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### Occurrence and characterization of a severe isolate of *Watermelon mosaic virus* from Argentina

María Cecilia Perotto • M. G. Celli • E. A. Pozzi • C. E. Luciani • V. C. Conci

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Abstract More than 50 viruses have been reported in cucurbit crops worldwide. In Argentina, cucurbit viruses have been associated with important yield losses. The most prevalent and widespread potyvirus is Watermelon mosaic virus (WMV). WMV was detected in Argentina in all cucurbit species with high incidence. In this study, a WMV isolate (WMV 1 SDE FF) was obtained from a naturally infected squash associated with a severe outbreak on melon and squash crops in an important cucurbit growing area in Santiago del Estero province (Argentina), during a survey conducted in November 2012. The fully sequenced WMV 1 SDE FF genome consists of 10,027 nucleotides and shares 96 % nt identity and 98 % aa identity with the French isolates JF273464.1|C07-014 and EU660581.1|FMF00-LL1 of the WMV molecular group 3. Using the recombination detection program RDP4, two statistically significant recombination events were identified: event 1, an 830-nt long recombinant fragment in the putative P1 coding region, and event 2,

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a 4071-nt recombinant fragment detected across the HC Pro, P3 and CI coding regions. The putative parental sequences detected for event 1 were the EU660586.1| FBR04–37 (major parent) and JF273468.1|C07–284 (minor parent), both from France. Putative parental sequences for event 2 were JX079685.1| WMV-ShanXi (major parent) and HQ384216.1|Dendrobium (minor parent), from China and USA, respectively. To our knowledge, this is the first complete genome of an Argentine WMV isolate. Our results provide evidence that WMV 1 SDE FF is the causal agent of the strong outbreak reported in melon and squash fields in recent years.

**Keywords** WMV· Cucurbitaceae · Virus recombination · Phylogenetic analysis

Cucurbit crops are affected by more than 50 different viruses from several different taxonomic groups. Viral diseases have been considered the main limiting factor in terms of yield and fruit quality in major cucurbit crops worldwide (Lecoq and Desbiez 2012; Lecoq et al. 1998; Zitter et al. 1996). In Argentina, the main cultivated cucurbits are squash (*Cucurbita moschata*), pumpkin (*Cucurbita pepo*), and melon (*Cucumis melo*) (FAOSTAT 2013).

To date, four of the most common and prevalent plant viruses worldwide all transmitted by aphids, were reported in Argentina. Three of those viruses belong to the genus *Potyvirus*: *Watermelon mosaic virus* (WMV), *Papaya ringspot virus* (PRSV) and *Zucchini yellow mosaic virus* (ZYMV) the remaining one, *Cucumber mosaic* 



virus (CMV), belongs to the genus Cucumovirus, In the earliest reports, detection was confirmed by serological and differential host assays. WMV was first detected in 1971 (Nome et al. 1974) and since then its presence was always confirmed in all surveys conducted in Argentina. Thus, WMV has become an important threat for production (Gracia and Feldman 1986). PRSV was detected in 1987 from samples of C. moschata in Santiago del Estero (Feldman and Gracia 1992). ZYMV and CMV were detected in squash (C. maxima and C. pepo) and in melon (C. melo) (Gracia 2000; Gracia and Feldman 1986).

Since 2011, the most important growing cucurbit areas of Argentina have been surveyed every year. The presence of viruses was confirmed for the first time by examining symptomatic plants using molecular tests. In 2012 melon and squash plants with severe mosaic, leaf deformation, blistering and vein clearing were observed in production areas of Santiago del Estero province, where a significant effect on fruit yield and quality was recorded. Later a similar situation occurred in 2014 with melon and squash in Cuyo region. In both cases, WMV was detected in all symptomatic plants by double antibody sandwich, enzyme-linked immunosorbent assay (DAS ELISA) and confirmed by Reverse transcription-polymerase chain reaction (RT-PCR) (Moreno et al. 2004) using specific primers in all symptomatic plants tested. The presence of WMV was detected at high frequencies 98 % in 2012 and 100 % in 2014 from 91 and 150 symptomatic plants respectively, in all cucurbit species. WMV has been the most prevalent and widespread virus found in all cucurbit growing regions in Argentina since the earliest record (unpublished data). In addition, a DAS-ELISA test was performed for the samples collected, using antisera against ZYMV, PRSV, CMV, Squash mosaic virus (SqMV) and Cucurbit yellow stunting disorder virus (CYSDV). The serological testing kits were all commercial antisera specific from Bioreba AG (Switzerland). In order to test the putative presence of other viruses, PCR was performed with universal primers for Begomovirus and RT-PCR was used with specific primers for Cucurbit aphidborne yellows virus (CABYV) and Cucumber vein yellowing virus (CVYV). The presence of PRSV, ZYMV and CMV was possible to detect in very low percentage, ranging from 2 % to 17 %.

Electron microscope observation of leaf dips from symptomatic plant revealed the presence of a virus with filamentous particles, and ultra-thin leaf sections showed cylindrical inclusions typically described for potyvirus. To date, four types of cylindrical inclusions have been recognized. Viruses inducing these types of inclusions have been separated into four subdivisions according to Edwardson (1992). Based on this classification, the WMV Argentinian isolate (WMV 1 SDE FF) belongs to type 3, showing pinwheels with scrolls and laminated aggregates presented in the cytoplasm of the infected host cells (not shown).

The aim of this work was to molecularly characterize the pathogen responsible for this severe disease in Argentina. A phylogenetic and recombination analysis was performed to analyze the sequence of the WMV isolate. Leaves of naturally infected plants were collected from a field-grown squash showing typical virus symptoms (Fig. 1). Total RNA was extracted from 100 mg of squash leaf tissue of a WMV-infected plant (WMV 1 SDE FF) using a Qiagen RNA kit. RNA was quantified and 5 µg were sent to INDEAR (Genomics and Bioinformatics Platform, INDEAR Inc., Rosario, Argentina) for synthesis of cDNA from polyadenylated RNA followed by deep sequencing using Illumina HiSeq 1500.

The reads of the full-length genome were assembled either de novo or by mapping of the Illumina reads to a reference WMV genome (GenBank NC 006262) using the Mira v 4 assembling method with a coverage of 16,454 reads/nt. The complete WMV 1 SDE FF genome sequence consisted of 10,027 nt (GenBank accession number KP164988). It presented a single large open reading frame (ORF), starting at nt 127 and ending at nt 9775 that encodes a large polyprotein with 3215 amino acids (aa). The putative cleavage sites of the genome for the viral-encoded proteinases yield all the 10 characteristic potyviral proteins, include the 11th protein PIPOwith estimated sizes of 439, 455, 347, 52, 634, 53, 190, 243, 517 and 283 aa for P1, HC-Pro, P3, 6 K1, CI, 6 K2, NIaVPg, VPg Pro, NIb and CP, respectively. The comparison of the complete genomic sequence of WMV 1 SDE FF with those of other WMV available in the GenBank by BLAST analysis showed the highest identity (96 % nt and 98 % aa) with the French isolates, JF273464.1and EU660581.1.

The virus sequence obtained from the RNA-seq was aligned with 35 sequences retrieved from GenBank (http://www.ncbi.nlm.nih.gov/) and with *Soybean mosaic virus* (SMV) (accession no. AJ628750) as an outgroup using the algorithm muscle in MEGA6, before conducting the phylogenetic analysis (Tamura et al. 2013). Neighbor-joining trees were constructed using the p-distance method with a bootstrap value of



**Fig. 1** Leaf and fruit symptoms in a WMV-infected plant





10,000; Maximum Likelihood trees using the Tamura-Nei model with a bootstrap value of 10,000; Minimum Evolution trees using the Number of differences model with a bootstrap value of 10,000.

Phylogenetic relationships among WMV isolates and the topologies inferred by the different methods (3) were all similar. The WMV isolates formed two distinct major groups based on genetic distances. Group B clustered five isolates and group A clustered the remaining 31 isolates. Group A was further divided into three subgroups: G1, G2 and G3. These three groups were defined by Desbiez et al. (2007). Our phylogenetic analysis showed that WMV 1 SDE FF belongs to subgroup G3 of WMV (Fig. 2). Among G3 isolates, four subgroups were defined, namely EM1 to EM4 (Desbiez et al. 2009); according to that classification, group A (this paper) corresponds to the EM3 subgroup. Intragroup variability in the whole sequences calculated with MEGA 6 was highest within group B  $(0.058 \pm 0.002)$ , followed by subgroups G3, G1 and G2 (0.049  $\pm$  0.001, 0.040  $\pm$  0.001 and 0.035  $\pm$  0.002, respectively). Intergroup variability was highest between groups B and G1 (0.094  $\pm$  0.002), being slightly lower between G1 and G2 (0.092  $\pm$  0.002). The lowest variability for group B was with G3, as expected  $(0.085 \pm 0.002)$ . Thus, the assignment of isolates to each subgroup is unambiguous with a high bootstrap support.

Little information is available on WMV genetic characterization based on full-length genomes. Furthermore, molecular characterization of this virus is scarce in the American continent. WMV has been reported in USA (HQ384216), Chile (EU660580 and EU660582) and more recently in Venezuela (KC292915) (Desbiez and Lecoq 2008; Romay et al. 2013). Desbiez et al. (2007) reported a sequence of 267 nt of the CP-NIb region from 1994 (DQ845043) of an isolate from Argentina that was included in G1. Another sequence analysis of the NIb-CP region showed that most of the Argentine isolates

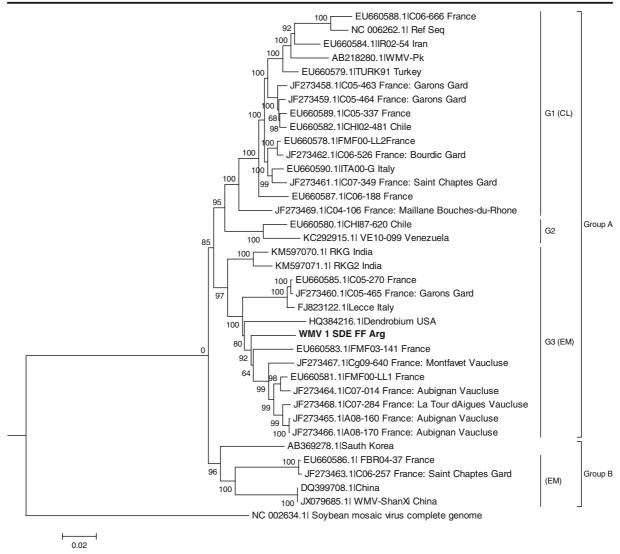
found nowadays are G3, few of them G2, but none of them are G1 (data not shown). Our results suggest a replacement of G1 with G3 in Argentina, as happened in other countries. Surveys carried out in several countries revealed a rapid replacement of local G1 classic strains (CL) with recently introduced G3 emerging ones (EM) (Fabre et al. 2010; Lecoq et al. 2011; Borodynko et al. 2009; Lecoq and Desbiez 2012; Finetti-Sialer et al. 2012; Kamberoglu et al. 2015). This replacement is very important since group classification is well correlated with differences in symptom intensity; isolates causing very severe symptoms belong to G3, whereas others causing mild symptoms are included in G1 (Desbiez et al. 2009).

The evidence that WMV 1 SDE FF belongs to molecular subgroup G3 is also supported by the fact that it potentially codes for the aa motif KEKET at position 3–7 in the N-terminal part of the CP, characteristic of molecular G3 isolates (Desbiez et al. 2007).

The Recombination Detection Program v.4.16 (RDP4) was used for recombination analysis (Martin and Rybicki 2000). Recombination events, likely parental isolates of recombinants, and recombination break points were analyzed. Default parameters were used for the seven programs implemented in the RDP4 program: RDP, GENECONV, Chimaera, MaxChi, BOOTSCAN, SISCAN and 3Seq including a Bonferroni corrected *P* value cutoff of 0.01. A recombination pattern was considered to be a firm event, and genuine evidence of actual recombination, if detected by four or more of these programs (Wylie and Jones 2009; Kehoe et al. 2014).

When the complete sequence of 35 WMV (retrieved from Genbank) and the WMV 1 SDE FF isolate were analyzed using RDP4, two statistically significant recombination events were identified. Event 1, comprises an 830-nt long recombinant fragment in the putative P1 coding region, and event 2, a 4071-nt recombinant





**Fig. 2** Neighbor-joining tree illustrating the phylogenetic relationships between WMV 1 SDE FF isolate and 35 published complete nucleotide sequences. The two main clusters (Groups A and B) and subgroups (G1(CL), G2 and G3(EM)) are indicated. The Argentine

isolate is shown in bold. SMV was included as an outgroup Bootstrap support (10,000 replicates) is shown next to the branches. Evolutionary analyses were conducted in MEGA6

fragment detected across the HC Pro, P3 and CI coding regions. The putative parentals detected for event 1 were

the EU660586.1| FBR04-37 (major parent) and JF273468.1|C07-284 (minor parent), both from France.



**Fig. 3** Analysis of recombination in full-length sequence of an WMV isolate from Argentina. Two recombination events were detected. The location of each potyvirus gene is shown (top). The different shadings indicate the viruses that supplied the different

regions with the sites and sizes of recombination patterns within them as assessed by six recombination-detection algorithms (RDP4 program)



Putative parental sequences for event 2 were JX079685.1| WMV-ShanXi (major parent) and HQ384216.1|Dendrobium (minor parent), from China and USA, respectively. In both events the major parentals belong to Group B while the minor parentals belong to G3. Both recombination events were detected using the RDP4 program and were positive for six out of seven algorithms in both cases. The recombination breakpoints (Rbps) were identified by RDP  $(P = 8,163 \times 10^{-17})$ , GENECONV  $(1211 \times 10^{-16})$ , BootScan  $(2893 \times 10^{-17})$ , Maxchi  $(1794 \times 10^{-11})$ , Chimaera  $(4316 \times 10^{-06})$  and SiScan  $(9852 \times 10^{-17})$  for event 1. Similar results were obtained for event 2, 3Seq  $(P = 8.932 \times 10^{-101})$ , GENECONV  $(1045 \times 10^{-05})$ , BootScan  $(5223 \times 10^{-06})$ , Maxchi  $(3139 \times 10^{-12})$ , Chimaera  $(2001 \times 10^{-09})$  and SiScan  $(8601 \times 10^{-20})$ . The sequences of the C07–014, FMF00-LL1 isolates show the same recombination event 2. As expected, our isolate shares the highest identity score with those isolates that were also confirmed by SDT 1.2 software JF273464.1|C07014 96.1 % and EU660581.1|FMF00LL1 France 96.0 % (Fig. 3).

In this study, we report the biological and molecular characterization of a WMV isolate, named WMV 1 SDE FF (accession KP164988), being the first complete genome of an Argentinian isolate. Our results provide evidence that WMV 1 SDE FF is associated with the strong outbreak reported in last years in melon and squash fields and that it is a recombinant of molecular emerging strain of the virus. We do not have any information on the temporal evolution of WMV populations in the country or how it was introduced into Argentina. Further studies should be planned and carried out to control this important disease in the cucurbit growing areas in Argentina.

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