.M. Bennett, K.J. Boote, S.L. Albrecht, and C.E. Maliro. 1989. Nitrogen accumulation and partitioning by three grain legumes in response to soil water deficits. *Field Crops Res.* 22:33–44. doi:10.1016/0378-4290(89)90087-7
- Di Ciocco, C., C. Coviella, E. Peñon, M. Diaz-Zorita, and S. López. 2008. Biological fixation of nitrogen and N balance in soybean crops in the pampas region. *Span. J. Agric. Res.* 6:114–119. doi:10.5424/sjar/2008061-5259
- Gan, Y., I. Stulen, F. Posthumus, H.V. Keulen, and P.J.C. Kuiper. 2002. Effects of N management on growth, N<sub>2</sub> fixation and yield of soybean. *Nutr. Cycling Agroecosyst.* 62:163–174. doi:10.1023/A:1015528132642
- Gelfand, I., and G.P. Robertson. 2015. A reassessment of the contribution of soybean biological nitrogen fixation to reactive N in the environment. *Biogeochemistry* 123:175–184. doi:10.1007/s10533-014-0061-4
- Gentry, L.E., F.E. Below, M.B. David, and J.A. Bergerou. 2001. Source of the soybean N credit in corn production. *Plant Soil* 236:175–184. doi:10.1023/A:1012707617126
- Gentry, L.E., S.S. Snapp, R.F. Price, and L.F. Gentry. 2013. Apparent red clover nitrogen credit to corn: Evaluating cover crop introduction. *Agron. J.* 105:1658–1664. doi:10.2134/agronj2013.0089
- George, T., P.W. Singleton, and B. Ben. 1988. Yield, soil nitrogen uptake, and nitrogen fixation by soybean from four maturity groups grown at three elevations. *Agron. J.* 80:563–567. doi:10.2134/agronj1988.00021962008000040004x
- Green, C.J., and A.M. Blackmer. 1995. Residue decomposition effects on nitrogen availability to corn following corn or soybean. *Soil Sci. Soc. Am. J.* 59:1065–1070. doi:10.2136/sssaj1995.03615995005900040016x
- Guafo, W., M.B. Peoples, D.F. Herridge, and B. Rerkasem. 1993. Nitrogen fixation, growth and yield of soybean grown under saturated soil culture and conventional irrigation. *Field Crops Res.* 32:257–268. doi:10.1016/0378-4290(93)90035-L
- Guffy, R.D., R.M. Vanden Heuvel, B.L. Vasilas, R.L. Nelson, M.A. Frobish, and J.D. Hesketh. 1989. Evaluation of the N<sub>2</sub> fixation capacity of four soybean genotypes by several methods. *Soil Biol. Biochem.* 21:339–342. doi:10.1016/0038-0717(89)90140-5
- Ham, G.E., and A.C. Caldwell. 1978. Fertilizer placement effects on soybean yield, N<sub>2</sub> fixation, and <sup>32</sup>P uptake. *Agron. J.* 70:779–783. doi:10.2134/agronj1978.00021962007000050020x
- Hardarson, G., F. Zapata, and S.K.A. Danso. 1984. Effect of plant genotype and nitrogen fertilizer on symbiotic nitrogen fixation by soybean cultivars. *Plant Soil* 82:397–405. doi:10.1007/BF02184277
- Herridge, D.F. 1982. Use of the ureide technique to describe the nitrogen economy of field-grown soybeans. *Plant Physiol.* 70:7–11. doi:10.1104/pp.70.1.7
- Hochman, Z., D. Holzworth, and J.R. Hunt. 2009. Potential to improve on-farm wheat yield and WUE in Australia. *Crop Pasture Sci.* 60:708–716. doi:10.1071/CP09064
- Hughes, R.M., and D.F. Herridge. 1989. Effect of tillage on yield, nodulation and nitrogen fixation of soybean in far north-coastal New South Wales. *Aust. J. Exp. Agric.* 29:671–677. doi:10.1071/EA9890671
- Itakura, M., Y. Uchida, H. Akiyama, Y.T. Hoshino, Y. Shimomura, S. Morimoto, K. Tago, Y. Wang, C. Hayakawa, Y. Uetake, C. Sánchez, S. Eda, M. Hayatsu, and K. Minamisawa. 2013. Mitigation of nitrous oxide emissions from soils by *Bradyrhizobium japonicum* inoculation. *Nat. Clim. Chang.* 3:208–212. doi:10.1038/nclimate1734
- Israel, D.W., and J.W. Burton. 1997. Nitrogen nutrition of soybean grown in Coastal Plain soils of North Carolina. *Technic. Bull.* 310. North Carolina Agricultural Research Service, North Carolina State University, Raleigh, NC.
- Jefing, Y., D.F. Herridge, M.B. Peoples, and B. Rerkasem. 1992. Effects of N fertilization on N<sub>2</sub> fixation and N balances of soybean grown after lowland rice. *Plant Soil* 147:235–242. doi:10.1007/BF00029075
- Jinfan, G., W. Qing, H. Zhongyan, Z. Hong, Z. Guibin, Z. Guizhi, W. Xiaoming, and X. Bao. 1987. Study of the symbiotic fixation nitrogen of soybean by nitrogen-15. *Soybean Sci.* 6:55–61.
- Johnson, J.W., L.F. Welch, and L.T. Kurtz. 1975. Environmental implications of N fixation by soybeans. *J. Environ. Qual.* 4:303–306. doi:10.2134/jeq1975.00472425000400030003x
- Kihara, J., C. Martius, A. Bationo, and P.L.G. Vieck. 2011. Effects of tillage and crop residue application on soybean nitrogen fixation in a tropical ferrasol. *Agriculture* 1:22–37. doi:10.3390/agriculture1010022
- Koenker, R. 2005. *Quantile regression*. University of Cambridge Press, Cambridge, UK.
- Koester, R.P., J.A. Skoneczka, T.R. Cary, B.W. Diers, and E.A. Ainsworth. 2014. Historical gains in soybean (*Glycine max* Merr.) seed yield are driven by linear increases in light interception, energy conversion, and partitioning efficiencies. *J. Exp. Bot.* 65:3311–3321. doi:10.1093/jxb/eru187
- Kucey, R.M.N., P. Chaiwanakupt, T. Arayangkool, P. Snitwongse, C. Siripaibool, P. Wadisirisuk, and N. Boonkerd. 1988a. Nitrogen fixation (15N dilution) with soybeans under Thai field conditions. II. Effect of herbicides and water application schedule. *Plant Soil* 108:87–92. doi:10.1007/BF02370103
- Kucey, R.M.N., P. Snitwongse, P. Chaiwanakupt, P. Wadisirisuk, C. Siripaibool, T. Arayangkool, N. Boonkerd, and R.J. Rennie. 1988b. Nitrogen fixation (15N dilution) with soybeans under Thai field conditions. I. Developing protocols for screening *Bradyrhizobium japonicum* strains. *Plant Soil* 108:33–41. doi:10.1007/BF02370097

- Kumudini, S., D.J. Hume, and G. Chu. 2001. Genetic Improvement in short season soybeans: I. Dry matter accumulation, partitioning, and leaf area duration. *Crop Sci.* 41:391–398. doi:10.2135/cropsci2001.412391x
- Kumudini, S., D.J. Hume, and G. Chu. 2002. Genetic improvement in short-season soybeans: II. Nitrogen accumulation, remobilization, and partitioning. *Crop Sci.* 42:141–145. doi:10.2135/cropsci2002.1410
- Kundu, S., S. Muneshwar, M.C. Manna, A.K. Tripathi, and P.N. Takkar. 1996. Effect of farmyard manure on nitrogen fixation in soybean (*Glycine max*) and its net potential contribution to N balance as measured by <sup>15</sup>N-tracer methodology. *Indian J. Agric. Sci.* 66:509–513.
- Leffel, R.C., P.B. Cregan, A.P. Bolgiano, and D.L. Thibeace. 1992. Nitrogen metabolism of normal and high-seed-protein soybean. *Crop Sci.* 32:747–750. doi:10.2135/cropsci1992.0011183X003200030034x
- Liu, M., Z. Yu, Y. Liu, and N. Konijn. 2006. Fertilizer requirements for wheat and maize in China: The QUEFTS approach. *Nutr. Cycling Agroecosyst.* 74:245–258. doi:10.1007/s10705-006-9002-5
- Lynch, D.H., and D.L. Smith. 1993. Soybean (*Glycine max*) nodulation and N<sub>2</sub>-fixation as affected by exposure to a low root-zone temperature. *Physiol. Plant.* 88:212–220. doi:10.1111/j.1399-3054.1993.tb05491.x
- Macedo, M.C.M., and C.H.B. Miranda. 2001. Fixação de nitrogênio pela soja em sistemas de cultivo contínuo e rotacionado com pecuária nos cerrados. Embrapa Gado de Corte, Campo Grande, Brazil. (Boletim de Pesquisa e Desenvolvimento).
- Maloney, T.S., K.G. Silveira, and E.S. Oplinger. 1999. Rotational vs. nitrogen-fixing influence of soybean on corn grain and silage yield and nitrogen use. *J. Prod. Agric.* 12:175–187. doi:10.2134/jpa1999.0175
- Mead, R., R.N. Curnow, and A.M. Hasted. 1993. Statistical methods in agriculture and experimental biology. Chapman and Hall, London. doi:10.1007/978-1-4899-6928-6
- Mengpei, Y., S. Keyong, L. Qizhen, L. Zenghui, C. Congyun, and D. Shujue. 1986. Studies on nutrition and fertilizer application method of summer-sown soybean. *Soybean Sci.* 5:317–326.
- Motulsky, H.J., and A. Christopoulos. 2003. Fitting models to biological data using linear and nonlinear regression. A practical guide to curve fitting. GraphPad Software, Inc., San Diego, CA.
- Munyinda, K., R.E. Karamanos, J.O. Legg, and S. Sanogho. 1988. Nitrogen fixation by soybeans (*Glycine max* L.) in Zambia. *Plant Soil* 109:57–63. doi:10.1007/BF02197580
- Neves, M.C.P., A.D. Didonet, F.F. Duque, and J. Dobereiner. 1985. Rhizobium strain effects on nitrogen transport and distribution in soybeans. *J. Exp. Bot.* 36:1179–1192. doi:10.1093/jxb/36.8.1179
- Ofosu-Budu, K.G., K. Fujita, and S. Ogata. 1990. Excretion of ureide and other nitrogenous compounds by the root system of soybean at different growth stages. *Plant Soil* 128:135–142. doi:10.1007/BF00011102
- Park, S.J., W.H. Kim, J.E. Lee, Y.U. Kwon, J.C. Shin, Y.H. Ryu, and R.C. Seong. 2005. Nitrogen balance and biological nitrogen fixation of soybean in soybean-barley cropping system. *Korean J. Crop Sci.* 50:1–4.
- Peoples, M.B., A.W. Faizah, B. Rerkasem, and D. Herridge. 1989. Methods for evaluating nitrogen fixation by nodulated legumes in the field. Australian Centre for International Agricultural Research, Canberra, Australia.
- Peoples, M.B., R.R. Gault, B. Lean, J.D. Sykes, and J. Brockwell. 1995. Nitrogen fixation by soybean in commercial irrigated crops of central and southern New South Wales. *Soil Biol. Biochem.* 27:553–561. doi:10.1016/0038-0717(95)98631-W
- R Development Core Team. 2009. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria.
- Ravuri, V., and D.J. Hume. 1993. Soybean stover nitrogen affected by dinitrogen fixation and cultivar. *Agron. J.* 85:328–333. doi:10.2134/agronj1993.000219620085000200031x
- Rennie, R., D. Rennie, C. Siripaibool, P. Chaiwanakupt, N. Boonkerd, and P. Snitwongse. 1988. N<sub>2</sub> fixation in Thai soybeans: Effect of tillage and inoculation on <sup>15</sup>N-determined N<sub>2</sub> fixation in recommended cultivars and advanced breeding lines. *Plant Soil* 112:183–193. doi:10.1007/BF02139994
- Rochester, I.J., M.B. Peoples, G.A. Constable, and R.R. Gault. 1998. Faba beans and other legumes add nitrogen to irrigated cotton cropping systems. *Aust. J. Exp. Res.* 38:253–260. doi:10.1071/EA97132
- Salsac, L., J.J. Drevon, M. Zengbq, J.C. Cleyet-Marel, and M. Obaton. 1984. Energy requirement of symbiotic nitrogen fixation. *Physiol. Veg.* 22:509–521.
- Salvagiotti, F., K.G. Cassman, J.E. Specht, D.T. Walters, A. Weiss, and A. Dobermann. 2008. Nitrogen uptake, fixation and response to fertilizer N in soybeans: A review. *F. Crop. Res.* 108:1–13. doi:10.1016/j.fcr.2008.03.001
- Salvagiotti, F., J.E. Specht, K.G. Cassman, D.T. Walters, A. Weiss, and A. Dobermann. 2009. Growth and nitrogen fixation in high-yielding soybean: Impact of nitrogen fertilization. *Agron. J.* 101:958–970. doi:10.2134/agronj2008.0173x
- Sanginga, N., K. Dashiell, J.A. Okogun, and G. Thottappilly. 1997. Nitrogen fixation and N contribution by promiscuous nodulating soybeans in the southern Guinea savanna of Nigeria. *Plant Soil* 195:257–266. doi:10.1023/A:1004207530131
- Sanginga, N., J.A. Okogun, B. Vanlauwe, and K. Dashiell. 2002. The contribution of nitrogen by promiscuous soybeans to maize based cropping the moist savanna of Nigeria. *Plant Soil* 241:223–231. doi:10.1023/A:1016192514568
- Santachiara, G., L. Borrás, F. Salvagiotti, J.A. Gerde, and J.L. Rotundo. 2017. Relative importance of biological nitrogen fixation and mineral uptake in high yielding soybean cultivars. *Plant Soil.* doi:10.1007/s11104-017-3279-9
- Schubert, K.R. 1982. The energetics of biological nitrogen fixation. American Society of Plant Physiologist, Rockville, MD.
- Setiyono, T.D., D.T. Walters, K.G. Cassman, C. Witt, and A. Dobermann. 2010. Estimating maize nutrient uptake requirements. *Field Crops Res.* 118:158–168. doi:10.1016/j.fcr.2010.05.006
- Shimada, S., H. Hamaguchi, Y. Kim, K. Matsuura, M. Kato, T. Kokuryu, J. Tazawa, and S. Fujimori. 2012. Effects of water table control by farm-oriented enhancing aquatic system on photosynthesis, nodule nitrogen fixation, and yield of soybeans. *Plant Prod. Sci.* 15:132–143. doi:10.1626/pp.s.15.132
- Singh, M., R.H. Wanjari, B.K. Agrawal, and P. Sharma. 2014. Biological N<sub>2</sub> fixation in soybean and contribution to soil in a 40-year-old experiment on Alfisols of Ranchi. *J. Indian Soc. Soil Sci.* 62:56–61.
- Sisworo, W.H., M.M. Mitrosuhardjo, H. Rasjid, and R.J.K. Myers. 1990. The relative roles of N fixation, crop residues and soil in supplying N in multiple cropping systems in a humid, tropical upland cropping system. *Plant Soil* 121:73–82. doi:10.1007/BF00013099
- SoyStat. 2016. International: Brazil & Argentina production. <http://soystats.com/international-brazil-argentina-production/> (accessed June 2017).
- Ta, T.C., F.D.H. McDowall, and M.A. Faris. 1986. Excretion of nitrogen assimilated from N<sub>2</sub> fixed by nodulated roots of alfalfa (*Medicago sativa*). *Can. J. Bot.* 64:2063–2067. doi:10.1139/b86-270
- Takahashi, Y., T. Chinushi, Y. Nagumo, T. Nakano, and T. Ohyama. 1991. Effect of deep placement of controlled release nitrogen fertilizer (coated urea) on growth, yield, and nitrogen fixation of soybean plants. *Soil Sci. Plant Nutr.* 37:223–231. doi:10.1080/00380768.1991.10415032

- Tamagno, S., and I.A. Ciampitti. 2017. Seed yield and biological N fixation for historical soybean genotypes. Kansas field research report. <http://newprairiepress.org/cgi/viewcontent.cgi?article=7436&context=kaesrr> (accessed Oct. 2017). doi:10.4148/2378-5977.7436
- Tamagno, S., G.R. Balboa, Y. Assefa, P. Kovács, S.N. Casteel, F. Salvagioti, F.O. García, W.M. Stewart, and I.A. Ciampitti. 2017. Nutrient partitioning and stoichiometry in soybean: A synthesis-analysis. *F. Crop. Res.* 200:18–27. doi:10.1016/j.fcr.2016.09.019
- Tewari, K., T. Sukanuma, H. Fujikake, N. Ohtake, K. Sueyoshi, Y. Takahashi, and T. Ohya. 2004. Effect of deep placement of N fertilizers and different inoculation methods of Bradyrhizobia on growth, N<sub>2</sub> fixation activity and N absorption rate of field-grown soybean plants. *J. Agron. Crop Sci.* 190:46–58. doi:10.1046/j.0931-2250.2003.00073.x
- Tewari, K., M. Onda, S. Ito, A. Yamazaki, H. Fujikake, N. Ohtake, K. Sueyoshi, Y. Takahashi, and T. Ohya. 2005. <sup>15</sup>N Analysis of the promotive effect of deep placement of slow-release N fertilizers on growth and seed yield of soybean. *Soil Sci. Plant Nutr.* 51:885–892. doi:10.1111/j.1747-0765.2005.tb00123.x
- Thies, J.E., P.W. Singleton, and B.B. Bohlool. 1995. Phenology, growth, and yield of fieldgrown soybean and bush bean as a function of varying modes of N nutrition. *Soil Biol. Biochem.* 27:575–583. doi:10.1016/0038-0717(95)98634-Z
- Tittonell, P., K.D. Shepherd, B. Vanlauwe, and K.E. Giller. 2008. Unraveling the effects of soil and crop management on maize productivity in smallholder agricultural systems of western Kenya—an application of classification and regression tree analysis. *Agric. Ecosyst. Environ.* 123:137–150. doi:10.1016/j.agee.2007.05.005
- Toomsan, B., J. McDonagh, V. Limpinuntana, and K. Giller. 1995. Nitrogen fixation by groundnut and soybean and residual nitrogen benefits to rice in farmers fields in north-east Thailand. *Plant Soil* 175:45–56. doi:10.1007/BF02413009
- Uchida, Y., and H. Akiyama. 2013. Mitigation of postharvest nitrous oxide emissions from soybean ecosystems: A review. *Soil Sci. Plant Nutr.* 59:477–487. doi:10.1080/00380768.2013.805433
- Unkovich, M.J., and J.S. Pate. 2000. An appraisal of recent field measurements of symbiotic N<sub>2</sub> fixation by annual legumes. *Field Crops Res.* 65:211–228. doi:10.1016/S0378-4290(99)00088-X
- USDA NASS. 2017. United States and all state data—crops. <http://www.nass.usda.gov/QuickStats/>. (accessed June 2017).
- van Kessel, C., and C. Hartley. 2000. Agricultural management of grain legumes: Has it led to an increase in nitrogen fixation? *F. Crop. Res.* 65:165–181. doi:10.1016/S0378-4290(99)00085-4
- Vasilas, B.L., and G.E. Ham. 1985. Intercropping nodulating and non-nodulating soybeans: Effects on seed characteristics and dinitrogen fixation estimates. *Soil Biol. Biochem.* 17:581–582. doi:10.1016/0038-0717(85)90030-6
- Vasilas, B.L., and J.J. Fuhrmann. 1993. Field response of soybean to increased dinitrogen fixation. *Crop Sci.* 33:785–787. doi:10.2135/cropsci1993.0011183X003300040031x
- Weber, C.R. 1966. Nodulating and non-nodulating soybean isolines. I. Agronomic and chemical attributes. *Agron. J.* 58:43–46. doi:10.2134/agronj1966.00021962005800010014x
- Witt, C., A. Dobermann, S. Abdulrachman, H.C. Gines, W. Guanghuo, R. Nagarajan, S. Satawatananont, T. Thuc Son, P. Sy Tan, and L. Van Tiem. 1999. Internal nutrient efficiencies of irrigated lowland rice in tropical and subtropical Asia. *Field Crops Res.* 63:113–138. doi:10.1016/S0378-4290(99)00031-3
- Wu, D., D.J. Hume, T.J. Vyn, and E.G. Beauchamp. 1998. N credit of soybean to a following corn crop in central Ontario. *Can. J. Plant Sci.* 78:29–33. doi:10.4141/P96-180
- Xinmin, L., D. Xintian, and L. Xiaoming. 1993. Evaluation of nodulation and nitrogen fixation ability of soybean cultivars grown in field. *Soybean Sci.*
- Yang, L., and Z. Cai. 2005. The effect of growing soybean (*Glycine max.* L.) on N<sub>2</sub>O emission from soil. *Soil Biol. Biochem.* 37:1205–1209. doi:10.1016/j.soilbio.2004.08.027
- Yong, T., W. Liu, L. Zhou, C. Song, F. Yang, L. Jiang, X. Wang, and W. Yang. 2015. Effects of reduced nitrogen application on nitrogen uptake and utilization efficiency in maize-soybean relay strip intercropping system. *Acta Ecol. Sin.* 35:4473–4482.
- Zapata, F., S.K.A. Danso, G. Hardarson, and M. Fried. 1987. Time course of nitrogen fixation in field-grown soybean using nitrogen-15 methodology. *Agron. J.* 79:172–176. doi:10.2134/agronj1987.00021962007900010035x
- Zhang, H., Z. Zhang, K. Zhao, X.M. Wang, B. Xu, and L. Zhao. 1986. Nitrogenase activity nodulation and the N<sub>2</sub> fixation of the indigenous rhizobium japonicum in black soil with soybean hosts. *Soybean Sci.* 5:47–56.
- Zingore, S., H.K. Murwira, R.J. Delve, and K.E. Giller. 2008. Variable grain legume yields, responses to phosphorus and rotational effects on maize across soil fertility gradients on African smallholder farms. *Nutr. Cycling Agroecosyst.* 80:1–18.
- Zotarelli, L. 2000. Balanço de nitrogênio na rotação de culturas em sistemas de plantio direto e convencional na região de Londrina-PR. Ms.Sc. thesis, Universidade Federal Rural do Rio de Janeiro, Brazil.