

## Article

# Increased Nitrogen Retention and Reduced Methane Emissions of Beef Cattle Grazing Legume vs. Grass Irrigated Pastures in the Mountain West USA

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**Abstract:** Grazing studies were carried out over a 5-year period using pregnant cows, yearling calves and 2-year-old heifers to investigate the influence of diet on intake, methane (CH<sub>4</sub>) emissions and retention of nitrogen (N). Monoculture legume (birdsfoot trefoil, BFT and cicer milkvetch, CMV) or grass (meadow brome grass, MBG) pastures were rotationally stocked, and during year 4 and year 5, treatments were contrasted with total mixed rations (TMR) fed in confinement. The sulfur hexafluoride (SF<sub>6</sub>) method was used to continuously measure enteric CH<sub>4</sub> emissions. Intake was greater on legume pastures and on TMR than on grass pastures, and enteric CH<sub>4</sub> emissions per unit of intake were lower on legumes compared with grass pastures. Legume pastures had elevated non-fiber carbohydrate (NFC) concentrations (400 g kg<sup>-1</sup> dry matter; DM) typical of perennial legumes cultivated in the Mountain West. A N balance calculated in 2017–2018 demonstrated that N retention was greater for TMR and legume than grass pastures. Enteric CH<sub>4</sub> emissions of grazing cow herds account for the majority of greenhouse gas (GHG) emissions from beef production and can be significantly reduced by using highly digestible forage legumes. The N retention of legumes can potentially enhance the efficiency of N use, thereby increasing the sustainability of grasslands.

**Keywords:** birdsfoot trefoil; cicer milkvetch; condensed tannins; enteric methane; meadow brome grass; nitrogen retention; non-fiber carbohydrates

## 1. Introduction

The sustainability of U.S. agriculture is threatened by the degradation and/or loss of ecosystem services due to global warming and anthropogenic interventions such as reduced biological diversity; water and air pollution; and loss of soil quality [1–3]. The sustainability of beef production has come under considerable scrutiny given increased concerns over the use of cereal grain for livestock feed [4,5] and the contributions of CH<sub>4</sub> and N<sub>2</sub>O from livestock production to global warming [6]. U.S. wetlands and grasslands have been converted to cropland for the production of annual feed grains, reducing ecosystem services provided by these lands. Of the annual grains produced in the U.S., 70% is used as livestock feed and approximately 35% of the grain consumed by livestock is fed to beef and dairy cattle [7]. Cereal grains require annual nitrogen (N) fertilization and periodic replacement of soil phosphorus and other mineral nutrients that are removed as they are harvested.

Mechanization and fuel are needed for their cultivation, planting and harvesting, and annual grain crop production is associated with significant soil loss via erosion. When grain is used for livestock feed instead of human consumption, the negative environmental effects accrue relative to animal agriculture, while the ecosystem service of provisioning is reduced by the inefficient conversion of grains to ruminant products such as meat and milk. However, the ruminant digestive system does not require concentrates such as grain and can derive energy from the cellulose of forages and other feeds that cannot be digested by swine or poultry. The ability to utilize plant fiber for energy places ruminants in a unique position in the world's economy [8], but this advantage is lost when concentrates are fed to ruminants. On the other hand, the nutrient density of grasses is low relative to concentrates [8], which adds to inefficiencies in nutrient use and results in increased time needed for the finishing process, which constitutes constraints that result in increments in greenhouse gas (GHG) emissions by the beef sector [9].

An increasing global population and improved standards of living provide a market for high-quality ruminant protein in meat and milk. In contrast to cereal grains and pasture grasses, perennial legumes fix their own N, present high nutritional quality and are productive for multiple years after establishment without additional cultivation or planting. Thus, by establishing perennial legume pastures on lands unsuitable for intensive cropping, ruminant productivity per land unit area could be optimized, making it unnecessary for ruminants to compete with humans for agricultural land or crops. In addition, legumes such as BFT and CMV are non-bloating and can, therefore, be grazed in pure stands, and legumes grown under irrigation in the Mountain West contain high concentrations of NFC that increase the efficiency of protein use.

Perennial legumes are digested more rapidly than grasses by ruminants; thus, intake and production are typically greater than for forage grasses. In addition, the unique plant secondary compounds (i.e., condensed tannins, CT) produced by some temperate legumes such as birdsfoot trefoil (*Lotus corniculatus* L.; BFT), as well as their high fiber digestibility [10–12] enhance the efficiency of energy and protein use in ruminants relative to other forages. Thus, it is likely that ruminant productivity per land unit area could be optimized with perennial legume pastures under management-intensive grazing. Consistent with this idea, the dry matter intake (DMI) and milk yields of cows fed with legume silage were greater than cows fed grass silage [13], but a meta-analysis of in vivo grass and legume ruminant methane (CH<sub>4</sub>) emission studies did not find differences in intake and therefore in CH<sub>4</sub> emissions [14]. While the proportion of perennial legumes such as alfalfa (*Medicago sativa* L.), red clover (*Trifolium pratense* L.) and white clover (*T. repens* L.) must be limited in pasture mixtures to prevent bloat, non-bloating legumes such as BFT, cicer milkvetch (*Astragalus cicer* L., CMV) and sainfoin (*Onobrychis viciifolia* Scop.) can be grazed alongside grasses or as monoculture alternatives to grass pastures.

It was demonstrated for Western Canadian beef cattle production systems that enteric CH<sub>4</sub> accounted for 63% of all GHG emissions in terms of carbon dioxide (CO<sub>2</sub>) equivalents and that 80% of GHG emissions occurred in the cow-calf phase [15]. Systems that include feedlot-finished cattle have been found to have a smaller carbon footprint than grazing-based systems largely because more time is needed for finishing (679 vs. 440 d for grass vs. feedlot, respectively; [9]), but red meat production can be nearly as rapid on Mountain West perennial legume pastures as in the feedlot, with comparable consumer appeal and an increased healthful balance of omega-6 to omega-3 fatty acids [16,17].

We designed the current study to determine the influence of forage nutritive value and intake on the relative enteric CH<sub>4</sub> emissions of cows, calves and heifers, the classes of animals comprising the grazing cowherd. In the final 2 years of the study, a N balance calculation of 2-year-old heifers grazing a CT-containing legume (BFT), a non-tannin legume (CMV) and a cool-season grass, meadow brome (*Bromus biebersteinii* Roem. and Schult., MBG) also included a total mixed ration (TMR) confinement diet treatment.

The legume BFT contains a relatively low concentration of CT that is sufficient to prevent bloating, while the non-tannin legume CMV is non-bloating because the digestion

of leaves is slowed by their structural anatomy [18]. The CT in BFT may favor N retention due to the formation of stable tannin–protein complexes in the rumen that reduce proteolysis and ammonia formation, while dissociating in the abomasum and thereby supplying high-quality protein to the intestines [19]. We used the cool-season grass, MBG, which is a bunchgrass with good nutritive value that is productive at high elevations of the Mountain West [20].

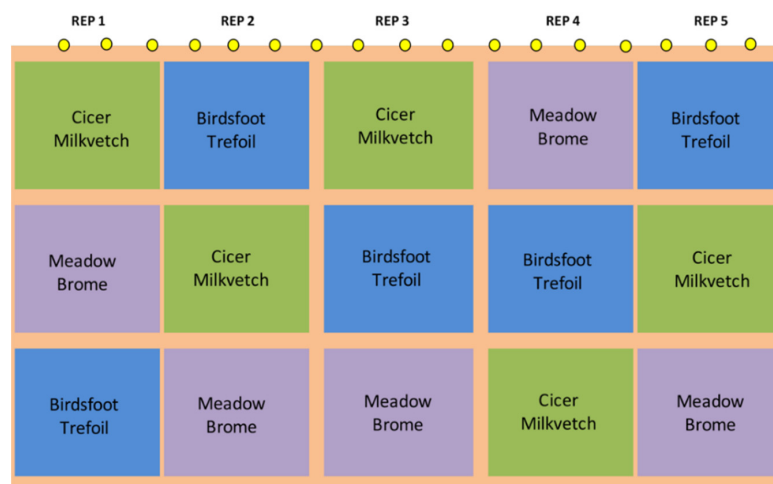
Birdsfoot trefoil and CMV have been found to accumulate high concentrations of non-fiber carbohydrates (NFC) when grown under irrigation in the Mountain West [21,22]. The enteric CH<sub>4</sub> emissions of dairy cattle fed with legume silages or pastures were less than those of cows fed with grass silage or pasture due to reduced fiber concentration, greater nutritive concentration and rapid reduction in particle size that allows legumes to leave the rumen more quickly than grasses [13,23]. The elevated NFC of Mountain West-grown perennial legumes complements their high N concentrations, which contributes to enhanced N retention [21,24].

Our hypothesis was that enteric CH<sub>4</sub> emissions would be significantly lower for cattle grazing legumes than grasses in a Mountain West environment due to greater nutritive value of legumes and resulting elevated intakes. Based on earlier studies [17,21], we hypothesized that N retention would be enhanced in cattle grazing legume pastures relative to grass pastures and similar to that of cattle fed a TMR.

## 2. Materials and Methods

### 2.1. Pasture Establishment

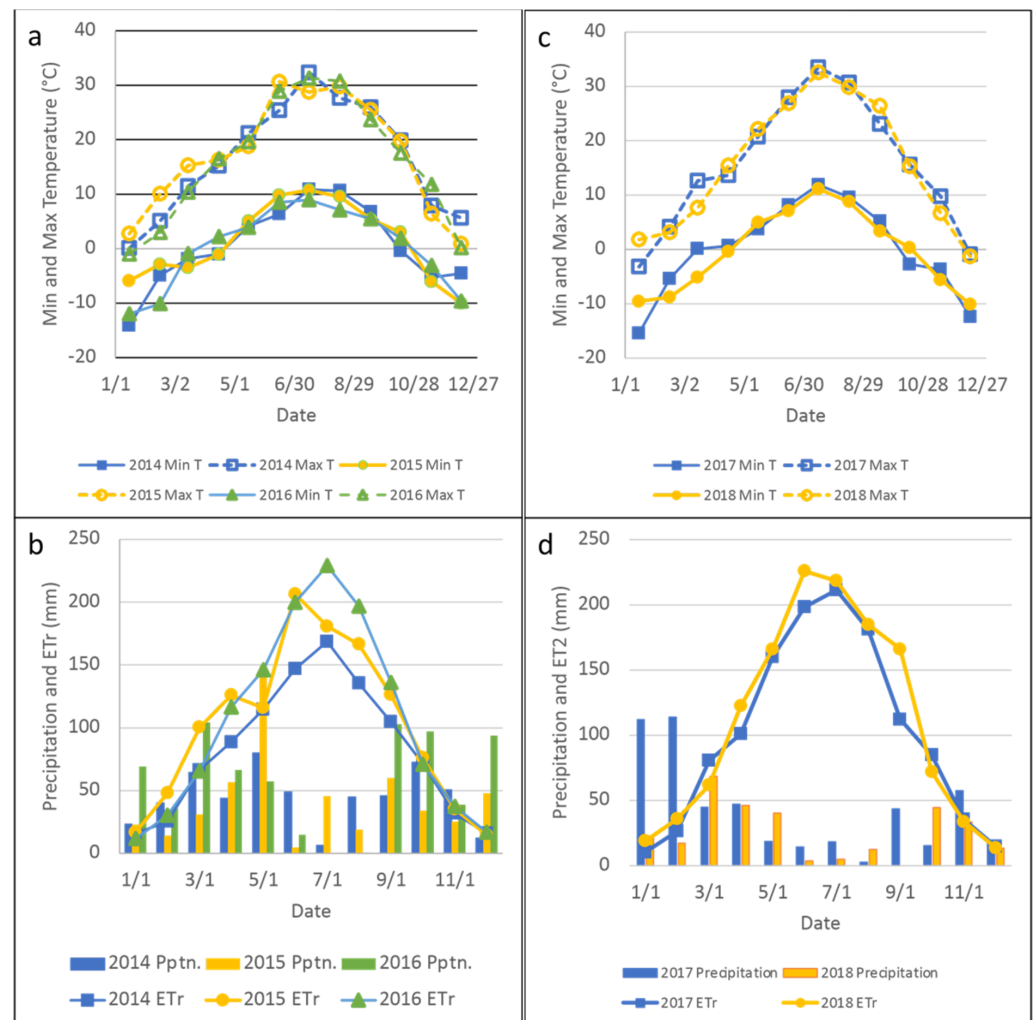
This study was carried out between 2014 and 2018 on 6.7 ha at the Utah State University (USU) Intermountain Irrigated Pasture Project in Lewiston, UT, USA (latitude 41°56' N, longitude 111°52' W; 1374 m a.s.l.). Two soil series are present: Kidman fine sandy loam (coarse-loamy, mixed and mesic Calcic Haploxeroll) and Lewiston fine sandy loam (coarse-loamy, mixed and mesic Aeric Calciaquoll). Soil tests were conducted during the summer of 2012, and deficiencies of phosphorus and potassium were addressed by adding 112 kg/ha P<sub>2</sub>O<sub>5</sub> and 135 kg/ha K<sub>2</sub>O prior to planting. Three monoculture pasture treatments, CMV cv. 'Monarch,' BFT cv. 'Langille' and MBG cv. 'Cache,' were replicated five times in a randomized complete block design (Figure 1); individual pastures were approximately 0.365 ha (64 × 57 m).



**Figure 1.** Plot plan of the grazing study. Cattle were assigned to one of fifteen plots between 2014 and 2018 and rotationally grazed for 7 to 12 weeks depending on the year. Yellow circles indicate irrigation risers.

Birdsfoot trefoil seed was inoculated with *Mesorhizobium loti* bacterium and CMV seed with *Rhizobium leguminosarum* bacterium at planting, and grass pastures received 56 kg/ha 34-0-0 fertilizer (N-P<sub>2</sub>O<sub>5</sub>-K<sub>2</sub>O) in early June, mid-July and early Sept. of each year. Pastures

were sprinkler irrigated for 12 h every 2 weeks during the growing season at a rate of 3.8 mm/h for a total of 46 mm of water per irrigation, equaling the available water-holding capacity of the soil. Pastures were broadcast seeded 14–15 August 2012 with CMV, BFT or MBG at 20, 34, and 37 kg pure live seed/ha, respectively. Temperature, evapotranspiration and solar radiation data were collected from an on-site Utah Climate Center meteorological weather station, and precipitation data were provided by a Utah Climate Center meteorological weather station approximately 7 km from the experimental site (Figure 2). The relatively alkaline soils, long (15 h) warm ( $>30\text{ }^{\circ}\text{C}$ ) sunny ( $350\text{ W m}^{-2}$ ) days and cool ( $60\text{ }^{\circ}\text{C}$ ) nights characteristic of the Mountain West growing season (Figure 2a,c) are favorable for the production of deep-rooted perennial legumes such as BFT, CMV and alfalfa, particularly under irrigation. Precipitation in the Mountain West is variable (Figure 2b,d) but typically occurs as rain in spring and autumn or as snowfall in winter. Evaporative demand has the opposite profile (Figure 2b,d), peaking in mid-summer, and can be addressed using irrigation except in periods of prolonged drought when water supply becomes limiting. Irrigation was unrestricted during this study.



**Figure 2.** (a) Temperature and (b) precipitation (columns) and evapotranspiration demand (lines) for 2014–2016 and (c) temperature and (d) precipitation and evapotranspiration demand for 2017 and 2018.

## 2.2. Grazing and Feeding

During each year of this study, the pasture experimental unit was a rotationally stocked pasture with its assigned animal or pair of animals (Table 1). In 2014, each pasture was

stocked with a pregnant 4- or 5-year-old Angus cow, in 2015 with a pair of yearling calves (Angus on 2 reps and Charolais on 3 reps), in 2016 with a 7- or 8-year-old dry Angus cow and in 2017 and 2018 with a 2-year-old Angus heifer. Each year, cattle were sorted into three groups of five animals (pairs in 2015) with similar body weight (BW), and one animal from each group was randomly assigned to each of the five replications of a given treatment. In 2015, pairs of Angus calves were randomly assigned to replications 3 and 4 of each treatment and pairs of Charolais calves were randomly assigned to replications 1, 2 and 5 of each treatment. Results for CH<sub>4</sub> from cows and calves in 2014 and 2015 were used to formulate models and tests that were applied to CH<sub>4</sub> data from 2017 to 2018 for 2-year-old heifers. In 2017 and 2018, a fourth treatment comprised five 2-year-old Angus heifers randomly assigned to one of five pens at the USU Animal Science Farm (latitude 41°40' N, longitude 111°53' W; 1370 m a.s.l.) and fed a TMR in confinement. Cattle were weighed before being moved to treatment pastures or confinement stalls and at the end of the study when they were removed from treatments. The yearling calves used in 2015 were previously on a diet of 50% alfalfa hay and 50% corn silage and were, therefore, not expected to gain weight during the 7 weeks they were assigned to grass and legume pastures.

**Table 1.** Details of cattle used in the grazing study along with grazing start and end dates. Values for 2017 and 2018 include the confinement treatment.

Year	Breed	Status	n Per Spatial Replication	Initial and Final Body Weight, kg	Start-End Date
2014	Angus	Pregnant cows	1	617 (±8.4) to 689 (±15.9)	9 June–22 August
2015	Angus	Yearlings	2, 2 reps	414 (±9.7) to 413 (±9.3)	6 July–21 August
2015	Charolais	Yearlings	2, 3 reps	462 (±9.3) to 450 (±9.7)	6 July–21 August
2016	Angus	Dry cows	1	654 (±19.2) to 664 (±17.4)	31 May–12 August
2017	Angus	2-year-old heifers	1	504 (±6.7) to 542 (±7.8)	22 May–17 August
2018	Angus	2-year-old heifers	1	587 (±12.9) to 614 (±13.4)	29 May–22 August

Cattle on pastures were moved to an ungrazed paddock within their pasture every 3.5 days; fresh water and trace-mineralized salt blocks (Morton iOFIXT T-M, Morton Salt Inc., Chicago, IL, USA) were provided ad libitum. In 2014, cows grazed for 5 weeks before CH<sub>4</sub> data collection began, whereas in 2015, 2017 and 2018, calves or heifers grazed for 2 weeks before CH<sub>4</sub> data collection began. In 2015, one calf from each pair was assigned to enteric CH<sub>4</sub> determinations and the other was used for DMI calculations. In 2016, dry cows grazed the pastures for 12 weeks but no CH<sub>4</sub> data were collected. Cattle were dewormed with albendazole (Valbazen broad spectrum dewormer, Pfizer Animal Health, Exton, PA, USA) at 8.8 mL/100 kg of BW before being permitted on pastures, and they were provided with permethrin ear tags (GardStar Plus, Y-Tex Corp., Cody, WY, USA) to reduce horn flies.

In 2017 and 2018, cattle in the confinement treatment were randomly assigned to individual adjacent pens measuring 5 × 10 m inside a covered barn and received a TMR consisting of 25% of alfalfa hay, 25% corn silage and 50% chopped barley (dry matter (DM) basis). The TMR was offered each day at 0900 h and 27 kg/animal was offered in both years. Refused feed was collected at 0850 h on the following day and weighed; fresh feed was offered immediately upon refusal collection. Feed intake was measured from day 5 to 9 of each collection period and the difference between amounts of feed offered and refused (on a DM basis) was recorded as feed intake. Each animal had ad libitum access to water and trace mineral salt blocks (Morton iOFIXT T-M). Methane data were collected in 2017 and 2018, but urine and fecal samples were only collected from feedlot animals in 2018.

### 2.3. Plant Production, Intake, Nutritive Value and Chemical Composition

Available herbage DM in the grazed paddock that cattle were entering, and remaining DM in the paddock that was being vacated, were non-destructively sampled using a Farmworks (Feilding, NZ) rising plate meter (RPM) calibrated for each forage species [25]. Forage DM disappearance was calculated as the difference between pre- and post-grazing DM availability [26,27]. Mean RPM values for pre- and post-grazing paddocks were



determined by walking in a “lazy W” pattern to accumulate a minimum of 30 RPM readings [25]. Calibration samples were collected in pre-grazing and post-grazing paddocks each week (2014–2016) or month (sampling period) (2017–2018) from the area under the RPM by clipping forage to a 1-cm height above the soil surface. Calibration samples were dried in a forced-air oven at 60 °C to constant weight. These samples were collected from a range of plant growth heights each year, and a linear regression of herbage dry mass on RPM readings was used to determine pasture production and forage disappearance.

To estimate intake for grazing animals during 2017 and 2018, we used fecal output estimates determined through the daily delivery of an external marker, chromic oxide (Cr<sub>2</sub>O<sub>3</sub>; see below) and forage DM digestibility estimated through near infrared spectroscopy (NIRS) analyses [28] (see below). Intake was calculated by the following equation.

$$\text{Intake on Pasture (kg/d)} = \text{Fecal output (kg/d)} / (1 - \text{Digestibility of DM (kg/kg)})$$

Intake values were then converted to units of percentage of body weight per day (% BW/d) for each legume. These percentages, in combination with the animals' individual BW, were used to estimate intake on pasture for years when fecal output estimates were not assessed (2014–2016).

Forage nutritive value samples were collected weekly (2014–2016) or monthly (2017 and 2018) from pre-grazing paddocks by clipping a small handful of stems every few steps to a height of approximately 7.5 cm along a diagonal transect of the next paddock to be grazed. Samples for determining the content of CT were taken in a similar manner but included only seeded pasture treatment species (BFT, CMV or MBG). Samples for nutritive value estimates and CT analyses were frozen under dry ice in the field, stored at –20 °C until freeze-dried and milled to pass the 1 mm screen of a Wiley mill (Thomas Scientific, Swedesboro, NJ, USA).

Forage nutritive value was determined using an NIRS analyzer (FOSS DS2500, Hillerød, Denmark) and NIRS Forage and Feed Consortium (Berea, KY, USA) forage nutritive value prediction equations. Crude protein (CP), neutral detergent fiber (aNDF), acid detergent fiber (ADF), lignin and ash of NIRS calibration samples were determined according to AOAC [29] methods 984.13, 2002.04, 973.18 and 942.05, respectively, and crude fat (CF) by AOAC [28] method 920.39. The *in vitro* true dry matter digestibility (IVTDMD) of calibration samples was determined by incubating samples in buffered rumen fluid for 48 h followed by refluxing indigestible residues in a neutral detergent solution [30,31]. Non-fiber carbohydrate concentration was calculated as suggested by NRC [32] as  $1000 - ((\text{NDF} - \text{NDFIP}) + \text{CP} + \text{CF} + \text{ash})$  where NDFIP (NDF insoluble protein) was estimated as  $\text{NDF} \times 0.93$  [33]. The distribution and boundaries of our forage sample spectra were well-represented by the population structure of spectra in the calibration set; thus, no additional wet chemistry was required.

In 2017 and 2018, representative TMR samples were collected during each sampling period, freeze dried, milled to 1 mm and analyzed by Cumberland Valley Analytical Services (Waynesboro, PA, USA) for NDF [34], ADF ([28] Method 973.18), total N ([28] Method 990.03) and NFC concentration as described above. Total digestible nutrients (TDN) were calculated from CP and fiber concentration based on equations by [35]. Samples were analyzed for CT using the butanol-hydrochloric acid-acetone-iron method of [36], with CT isolated from BFT [37] serving as the standard.

#### 2.4. Enteric Methane Emissions

Enteric CH<sub>4</sub> emissions were determined using the sulfur hexafluoride (SF<sub>6</sub>) trace gas technique [38,39]. Before grazing began, animals to be used for CH<sub>4</sub> sampling were trained to wear halters and polyvinyl chloride canisters. Canisters equipped with quick-connect couplers were clipped to halters under the chin and attached to capillary tubing running along the halter from above the nose and mouth. Canisters were constructed of schedule 40 polyvinyl chloride 10-cm-diameter, 28-cm-long with slip caps attached to both ends; canister volume was approximately 2.5 L. Canisters were fitted with Swagelok ball valves

and quick connect fittings. A brass permeation tube with a known release rate of SF<sub>6</sub> that served as an internal standard for respiration volume [39] was placed in the reticulorumen of each animal using a bolus gun. The SF<sub>6</sub> release rate of each permeation tube was determined gravimetrically during six weeks of *in vitro* incubation at 39 °C.

Enteric CH<sub>4</sub> was collected for 4 consecutive days from one (2014) or two (2015) replications of each pasture treatment each week for five weeks or for 5 consecutive days at monthly intervals (2017 and 2018). Before CH<sub>4</sub> collection, canisters were evacuated to a tension of approximately 0.250 psi using a diaphragm vacuum pump (Vacuubrand Model MZ2NT, Wertheim, Germany) and an inline digital pressure meter (Druck, Model DPI 705, Druck Ltd., Groby, Leicester, UK). Halters were fitted with a 50 cm length of 125 µm ID × 1/16" OD U160 capillary tubing (IDEX, Oak Harbor, WA, USA) that connected a filtered inlet above the mouth and nose to a quick connect fitting near the chin. Each sampled animal was fitted with a halter and evacuated canister, and the canister was connected to the capillary tubing on the halters: Its valve was opened, the time was noted, and after approximately 24 h its valve was closed, the time was noted, and canisters were disconnected from the collection system and returned to the lab. After 24 h of collection, acceptable final tensions in canisters were between 0.25 and 0.67 atm. Tensions above or below that range indicated a leak or blockage, respectively [39].

Before field collection began, canisters fitted with capillary tubing systems were placed in pastures to determine background SF<sub>6</sub>. During each collection period, control canisters were placed in grazed sections of each treatment pasture. The inlet was positioned on top of a fence post at 1.5 m height, and data were used to correct values obtained from cattle for ambient CH<sub>4</sub> [40].

### 2.5. Methane Analysis

At the end of the 24-h collection period, the tension remaining in each sample canister was recorded; canisters were pressurized to 1.1 atm with high-purity N<sub>2</sub> gas; and the exact dilution pressure was recorded. Duplicate gas subsamples were analyzed for CH<sub>4</sub> and SF<sub>6</sub> concentrations at the Lethbridge Research and Development Centre, Agriculture and Agri-Food Canada, Lethbridge, Alberta, Canada. SF<sub>6</sub> corrects CH<sub>4</sub> for respiration volume because both gases are exhaled from the rumen at once and mixed with ambient air at the same dilution rates. Therefore, the CH<sub>4</sub> emission rate was calculated as the product of the known release rate of SF<sub>6</sub> and the ratio of CH<sub>4</sub> and SF<sub>6</sub> corrected for control canister concentrations [39,40].

$$\text{CH}_4 \text{ emission rate (g d}^{-1}\text{)} = \text{SF}_6 \text{ release rate (g d}^{-1}\text{)} \times [(\text{sample CH}_4 \text{ (g d}^{-1}\text{)} - \text{control CH}_4 \text{ (g d}^{-1}\text{)}) / (\text{sample SF}_6 \text{ (g d}^{-1}\text{)} - \text{control SF}_6 \text{ (g d}^{-1}\text{)})]$$

### 2.6. Fecal Output (2017–2018)

During 9-day (Thursday–Saturday) sampling periods spaced 4 weeks apart in 2017 and 2018, 2-year-old heifers were moved each day between 0800 and 0900 h to a holding area with a squeeze chute. A gelatin capsule containing 15 g of the indigestible marker Cr<sub>2</sub>O<sub>3</sub> lubricated with mineral oil was administered to each animal using a bolus gun. Fecal grab samples of approximately 50 g DM were collected on days 5 to 9 of the sampling period and transported to the lab on dry ice where they were stored at −20 °C. At the end of each collection period, fecal samples were thawed, and daily fecal samples for each animal were composited for that period. The homogenized composited samples were frozen and lyophilized (Free Zone 18 L, Labconco Corporation, Kansas City, MO, USA). The dry sample weight was recorded, and feces were milled to pass the 1 mm screen of a Wiley mill (Thomas Scientific, Swedensboro, NJ, USA). Composite fecal samples for each animal and period were analyzed for Cr<sub>2</sub>O<sub>3</sub> concentration using the method of [41], and the concentration of fecal Cr<sub>2</sub>O<sub>3</sub> was used to calculate fecal output as described by Parker et al. [42] using the following equation.

$$\text{Fecal output (g d}^{-1}\text{)} = \text{daily Cr}_2\text{O}_3 \text{ dose (g)} \div \text{Cr}_2\text{O}_3 \text{ concentration in feces (g Cr}_2\text{O}_3 * \text{g}^{-1} \text{ feces DM)}$$

Fecal samples were also analyzed for DM concentration by drying further at 100 °C for 24 h ([28]; Method 967.03). All freeze-dried samples were analyzed for NDF, ADF and total N concentration as previously described.

### 2.7. Blood and Urine Collection and Nitrogen Balance (2017–2018)

Blood and urine samples were collected at the end of each sampling period for each animal. Blood was taken from the coccygeal vein of the tail using 10 mL vacuum tubes without additives [43]. Blood was allowed to clot at room temperature; serum was separated by centrifugation at 2000 × g for 20 min; and aliquots of serum were removed and frozen at −20 °C until analysis for blood urea N (BUN). Blood serum samples were analyzed for BUN using Siemens Urea Nitrogen Flex Reagent (Siemens Healthcare Diagnostics, Newark, DE, USA) at the Utah Veterinary Diagnostic Laboratory.

Urine was collected through vulva massage. A volume of 25 mL/cow was measured using a calibrated cylinder, and 3.125 mL of HCl 6N was added to reduce pH in order to avoid N volatilization. Samples were stored at −20 °C until N analysis [44]. Urine samples were analyzed for urinary N concentration (Leco Corporation FP-528 Protein/Nitrogen Determinator) expressed in grams per liter (g/L). Daily urinary N excretion was calculated using the following equation [45].

$$\text{Urinary N output (g d}^{-1}\text{)} = \text{Clearance rate (L blood cleared of BUN d}^{-1} \text{ kg}^{-1} \text{ BW)} * \text{BUN (g L}^{-1}\text{)} * \text{BW (kg)}.$$

The clearance rate for cattle was determined to be 1.3 L of blood cleared d<sup>−1</sup> kg BW<sup>−1</sup> [45]. Dietary N intake was calculated as follows.

$$\text{Dietary N intake (g d}^{-1}\text{)} = \text{CP content of the forage consumed (g kg}^{-1} \text{ DM)} * 0.16 \text{ (proportion of N in CP)} * \text{DMI (kg DM d}^{-1}\text{)} \text{ calculated as described above.}$$

Finally, N retained (g d<sup>−1</sup>) by the animals was calculated by subtracting the amount of N excreted in feces and urine from N intake. Total N excretion in urine and feces was estimated as a percentage of N intake (total N excretion, percentage (%) of intake); N retention by the animals was calculated as a percentage of N intake (N retention, percentage (%) of intake); and urinary and fecal N excretion as a percentage of N intake (urinary N, percentage (%) of intake and fecal N, percentage (%) of intake), respectively.

### 2.8. Statistical Analysis

Trials carried out between 2014 and 2016 employed a different class of cattle in each year; thus, data were analyzed separately for each year, while data for 2-year-old heifers collected in 2017 and 2018 were combined. Data from 2014–2016 informed the statistical models and tests for data collected in 2017 and 2018, including a confinement-fed TMR treatment. Data from 2017 and 2018 were pooled because the same class of cattle was managed under the same experimental conditions. Herbage biomass (pre-grazing, post-grazing and their difference) and the nutritive composition (CP, aNDF, ADF, fat, lignin, ash, IVTDMD, NFC and CT) of samples collected before grazing began in each paddock. BW, DMI and CH<sub>4</sub> emissions per animal per day on the basis of DMI were analyzed using a mixed model with repeated measures in which species (treatment) and week or period were fixed factors. Random factors were replication, species–replication interaction and year (for 2017 and 2018 data). The error variance-covariance matrix was unstructured and varied by year. Interaction between species and period or week was either not significant or marginally significant. Examination of the interaction revealed that the interaction was mainly caused by the magnitude of species differences; the nature of the species difference remained the same regardless of week or period. Therefore, the main effects of species



were estimated and compared. All analyses were conducted using SAS PROC GLIMMIX (SAT/STAT 15.1, SAS Institute, Cary, NC, USA). Least squares means (LSMEANS) were estimated and compared pairwise with the Tukey–Kramer method to adjust for multiplicity. Statistical significance is specified at  $\alpha = 0.05$ .

### 3. Results

In all 5 years, legumes had greater CP, IVTDMD and NFC than the grass, while grass had greater aNDF, ADF and CF than legumes (Table 2). Digestibility was greater for the legumes than for the grass; the TDN of legumes was always greater than 700 g kg<sup>-1</sup> DM, and in 2017 and 2018 it was greater than that of the high-forage confinement TMR (Table 2). The available forage DM in pastures before they were grazed was greater for CMV than for MBG in most years (Table 3). Forage utilization rarely exceeded 50% and, thus, forage intake was not limited by availability.

**Table 2.** Grazing period forage nutritive value characteristics determined using near infrared spectroscopy (NIRS).

Species <sup>2</sup>	CP	aNDF	ADF	Fat	Lignin g kg <sup>-1</sup> DM	Ash	IVTDMD	NFC	TDN	CT
2014										
BFT	271 a	188 b	188 b	24 b	62 a	85 c	942 a	432 a	778 a	22.14 a
CMV	254 b	186 b	173 c	23 c	60 a	89 b	924 b	444 a	754 b	2.84 b
MBG	178 c	492 a	311 a	40 a	35 b	126 a	862 c	185 b	572 c	3.10 b
2015										
BFT	219 b	324 b	271 b	14 b	67 a	59 c	790 b	382 b	708 b	15.12 a
CMV	258 a	244 c	220 c	14 b	59 b	76 b	912 a	415 a	773 a	2.44 b
MBG	199 c	531 a	317 a	24 a	31 c	92 a	805 b	169 c	663 c	3.32 b
2016										
BFT	201 b	325 b	251 b	16 b	60 a	52 b	803 b	406 a	735 a	16.41 a
CMV	228 a	289 c	243 b	14 c	57 b	68 a	867 a	400 a	744 a	1.91 b
MBG	139 c	583 a	354 a	25 a	39 c	70 a	749 c	194 b	619 b	1.73 b
2017										
BFT	206 b	333 b	237 b	20 b	49 ab	62 ab	863 b	387 ab	745 a	19.29 a
CMV	253 a	250 c	211 c	18 c	45 b	78 a	915 b	408 a	775 a	1.89 b
MBG	129 c	568 a	348 a	27 a	51 a	40 b	766 c	232 b	618 b	1.25 b
Confinement <sup>1</sup>	147	329	206	24	39	85		435	724	0.71
2018										
BFT	200 a	310 b	233 b	23 b	57 a	60 b	827 b	401 a	754 a	12.94 a
CMV	226 a	261 c	225 b	15 c	64 a	73 a	870 a	429 a	765 a	0.85 b
MBG	73 b	639 a	406 a	37 a	49 b	66 ab	713 c	201 b	557 b	-0.62 b <sup>2</sup>
Confinement <sup>1</sup>	121	359	248	24	41	85		431	680	0.68
Means										
BFT	219	296	236	19	59	64	845	402	744	17.18
CMV	244	246	214	17	57	77	898	419	762	1.99
MBG	144	563	347	31	41	79	779	196	606	1.76
Confinement <sup>1</sup>	134	344	227	24	40	85		433	702	0.70

Values in columns within years with different letters are significantly different at  $p < 0.05$ . BFT, birdfoot trefoil; CMV, cicer milkvetch; MBG, meadow bromegrass; CP, crude protein; aNDF, neutral detergent fiber determined with the addition of amylase; ADF, acid detergent fiber; IVTDMD, in vitro true dry matter digestibility; NFC, non-fiber carbohydrates; TDN, total digestible nutrients; CT, condensed tannins. <sup>1</sup> Confinement TMR samples are the mean of unreplicated samples for 2 (2017) or 3 (2018) periods. <sup>2</sup> A negative value resulted from absorbance less than that of the assay blank.

**Table 3.** Mean pre-grazing and post-grazing dry matter (DM), their difference, forage disappearance (SEM) and resulting pasture utilization.

Species	Pre-Grazing	Post-Grazing kg DM/ha	Forage Disappearance	Utilization %
2014				
BFT	4089 (178) a	2479 (132) a	1113 (124) b	27
CMV	3781 (165) a	2031 (105) b	1528 (120) a	40
MBG	2741 (119) b	2038 (148) b	1485 (174) a	54
2015				
BFT	4687 (132) b	2719 (98) a	1998 (139) b	43
CMV	5345 (132) a	2652 (98) a	2733 (139) a	51
MBG	3060 (132) c	1396 (98) b	1664 (136) b	54
2016				
BFT	5641 (168) b	3057 (77) b	2584 (140) a	46
CMV	6398 (168) a	3713 (77) a	2685 (140) a	42
MBG	4862 (168) c	2805 (77) b	2047 (140) b	42
2017 and 2018				
Period 1				
BFT	4568 (684) c	2547 (402) b	2365 (412) b	52
CMV	7810 (1170) a	4242 (670) a	3886 (412) a	50
MBG	6232 (933) b	4653 (735) a	1705 (412) b	27
Period 2				
BFT	4030 (603) b	2821 (446) b	1035 (412) b	26
CMV	6316 (946) a	3263 (516) b	3012 (412) a	48
MBG	6126 (917) a	4365 (690) a	1861 (412) ab	30
Period 3				
BFT	4665 (698) a	3022 (478) a	1698 (412) a	36
CMV	5200 (779) a	3191 (504) a	2004 (412) a	39
MBG	4747 (711) a	3558 (562) a	1302 (412) a	27

Values in columns within years with different letters are significantly different at  $p < 0.05$ . BFT, birdfoot trefoil; CMV, cicer milkvetch; MBG, meadow bromegrass.

The DMI ( $\text{kg d}^{-1}$ ) of cattle on BFT and CMV pastures was greater than for cattle on MBG pasture in all 5 years (Figure 3a–d). The difference in the BW of cows in 2014 and 2016, yearling calves in 2015 and 2-year-old heifers in 2017 and 2018 is reflected in the magnitude of DMI in these years. Enteric  $\text{CH}_4$  emissions per unit of DMI were less for cattle grazing perennial legume pastures than for cattle on grass pastures in every year of the study (Figure 3e–g). In 2017 and 2018,  $\text{CH}_4$  emissions by heifers fed with TMR was intermediate to cattle on legume and grass pastures (Figure 3g).

The N balance calculated for 2017–2018 data (Table 4) demonstrated that greater DMI and CP concentration of legumes resulted in greater dietary N intake and greater urinary N output for cattle grazing legume pastures than for cattle grazing MBG or offered the TMR (Table 4). The N retention of legume-fed cattle was also much greater for cattle grazing legumes than for cattle on grass or fed with TMR, with the opposite pattern observed for the proportion of N excreted relative to N intake (N excretion, percentage (%) intake; Table 4). The urinary output was greater for legumes than for other treatments due to the greater concentration and intake of dietary N, and urinary N excretion as a proportion of N ingested (urinary N, % intake) was greater for CMV and BFT. Finally, fecal N as a proportion of N intake was the lowest for legumes, with the opposite pattern for the urinary:fecal N ratio (Table 4).



**Figure 3.** (a–d) Least squares means (LSMEANS) of daily dry matter intake (DMI) based on forage quality and body weight and (e–g) methane (CH<sub>4</sub>) emissions as a function of DMI for (a,e) pregnant cows (653 kg) in 2014, (b,f) yearling calves (439 kg) in 2015, (c) dry cows (659 kg) in 2016 and (d,g) 2-year-old heifers (562 kg) in 2017–2018 on the three pasture treatments (BFT, birdfoot trefoil; CMV, cicer milkvetch; MBG, meadow brome) and on a confinement total mixed ration (TMR) in 2017 and 2018. Data for DMI and CH<sub>4</sub> were analyzed separately for 2014 and 2015 and combined for 2017 and 2018; no CH<sub>4</sub> data were collected in 2016. Error bars are standard errors of the estimate. LSMEANS for each treatment within each year with the same letter are not different at  $\alpha = 0.05$  (a–c). Error bars represent  $\pm$  SEM.

**Table 4.** Intake, retention and output of nitrogen (N) in urine and feces in 2017 and 2018.

	BFT	CMV	MBG	Confinement
Dietary N intake, g/d	425.57 a	449.40 a	162.91 b	201.60 b
Urinary N output, g/d	133.75 a	131.38 a	54.07 b	66.77 b
Fecal N output, g/d	97.72 a	70.49 b	68.15 b	101.73 a
Total N excretion, g/d	221.89 a	200.66 a	128.73 b	157.93 ab
N retention, g/d	202.99 a	263.57 a	44.30 b	42.72 a
Urinary N, % of intake	34.15 ab	31.87 b	42.75 a	22.12 c
Fecal N, % of intake	25.81 b	17.05 c	47.98 a	41.94 a
Total N excretion, % of intake	55.85 bc	45.87 c	86.82 a	64.10 b
N retention, % of intake	42.58 a	53.65 a	27.31 b	39.51 ab
Urinary: Fecal N	1.21 b	1.70 a	0.71 c	0.53 c
Blood urea N, mg/dL	18.24 a	19.22 a	7.68 b	8.96 b
Urinary N, g/L	5.5 a	4.8 a	2.7 b	4.0 ab

Least squares means (LSMEANS) for each treatment within each year with the same letter are not different at  $\alpha = 0.05$ .

#### 4. Discussion

The Mountain West climate has a unique influence on forage quality. Low annual precipitation and high evapotranspiration have resulted in moderately alkaline soils that are naturally high in calcium, phosphorus and potassium. The growing-season climate regimen includes high daytime temperatures, high solar radiation and low nighttime temperatures, maximizing photosynthesis and minimizing aerobic respiration. We have recently determined that perennial legumes (e.g., alfalfa, BFT, sainfoin and CMV) not only

persist and yield well in this environment, but they accumulate levels of NFC comparable to beet pulp or corn silage [24,46]. For comparison with forage yields from other locations, annual yields of four replications of 0.8-ha monoculture stands of 'NexGrow 6409 HVRX' alfalfa and 'Cache' MBG planted in fall of 2019 and harvested in early June, mid-July and late August of 2020 and 2021 averaged 12,296 and 10,475 kg ha<sup>-1</sup>, respectively (MacAdam, pers. comm.).

This study sought to demonstrate the value of perennial legumes for reducing environmental impacts of Mountain West beef cattle production. We investigated enteric CH<sub>4</sub> emissions and the retention of N in cows, calves and heifers because beef cows produce the greatest proportion of the emissions from the cowherd [15], and we sought to determine the potential of perennial legumes to reduce beef system GHG by contrasting cattle grazing legume monocultures with cattle grazing grass monocultures or those fed with TMR in confinement.

#### 4.1. Nutritional Quality and Intake

The nutritional quality of the legumes explains the high values of DM disappearance observed for these treatments. For data reported in Table 2, the 5-year mean of aNDF concentration of the two legumes was 271 g kg<sup>-1</sup> DM while the mean aNDF concentrations of the grass and TMR diet were 563 and 344 g kg<sup>-1</sup> DM, respectively. The 5-year mean NFC concentration of the two legumes was 410 g kg<sup>-1</sup> DM and their mean TDN concentration was 752 g kg<sup>-1</sup> DM, whereas NFC mean concentrations for the grass and TMR diet were 196 and 433, respectively, and their mean TDN concentrations were 606 and 702 g kg<sup>-1</sup> DM. The CP concentrations of the legumes were much greater than of grass or TMR. Legumes with lower cell wall (aNDF) content allow for increased passage rates [47], and greater CP and NFC concentrations enhanced fermentation rates and intake in ruminants [8].

The DMI of animals grazing BFT and CMV was always greater than the DMI of animals grazing MBG (Figure 3a–d), and in 2017 and 2018, the DMI of heifers on BFT and CMV pastures did not differ from the intake of TMR-fed confinement heifers. There was some deterioration of the BFT pastures over time, with the presence of weeds revealed by increments in aNDF and ADF concentrations relative to CMV (Table 2).

#### 4.2. Methane Emissions

Enteric CH<sub>4</sub> emission per unit of DMI was less for cattle grazing perennial legume pastures than for cattle on grass and did not differ from cattle fed with TMR. Diet quality affects the amount of CH<sub>4</sub> emitted by ruminants. Forages with higher fiber concentrations (i.e., MBG in this study) constrain passage rate and increase ruminal retention time [47,48], which in turn results in an increment of CH<sub>4</sub> production per unit of forage intake (CH<sub>4</sub> yield), because as fiber increases the extent of rumen fermentation increases and there is more hydrogen available as a substrate for methanogenic archaea [49]. In addition, a more fibrous diet usually results in greater acetic acid production, which increases CH<sub>4</sub> production [50,51].

On the other hand, forages with lower fiber concentration have greater passage rates and favor propionate production, which is considered a competitive pathway for hydrogen use in the rumen [49]. In addition, legumes with high NFC concentrations that are rapidly fermented in the rumen and a low proportion of structural carbohydrates can yield proportions of ruminal microorganisms similar to those present in grain-fed animals, increasing proportions of propionate-forming bacteria, decreasing H<sub>2</sub> production and resulting in decreased CH<sub>4</sub> emissions relative to forages with lower concentrations of NFC, such as grasses [52]. Finally, it is known that CT, such as those present in BFT, may inhibit CH<sub>4</sub> production in the rumen either in absolute amounts (g/d) or as CH<sub>4</sub> yield (g/kg DMI). The CT concentration of BFT was borderline (12 to 20 g/kg DM) during the 5 years of this study compared with the effective BFT range (20 to 50 g/kg DM) known to decrease CH<sub>4</sub> emissions [53–55], and CH<sub>4</sub> emissions for cattle on BFT pastures were no less

than that of cattle on CMV pastures. Thus, it is likely that high NFC concentrations in BFT were more consequential than CT for reducing CH<sub>4</sub> emissions in this study.

#### 4.3. Nitrogen Retention and Emissions

Nitrogen lost to the environment as nitrous oxide (N<sub>2</sub>O) has far greater warming potential per weight of gas than either CO<sub>2</sub> or CH<sub>4</sub>, and ammonia can contribute to particulate (PM<sub>2.5</sub>) air pollution. The CP concentration of the two pasture legumes was greater than for the TMR which was greater than for MBG. Along with the greater DMI of animals consuming legumes, this led to greater N intake and greater urinary N excretion but reduced total N excretion as a proportion of intake, compared with the grass. The N retention by cattle grazing legumes was greater than for the grass and the same as for TMR diets, suggesting that N consumed from legume pastures will contribute more to meat, milk and wool production than N consumed from grass pastures (i.e., greater N efficiencies). Nevertheless, it is worth mentioning that N balance studies are prone to measurement errors (e.g., N volatilization and fecal N excretion) and, in particular, under grazing studies with a high degree of spatial and animal variation [56,57] that adds uncertainty to the estimates.

The high N retention values observed in this study are consistent with previous results reported for legume-based diets, e.g., [21,58,59]. Legumes present high concentrations of NFC, and the provision of readily available sources of energy such as NFC to the rumen may enhance the efficiency of N use as these carbohydrates provide substrate, along with ammonia, for the production of microbial protein. In addition, increased N retention values in legume diets have been attributed to augmented levels of highly digestible CP intake consumed with synchronous sources of carbohydrates [58]. Finally, the CT in BFT may have contributed to enhanced fecal N excretion relative to CMV, resulting in a greater urinary:fecal N ratio in cattle grazing CMV.

## 5. Conclusions

The legume pastures grazed in this study resulted in reductions in CH<sub>4</sub> emissions as a function of intake of between 25 and 63% relative to grass pasture and reductions in excretion as a proportion of dietary N between 36 and 47% compared with animals on the grass pasture. The elevated concentrations of both NFC and CP and reduced concentrations of aNDF in legume pastures compared with the grass pasture resulted in intake and methane emissions comparable to TMR confinement-fed cattle in 2017–2018 and in N retention greater than grass-fed cattle and not different from cattle fed with TMR containing 50% grain, thereby enhancing the sustainability of beef agroecosystems. Condensed tannins produced by BFT and other temperate legumes have the potential to further enhance the efficiency of energy and protein use in ruminants relative to other perennial species. Life cycle assessment will be used to determine the implications of these results for beef system GHG emissions.

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**Data Availability Statement:** The data generated during the study are available upon request from the corresponding author.



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### Abbreviations

ADF	Acid detergent fiber
BFT	Birdsfoot trefoil
BUN	Blood urea nitrogen
BW	Body weight
CO <sub>2</sub>	Carbon dioxide
Cr <sub>2</sub> O <sub>3</sub>	Chromic oxide
CMV	Cicer milkvetch
CT	Condensed tannins
CF	Crude fat
CP	Crude protein
DM	Dry matter
DMI	Dry matter intake
GHG	Greenhouse gas
IVTDMD	In vitro true dry matter digestibility
LSMEANS	Least squares means
MBG	Meadow brome grass
CH <sub>4</sub>	Methane
NDFIP	NDF insoluble protein
NIRS	Near infrared spectroscopy
aNDF	Neutral detergent fiber
N	Nitrogen
N <sub>2</sub> O	Nitrous oxide
NFC	Non-fiber carbohydrates
P <sub>2</sub> O <sub>5</sub>	Phosphorus pentoxide
K <sub>2</sub> O	Potassium oxide
RPM	Rising plate meter
SF <sub>6</sub>	Sulfur hexafluoride
TMR	Total mixed ration
USU	Utah State University

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