



BRIEF REPORT

Carbon-substrate utilization profiles by *Cladorrhinum* (Ascomycota)

Viviana A. Barrera^{a,*}, Mara E. Martin^{a,b,*}, Mónica Aulicino^c, Sofía Martínez^a, Guido Chiessa^a, Mario C.N. Saparrat^{b,d,e,*}, Amelia L. Gasoni^a

^a Instituto de Microbiología y Zoología Agrícola, Instituto Nacional de Tecnología Agropecuaria, CC 25 (1712) Castelar, Buenos Aires, Argentina

^b Consejo Nacional de Investigaciones Científicas y Técnicas, Argentina

^c Facultad de Ciencias Agrarias, UNLZ, Camino de Cintura y Juan XXIII, Lomas de Zamora, Argentina

^d Instituto de Fisiología Vegetal (INFIVE), UNLP, CCT-La Plata-CONICET, Diag. 113 y 61, CC 327, 1900 La Plata, Argentina

^e Instituto de Botánica Spegazzini, Facultad de Ciencias Naturales y Museo, UNLP, 53 # 477, 1900, La Plata, Argentina

Received 20 April 2018; accepted 24 September 2018

KEYWORDS

Cladorrhinum;
Metabolic profile;
Biolog[®] FF system

Abstract Fungi from the genus *Cladorrhinum* (Ascomycota) are promising agents in the bio-control of phytopathogens, in the promotion of plant growth, and in the production of enzymes with technological application. We analyzed comparatively the ability of 5 native strains of *Cladorrhinum samala* and *Cladorrhinum bulbiliosum* with reference strains belonging to the same genus. We used 95 individual carbon sources available in microplates from the Biolog[®] FF system. Although most of the strains mainly used soluble carbohydrates, the metabolic profile was highly dependent upon each isolate and it revealed intraspecific physiological variability in *Cladorrhinum* species.

© 2018 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

PALABRAS CLAVE

Cladorrhinum;
Perfiles metabólicos;
Biolog[®] FF system

Perfiles de utilización de sustratos carbonados de *Cladorrhinum* (Ascomycota)

Resumen Los hongos del género *Cladorrhinum* (Ascomycota) son agentes prometedores en el biocontrol de fitopatógenos, la promoción del crecimiento de las plantas y la producción de enzimas con aplicación tecnológica. En este trabajo se analizaron comparativamente las habilidades de 5 cepas nativas pertenecientes a las especies *Cladorrhinum samala* y *Cladorrhinum bulbiliosum* con cepas de referencia del mismo género. Se usaron 95 fuentes individuales de

* Corresponding authors.

E-mail addresses: barrera.viviana@inta.gob.ar (V.A. Barrera), martin.mara@inta.gob.ar (M.E. Martin), masaparrat@yahoo.com.ar (M.C.N. Saparrat).

<https://doi.org/10.1016/j.ram.2018.09.005>

0325-7541/© 2018 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

carbón, disponibles en microplacas de Biolog® FF system. Aunque la mayoría de las cepas utilizaron principalmente carbohidratos solubles, el perfil metabólico fue altamente dependiente de cada aislamiento y reveló variabilidad fisiológica intraespecífica en las especies de *Cladorrhinum*.

© 2018 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

The genus *Cladorrhinum* Sacc. and Marchal (Lasiosphaeriaceae, Sordariomycetes, Ascomycota [IndexFungorum; <http://www.indexfungorum.org/names/Names.asp>]) includes a fungal group of fundamental importance for agriculture and livestock, because some species have potential as agents in the biocontrol of fungal phytopathogens, in the promotion of plant growth, and in the production of phytases (U.S. Patent No. 6,514,495 from strain *Cladorrhinum foecundissimum* CBS 427.97)¹⁶. This genus includes representatives with a diagnostic conidial system that can be found in roots as endophytes or as saprotrophic forms on dung, soil or plant material, and is considered an ammonia fungus¹⁷. However, some species have also been associated with human and animal opportunistic diseases⁶.

Today the use of microbial-based fertilizers has gained significance in the effort to reduce the negative environmental effects generated by the excessive and/or improper application of chemical fertilizers. Although some *Cladorrhinum* strains have been proposed as promising agents in the development of biofertilizers for plant production, the knowledge of the nutritional features of these fungi, which are important in the industrial manufacturing of new biofertilizers using them, is scarce⁷. Carmarán et al.³ reported data about the growth of three strains in a standard agar medium under a narrow range of temperature. However, analysis of nutritional preferences based on carbon substrate utilization profiles can be used to identify and characterize phenotypical diversification in *Cladorrhinum* strains and to characterize the Biolog FF MicroPlates carbon compounds for fungal growth.

The aim of this work was to characterize 10 strains from the genus *Cladorrhinum* through carbon-substrate utilization profiles by the Biolog® system (Biolog Inc., Hayward, CA) and evaluate the physiological behavior of the strains related to the taxonomic delimitation of the species of the genus by comparison with the type strains.

In this study we used 5 reference strains from *Cladorrhinum samala*, *Cladorrhinum bulbillosum* and *Cladorrhinum foecundissimum* and 5 native strains corresponding to *Cladorrhinum samala* and *Cladorrhinum bulbillosum* deposited in the fungal collection at the Instituto de Microbiología y Zoología Agrícola, Instituto Nacional de Tecnología Agropecuaria (INTA), Argentina. The fungi were preserved at -20°C in tubes containing media developed by Butler¹⁸ and at 4°C in glycerol media. Table 1 shows the strains of *Cladorrhinum* spp. included in this study.

Carbon assimilation was investigated using Biolog FF MicroPlates. These plates are especially developed for

cultivating filamentous fungi through the 95 individual carbon source utilization analysis (Biolog Inc. USA). The FF-IF broth (filamentous fungi-inoculation fluid) was prepared in a borosilicate test tube by mixing 0.25% Phytigel (P8169, Sigma) and 0.03% Tween 40 (P1504, Sigma) in distilled water. The solution was stirred until all the components were dissolved and sterilized by autoclaving for 20 min at 121°C . Biolog FF MicroPlates (cat. no. 1006) were stored at 4°C until use. Pure cultures from the frozen stocks of *Cladorrhinum* spp. were firstly subcultured onto Potato Dextrose Agar (PDA) and then onto Malt Extract Agar (MEA) at 25°C . To promote sporulation, strains of *Cladorrhinum* spp. were incubated for 20 days under UV light with 12-hour photoperiod. Conidia were collected with sterile cotton-tipped swabs and suspended in a 16 ml tube containing sterile IF-FF broth. The suspension was agitated in a vortex mixer for about 5 s and the inoculum density was adjusted to 75% transmittance at 590 nm wavelength. Three Biolog FF MicroPlates, which contain 95 individual carbon sources, were inoculated with the conidial suspension of each isolate and incubated at 25°C in the dark. After 96 h incubation, absorbance readings were taken at 750 nm, which corresponds to turbidity reflecting mycelial production¹⁰. It was done in a microplate reader Emax™ (Molecular Devices®, Inc., Sunnyvale, CA, USA).

Statistical analyses were performed using InfoStat Software⁴. Absorbance values in each well of Biolog FF MicroPlates after 96 h incubation were used instead of binary data to perform statistical analyses¹⁵. The optical density (OD) values of Biolog FF MicroPlates wells were corrected considering the background color developed in control well A1. Negative scores were set to zero. The average well color development (AWCD) was obtained as the sum of absorbance units of all positive wells divided by their total number. The average plate value was calculated using the media in triplicate. In order to reduce the variable-to-sample ratio in the microplates, the 95 carbon individual sources were grouped into eight chemical groups (carbohydrates, carboxylic acids, esters, polymers, alcohols, chemical phosphorylated, amines/amides, and amino acids). The average absorbance for the wells corresponding to each group was calculated².

An analysis of variance of a factor and contrast ($p < 0.05$) using the least significant difference (LSD) was applied to demonstrate whether the AWCD of fungal strains was differential. Ten *Cladorrhinum* spp. strains were characterized using Biolog FF MicroPlates to obtain data

Table 1 *Cladorrhinum* spp. strains used in this study.

Species	Strain code	Isolation source
<i>C. bulbillosum</i>	INTA-AR 54	Soybean crop; Buenos Aires province, Argentina (S 34° 36' W 58° 40')
<i>C. bulbillosum</i>	INTA-AR 104	Fallow land; Buenos Aires province, Argentina (S 34° 36' W 58° 40')
<i>C. bulbillosum</i>	CBS 304.90	Sand; Western Desert, Oasis Dakhla, Egypt; reference culture from holotypus
<i>C. foecundissimum</i>	CBS 180.66	Soil; Netherlands; reference culture from neotypus
<i>C. foecundissimum</i>	MUCL 6980	<i>Triticum sativum</i> soil; Schleswig-Holstein, Kiel, Kitzeberg, Germany
<i>C. foecundissimum</i>	CBS 341.92	Maryland, Beltsville, USA
<i>C. samala</i>	INTA-AR 156	Soybean crop; Santa Fe province, Argentina (S 31° 36' W 60° 47')
<i>C. samala</i>	INTA-AR 1	Alfalfa crop; Buenos Aires province, Argentina (S 34° 36' W 58° 40')
<i>C. samala</i>	INTA-AR 20	Alfalfa crop; Buenos Aires province, Argentina (S 34° 36' W 58° 40')
<i>C. samala</i>	CBS 302.90	<i>Triticum sativum</i> soil; Western Desert, Oasis Dakhla, Egypt; reference culture from neotypus

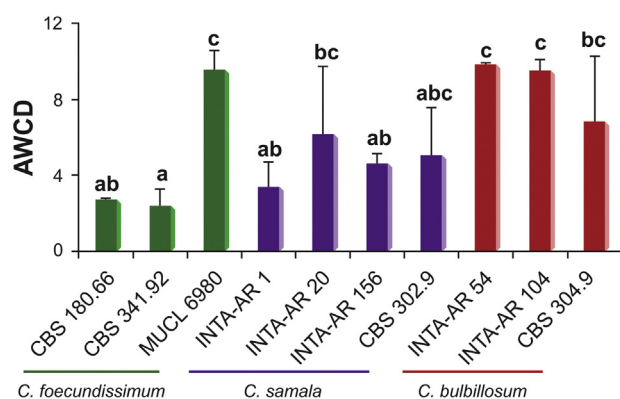


Figure 1 AWCD (Average Well Color Development) at 750 nm by *Cladorrhinum* spp. strains. Bars with the same letter are not significantly different at 5% (LSD). Different colors correspond to different species.

on C-substrate utilization. The results obtained from the analysis of variance indicated F2.24: 14.48 ($p < 0.0001$).

Figure 1 shows mycelial production (estimated by measuring average well color development [AWCD] at 750 nm) by several *Cladorrhinum* spp. strains. While the strains *C. foecundissimum* MUCL 6980, *C. samala* INTA-AR 20, *C. bulbillosum* INTA-AR 54, INTA-AR 104 and CBS 304.90 revealed the highest biomass levels, the lowest biomass production was measured for isolate *C. foecundissimum* CBS 341.92. Based on the intraspecific responses, *C. foecundissimum* strains showed more variability than the *C. bulbillosum* and *C. samala* strains tested.

Relative consumption of several C compounds by *Cladorrhinum* spp. strains is reported in **Figure 2**. Carbohydrates were mainly consumed (over 45%) by most strains, with the exception of *C. foecundissimum* CBS 180.66, *C. samala* INTA-AR 1, and *C. bulbillosum* INTA-AR 54, which used mainly esters or polymers.

The comparison of the relative use of carbon sources by 10 strains belonging to three *Cladorrhinum* species using Biolog FF MicroPlates revealed variability among these abilities. Moreover, differences were found when several strains from the same species were compared. Although similar working strategies were reported as screening and evaluation tools for the physiological characterization of

bacterial and fungal strains⁵, no data are available about the use of the microplates method for studying the biology of *Cladorrhinum* species. This Biolog FF MicroPlates analysis proved that all the strains tested might be considered different individuals due to specific biomass levels.

The results indicate that there was no species-specific behavior associated with the group of C-source assimilation in all the strains. Even though the preferential utilization of carbohydrates might be explained by the fact that carbohydrates and carboxylic acids are the primary sources for cellular metabolism⁸, other carbon sources such as amino acids also contributed to growth in the strains. The total consumption of these three compounds was 75% for most strains. The strains could be divided into three groups which were associated with: (a) intermediate to low mycelial production (*C. samala* INTA-AR 1 and INTA-AR 156, *C. foecundissimum* CBS 180.66 and CBS 341.92); (b) higher production of biomass, such as that found in *C. bulbillosum* (INTA-AR 54, INTA-AR 104, and CBS 304.90) and *C. foecundissimum* MUCL 6980; and (c) highly variable production, such as that found for some strains of *C. samala* (INTA-AR 20 and CBS 302.90). The *C. bulbillosum* INTA-AR 54 strain presented nutritional preferences for polymers, and *C. samala* INTA-AR 1 was differentiated by ester consumption in the group with low biomass production. In a taxonomic study analyzing the growth response by temperature, Madrid et al.¹¹ reported a lower growth for the *C. foecundissimum* CBS 180.66 strain than for *C. samala* CBS 302.90 and *C. bulbillosum* CBS 304.90. Carmarán et al.³ observed the same trend for the strains analyzed in the present study. The existence of intraspecific variability in *C. foecundissimum* and *C. samala* is remarkable. It is known that microorganisms including fungi use certain C-substrates to increase biomass and for housekeeping reactions needed for fungal survival⁹. The differences found between the strains studied could be explained, in part, by the balance between the metabolism for growth and for fungal survival. Since several strains of *C. foecundissimum* and *C. samala* have potential as biocontrol agents against important fungal phytopathogens⁷, the ability of specific isolates to assimilate certain C sources might be related to their competitiveness under specific ranges of nutritional conditions. Variability in carbon source utilization may be associated with different ecological behaviors.

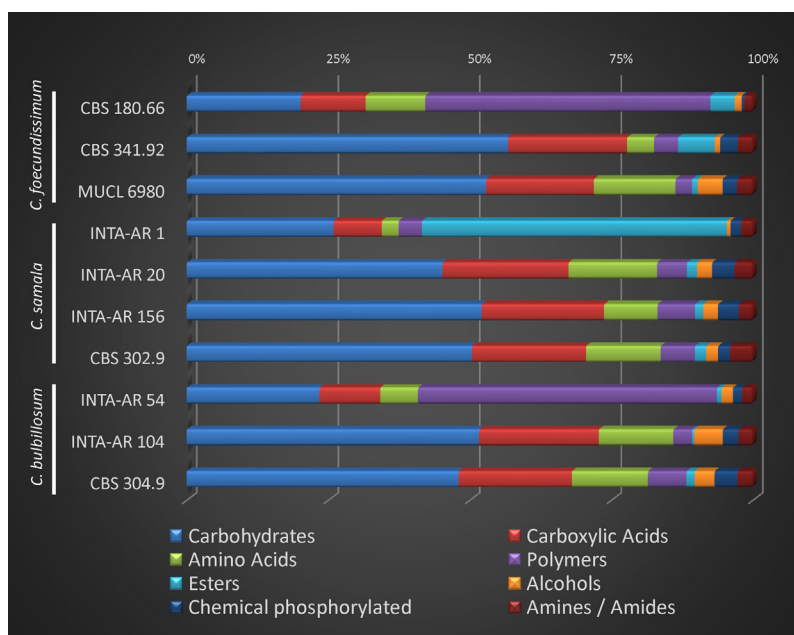


Figure 2 Relative consumption of carbon sources grouped into eight chemical carbon classes (in percentage) used by *Cladorrhinum* spp. strains measured at 750 nm.

Likewise, the functions of organisms in an ecosystem are influenced by the environment, and the particular traits of these organisms are their nutrition mode, host or substrate preference, and specificity. Rice and Currah¹³ reported that the differences observed between strains within a species reflect ecological differentiation or adaptation to different habitats. This behaviour suggests that the colonization of roots in different crops by certain *Cladorrhinum* spp. isolates might be related to an adaptative specialization. According to Sagara¹⁴, *C. foecundissimum* is a representative component of the ecophysiological group "ammonia fungi". The ability of these fungi to use amino acids as C-source could be indicative of their possible role in the ammonification processes at the rhizosphere level. The liberation of ammonium by *Cladorrhinum* spp. strains could be relevant since it could represent an additional role of these fungi in the promotion of plant growth. The use of different compounds containing low-molecular-mass nitrogen by these fungi, could play a role in the interaction of *Cladorrhinum* strains and roots and their effect on the plant promoting growth.

In agreement with Kubicek et al.¹⁰, who worked with *Trichoderma harzianum* strains, our physiological data did not reflect the taxonomical delimitation of *Cladorrhinum* spp. species. A similar situation was observed when morphological and physiological features were used to separate strains of *Trichoderma* spp. selected for biological control activity against phytopathogens^{1,12}.

To conclude, the physiological behavior of the studied *Cladorrhinum* spp. strains did not correspond to the taxonomic delimitation of the species. Further research is needed to correlate the high intraspecific variability found in the requirements of carbon sources related to the ecological behavior of the strains.

Conflict of interest

The authors declare that they have no conflicts of interest.

Financial support

INTA PROJECTS PNPV-1135023, PNAIyAV-1130034, PICT 2015-1620.

Acknowledgments

This work was funded by the Instituto Nacional de Tecnología Agropecuaria (INTA) through the following projects: PNPV-1135023 and PNAIyAV-1130034 and by the Agencia Nacional de Promoción Científica y Tecnológica (ANPCyT) of the Ministerio de Ciencia, Tecnología e Innovación Productiva through the project PICT 2015-1620 (M. C. N. Saparrat), CONICET (PUE INFIVE), CICPBA and UNLP, Argentina. Martin, M. is a recipient of a scholarship from CONICET, Argentina. Saparrat, M. C. N. is a researcher from CONICET, Argentina.

References

1. Barrera VA. El género *Hypocrea* Fr. (Hypocreales, Ascomycota) en la Argentina. Estudio de la variabilidad molecular de su estado anamórfico *Trichoderma*. Tesis doctoral en Ciencias Biológicas. 2012. Facultad de Ciencias Exactas y Naturales (UBA). http://digital.bl.fcen.uba.ar/Download/Tesis/Tesis_5226.Barrera.pdf.
2. Buyer S, Roberts DP, Millner P, Russek-Cohen E. Analysis of fungal communities by sole carbon source utilization profiles. *J Microbiol Methods*. 2001;45:53–60.
3. Carmarán CC, Berretta M, Martínez S, Barrera V, Munaut F, Gasoni L. Species diversity of *Cladorrhinum* in Argentina

- and description of a new species, *Cladorrhinum australe*. Mycol Prog. 2015;14:94, <http://dx.doi.org/10.1007/s11557-015-1106-3> [on-line].
- DiRienzo JA, Casanoves F, Balzarini MG, González L, Tablada M, Robledo CW. InfoStat versión 2017. Grupo InfoStat, FCA, Universidad Nacional de Córdoba, Argentina. <http://www.infostat.com.ar> [on-line].
 - Frąc M, Gryta A, Oszust K, Kotowicz N. Fast and accurate microplate method (Biolog MT2) for detection of *Fusarium* fungicides resistance/sensitivity. Front Microbiol. 2016;7:489, <http://dx.doi.org/10.3389/fmicb.2016.00489> [on-line].
 - Gajjar DU, Pal AK, Santos JM, Ghodadra BK, Vasavada AR. Severe pigmented keratitis caused by *Cladorrhinum bulbiliosum*. Indian J Med Microbiol. 2011;29:434–7. <http://www.ijmm.org/text.asp?2011/29/4/434/90191> [on-line].
 - Gasoni L, Stegman de Gurfinkel B. Biocontrol of *Rhizoctonia solani* by the endophytic fungus *Cladorrhinum foecundissimum* in cotton plants. Australas Plant Pathol. 2009;38:389–91.
 - Hothersall JS, Ahmed A. Metabolic fate of the increased yeast amino acid uptake subsequent to catabolite derepression. J Amino Acids. 2013;2013:461901, <http://dx.doi.org/10.1155/2013/461901> [on-line].
 - Kempes CP, Dutkiewicz S, Follows MJ. Growth, metabolic partitioning, and the size of microorganisms. Proc Natl Acad Sci USA. 2012;109:495–500.
 - Kubicek CP, Bissett J, Druzhinina I, Kullnig-Gradinger C, Szakacs G. Genetic and metabolic diversity of *Trichoderma*: a case study on South-East Asian isolates. Fungal Genet Biol. 2003;38:310–9.
 - Madrid H, Cano J, Gené J, Guarro J. Two new species of *Cladorrhinum*. Mycologia. 2011;103:795–805.
 - Ortiz EM, Duchicela J, Debut A. Scanning electron microscopic observations of early stages of interaction of *Trichoderma harzianum*, *Gliocladium virens* and *Bacillus subtilis* with *Acaulospora colombiana*. Rev Argent Microbiol. 2018;50:227–9.
 - Rice AV, Currah RS. Profiles from Biolog FF plates and morphological characteristics support the recognition of *Oidiodendron fimicola* sp. nov. Stud Mycol. 2005;53:75–82.
 - Sagara N. Ammonia fungi: a chemoecological grouping of terrestrial fungi. Contrib Biol Lab Kyoto Univ. 1975;24:205–76.
 - Singh MP. Application of Biolog FF MicroPlate for substrate utilization and metabolite profiling of closely related fungi. J Microbiol Methods. 2009;77:102–8.
 - Svendsen A, Lassen SF, Kostrewa D, Pasamontes L, Lehmann M, Tomschy A, Van Loon A, Vogel K, Wyss M. U.S. Patent No. 6,514,495. 2003. Washington, DC: U.S. Patent and Trademark Office.
 - Suzuki A. Propagation strategy of ammonia fungi. Mycoscience. 2009;50:39–51.
 - Webb KM, Hill AL, Laufman J, Hanson LE, Panella L. Long-term preservation of a collection of *Rhizoctonia solani* using cryogenic storage. Ann Appl Biol. 2011;158:297–304.